

(19) World Intellectual Property Organization
International Bureau(43) International Publication Date
20 December 2001 (20.12.2001)

PCT

(10) International Publication Number
WO 01/95702 A1(51) International Patent Classification⁷: **A01H 5/00**,
C12N 9/00, 9/02, 9/04, 9/10, 15/29

(21) International Application Number: PCT/AU01/00699

(22) International Filing Date: 14 June 2001 (14.06.2001)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
PQ 8154 14 June 2000 (14.06.2000) AU(71) Applicants (for all designated States except US): **STATE OF VICTORIA AS REPRESENTED BY DEPARTMENT OF NATURAL RESOURCES AND ENVIRONMENT [AU/AU]**; 15th Floor, 8 Nicholson Street, East Melbourne, Victoria 3002 (AU). **THE UNIVERSITY OF ADELAIDE [AU/AU]**; North Terrace, Adelaide, South Australia 5005 (AU). **INTERNATIONAL MAIZE**

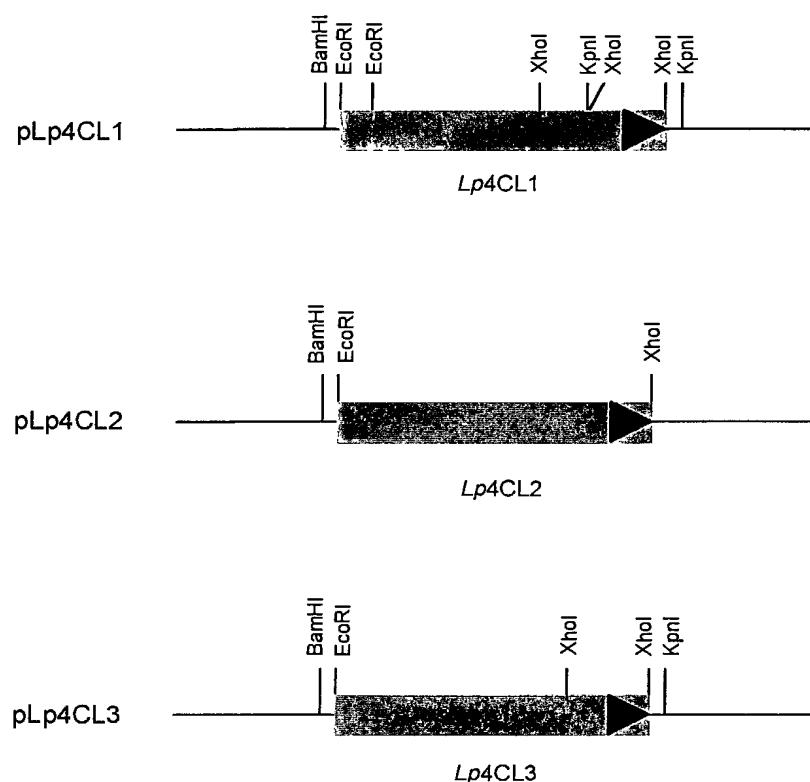
AND WHEAT IMPROVEMENT CENTER [MX/MX]; Lisboa 27, Apartado Postal 6-641, Mexico, D.F. 06600 (MX). **STATE OF SOUTH AUSTRALIA** as represented by **SOUTH AUSTRALIAN RESEARCH AND DEVELOPMENT INSTITUTE [AU/AU]**; Waite Road, Glen Osmond, South Australia 5064 (AU). **SOUTHERN CROSS UNIVERSITY [AU/AU]**; Military Road, Lismore, New South Wales 2580 (AU). **DAIRY RESEARCH AND DEVELOPMENT CORPORATION [AU/AU]**; Level 3, 84 William Street, Melbourne, Victoria 3000 (AU).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **SPANGENBERG, German, Carlos [UY/AU]**; 56 Arthur Street, Bundoora, Victoria 3083 (AU). **LIDGETT, Angela, Jane [AU/AU]**; 13 Moore Street, Richmond, Victoria 3121 (AU). **HEATH, Robyn, Louise [AU/AU]**; 3 Berry Street, Clifton Hill, Victoria 3068 (AU). **MCINNES, Russell, Leigh [AU/AU]**; Glenn College, La Trobe University, Bundoora, Victoria 3083 (AU). **LYNCH, Damian, Paul [NZ/AU]**; Unit 10, 141 Elm Street, Northcote, Victoria 3070 (AU).

[Continued on next page]

(54) Title: MODIFICATION OF LIGNIN BIOSYNTHESIS



(57) Abstract: The present invention relates to the modification of lignin biosynthesis in plants, using the nucleotide sequences encoding the enzymes 4-coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR), cinnamyl alcohol dehydrogenase (CAD) of the lignin biosynthetic pathway, from ryegrass (*Lolium*) and fescue (*Festuca*). The present invention also relates to regulatory elements, promoters capable of causing expression of exogenous genes in plants, wherein the regulatory elements are from the genes for caffeic acid Omethyl transferase (OMT), 4CL, CCR or CAD. The invention also relates to vectors including the nucleic acids and regulatory elements of the invention, plant cells, plants, plant seeds and other plant parts transformed with the regulatory elements, nucleic acids and vectors and methods using the nucleic acids, regulatory elements and vectors.



(74) **Agent:** FREEHILLS CARTER SMITH BEADLE;
Level 43, 101 Collins Street, Melbourne, Victoria 3000
(AU).

(81) **Designated States (national):** AE, AG, AL, AM, AT, AU,
AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU,
CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH,
GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC,
LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW,
MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK,
SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA,
ZW.

(84) **Designated States (regional):** ARIPO patent (GH, GM,
KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian
patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European
patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE,
IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF,
CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published:

— with international search report

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

MODIFICATION OF LIGNIN BIOSYNTHESIS

The present invention relates to the modification of lignin biosynthesis in plants and, more particularly, to enzymes involved in the lignin biosynthetic pathway and nucleic acids encoding such enzymes.

5 The present invention also relates to a regulatory element and, more particularly, to a promoter capable of causing expression of an exogenous gene in plant cells, such as a gene encoding an enzyme involved in the lignin biosynthetic pathway in plants.

10 The invention also relates to vectors including the nucleic acids and regulatory elements of the invention, plant cells, plants, seeds and other plant parts transformed with the regulatory elements, nucleic acids and vectors, and methods of using the nucleic acids, regulatory elements and vectors.

15 Lignins are complex phenolic polymers that strengthen plant cell walls against mechanical and chemical degradation. The process of lignification typically occurs during secondary thickening of the walls of cells with structural, conductive or defensive roles. Three monolignol precursors, sinapyl, coniferyl and *p*-coumaryl alcohol combine by dehydrogenative polymerisation to produce respectively the syringyl(S), guaiacyl(G) and hydroxyl(H) subunits of the lignin polymer, which can also become linked to cell-wall polysaccharides 20 through the action of peroxidases and other oxidative isozymes. In grasses, biosynthesis of the monolignol precursors is a multistep process beginning with the aromatic amino-acids phenylalanine and tyrosine. It is the final two reduction/ dehydrogenation steps of the pathway, catalysed by Cinnamoyl CoA Reductase (CCR) and Cinnamyl Alcohol Dehydrogenase (CAD) that are 25 considered to be specific to lignin biosynthesis. The proportions of monolignols incorporated into the lignin polymer vary depending on plant species, tissue, developmental stage and sub-cellular location.

Caffeic acid *O*-methyl transferase (OMT), 4 coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR) and cinnamyl alcohol dehydrogenase

- 2 -

(CAD) are key enzymes involved in lignin biosynthesis.

Worldwide permanent pasture is estimated to cover 70% of agriculturally cultivated area. Ryegrasses (*Lolium* spp.) together with the closely related fescues (*Festuca* spp.) are of significant value in temperate 5 grasslands. The commercially most important ryegrasses are Italian or annual ryegrass (*L. multiflorum* Lam.) and perennial ryegrass (*L. perenne* L.). They are the key forage species in countries where livestock production is an intensive enterprise, such as the Netherlands, United Kingdom and New 10 Zealand. The commercially most important fescues are tall fescue (*F. anundinacea* Schreb.), meadow fescue (*F. pratensis*) and red fescue (*F. rubra*).

Perennial ryegrass (*Lolium perenne* L.) is the major grass species sown in temperate dairy pastures in Australia, and the key pasture grass in temperate climates throughout the world. A marked decline of the feeding 15 value of grasses is observed in temperate pastures of Australia during late spring and early summer, where the nutritive value of perennial ryegrass based pasture is often insufficient to meet the metabolic demands of lactating dairy cattle. Perennial ryegrass is also an important turf grass.

Grass and legume *in vitro* dry matter digestibility has been negatively 20 correlated with lignin content. In addition, natural mutants of lignin biosynthetic enzymes in maize, sorghum and pearl millet that have higher rumen digestibility have been characterised as having lower lignin content and altered S/G subunit ratio. Thus, lignification of plant cell walls is the major factor identified as responsible for lowering digestibility of forage tissues as they 25 mature.

It would be desirable to have methods of altering lignin biosynthesis in plants, including grass species such as ryegrasses and fescues, by reducing the activity of key biosynthetic enzymes in order to reduce lignin content and/or alter lignin composition for enhancing dry matter digestibility and improving 30 herbage quality. However, for some applications it may be desirable to

- 3 -

enhance lignin biosynthesis to increase lignin content and/or alter lignin composition, for example to increase mechanical strength of wood, to increase mechanical strength of turf grasses, to reduce plant height and reduce lodging or improve disease resistance.

5 While nucleic acid sequences encoding some of the enzymes involved in the lignin biosynthetic pathway have been isolated for certain species of plants, there remains a need for materials useful in the modification of lignin biosynthesis in plants, particularly grass species such as ryegrasses and fescues.

10 Other phenotypic traits which may be improved by transgenic manipulation of plants include disease resistance, mineral content, nutrient quality and drought tolerance.

15 However, transgenic manipulation of phenotypic traits in plants requires the availability of regulatory elements capable of causing the expression of exogenous genes in plant cells.

It is an object of the present invention to overcome, or at least alleviate, one or more of the difficulties or deficiencies associated with the prior art.

20 In one aspect, the present invention provides substantially purified or isolated nucleic acids or nucleic acid fragments encoding the following enzymes from a ryegrass (*Lolium*) or fescue (*Festuca*) species: 4 coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR) and cinnamyl alcohol dehydrogenase (CAD).

25 The ryegrass (*Lolium*) or fescue (*Festuca*) species may be of any suitable type, including Italian or annual ryegrass, perennial ryegrass, tall fescue, meadow fescue and red fescue. Preferably the ryegrass or fescue species is a ryegrass, more preferably perennial ryegrass (*Lolium perenne*).

The nucleic acid or nucleic acid fragment may be of any suitable type

- 4 -

and includes DNA (such as cDNA or genomic DNA) and RNA (such as mRNA) that is single- or double- stranded, optionally containing synthetic, non-natural or altered nucleotide bases, and combinations thereof.

The term "isolated" means that the material is removed from its original environment (eg. the natural environment if it is naturally occurring). For example, a naturally occurring nucleic acid present in a living plant is not isolated, but the same nucleic acid separated from some or all of the coexisting materials in the natural system, is isolated. Such nucleic acids could be part of a vector and/or such nucleic acids could be part of a composition, and still be isolated in that such a vector or composition is not part of its natural environment.

In a preferred embodiment of this aspect of the invention, the substantially purified or isolated nucleic acid or nucleic acid fragment encoding 4CL includes a nucleotide sequence selected from the group consisting of (a) sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 1, 3 and 5; respectively) (b) complements of the sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 1, 3 and 5, respectively); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

In a further preferred embodiment of this aspect of the invention, the substantially purified or isolated nucleic acid or nucleic acid fragment encoding CCR includes a nucleotide sequence selected from the group consisting of (a) the sequence shown in Figure 10 hereto (Sequence ID No: 7); (b) the complement of the sequence shown in Figure 10 hereto (Sequence ID No: 7); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

In a still further preferred embodiment of this aspect of the invention, the substantially purified or isolated nucleic acid or nucleic acid fragment encoding CAD includes a nucleotide sequence selected from the group consisting of (a)

- 5 -

the sequences shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 9, 11, 14 and 16, respectively); (b) complements of the sequences shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 9, 11, 14 and 16, respectively); (c) sequences antisense to the sequences recited in (a) and (b); 5 and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

By "functionally active" is meant that the fragment or variant (such as an analogue, derivative or mutant) is capable of modifying lignin biosynthesis in a plant. Such variants include naturally occurring allelic variants and non- 10 naturally occurring variants. Additions, deletions, substitutions and derivatizations of one or more of the nucleotides are contemplated so long as the modifications do not result in loss of functional activity of the fragment or variant. Preferably the functionally active fragment or variant has at least approximately 80% identity to the relevant part of the above mentioned 15 sequence, more preferably at least approximately 90% identity, most preferably at least approximately 95% identity. Such functionally active variants and fragments include, for example, those having nucleic acid changes which result in conservative amino acid substitutions of one or more residues in the corresponding amino acid sequence. Preferably the fragment 20 has a size of at least 10 nucleotides, more preferably at least 15 nucleotides, most preferably at least 20 nucleotides.

In a second aspect of the present invention there is provided a vector including a nucleic acid or nucleic acid fragment according to the present invention.

25 In a preferred embodiment of this aspect of the invention, the vector may include a regulatory element such as a promoter, a nucleic acid or nucleic acid fragment according to the present invention and a terminator; said regulatory element, nucleic acid or nucleic acid fragment and terminator being operatively linked.

30 By "operatively linked" is meant that said regulatory element is capable

- 6 -

of causing expression of said nucleic acid or nucleic acid fragment in a plant cell and said terminator is capable of terminating expression of said nucleic acid or nucleic acid fragment in a plant cell. Preferably, said regulatory element is upstream of said nucleic acid or nucleic acid fragment and said terminator is 5 downstream of said nucleic acid or nucleic acid fragment.

The vector may be of any suitable type and may be viral or non-viral. The vector may be an expression vector. Such vectors include chromosomal, non-chromosomal and synthetic nucleic acid sequences, eg. derivatives of plant viruses; bacterial plasmids; derivatives of the Ti plasmid from 10 *Agrobacterium tumefaciens*; derivatives of the Ri plasmid from *Agrobacterium rhizogenes*; phage DNA; yeast artificial chromosomes; bacterial artificial chromosomes; binary bacterial artificial chromosomes; vectors derived from combinations of plasmids and phage DNA. However, any other vector may be used as long as it is replicable or integrative or viable in the plant cell.

15 The regulatory element and terminator may be of any suitable type and may be endogenous to the target plant cell or may be exogenous, provided that they are functional in the target plant cell.

Preferably the regulatory element is a promoter. A variety of promoters which may be employed in the vectors of the present invention are well known 20 to those skilled in the art. Factors influencing the choice of promoter include the desired tissue specificity of the vector, and whether constitutive or inducible expression is desired and the nature of the plant cell to be transformed (eg. monocotyledon or dicotyledon). Particularly suitable promoters include the Cauliflower Mosaic Virus 35S (CaMV 35S) promoter, the 25 maize Ubiquitin promoter, the rice Actin promoter, and ryegrass endogenous OMT, 4CL, CCR or CAD promoters.

A variety of terminators which may be employed in the vectors of the present invention are also well known to those skilled in the art. The terminator may be from the same gene as the promoter sequence or a different gene. 30 Particularly suitable terminators are polyadenylation signals, such as the

CaMV 35S polyA and other terminators from the nopaline synthase (*nos*) and the octopine synthase (*ocs*) genes.

The vector, in addition to the regulatory element, the nucleic acid or nucleic acid fragment of the present invention and the terminator, may include 5 further elements necessary for expression of the nucleic acid or nucleic acid fragment, in different combinations, for example vector backbone, origin of replication (ori), multiple cloning sites, spacer sequences, enhancers, introns (such as the maize Ubiquitin Ubi intron), antibiotic resistance genes and other 10 selectable marker genes [such as the neomycin phosphotransferase (*npt2*) gene, the hygromycin phosphotransferase (*hph*) gene, the phosphinothricin acetyltransferase (*bar* or *pat*) gene], and reporter genes (such as beta-glucuronidase (GUS) gene (*gusA*]). The vector may also contain a ribosome 15 binding site for translation initiation. The vector may also include appropriate sequences for amplifying expression.

15 As an alternative to use of a selectable marker gene to provide a phenotypic trait for selection of transformed host cells, the presence of the vector in transformed cells may be determined by other techniques well known in the art, such as PCR (polymerase chain reaction), Southern blot hybridisation analysis, histochemical GUS assays, northern and Western blot 20 hybridisation analyses.

Those skilled in the art will appreciate that the various components of the vector are operatively linked, so as to result in expression of said nucleic acid or nucleic acid fragment. Techniques for operatively linking the components of the vector of the present invention are well known to those 25 skilled in the art. Such techniques include the use of linkers, such as synthetic linkers, for example including one or more restriction enzyme sites.

The vectors of the present invention may be incorporated into a variety of plants, including monocotyledons (such as grasses from the genera *Lolium*, *Festuca*, *Paspalum*, *Pennisetum*, *Panicum* and other forage and turf grasses, 30 corn, oat, sugarcane, wheat and barley), dicotyledons (such as *arabidopsis*,

tobacco, legumes, alfalfa, oak, eucalyptus, maple, canola, soybean and chickpea) and gymnosperms. In a preferred embodiment, the vectors are used to transform monocotyledons, preferably grass species such as ryegrasses (*Lolium* species) and fescues (*Festuca* species), more preferably 5 perennial ryegrass (*Lolium perenne*) including forage and turf type cultivars.

Techniques for incorporating the vectors of the present invention into plant cells (for example by transduction, transfection or transformation) are well known to those skilled in the art. Such techniques include *Agrobacterium* mediated introduction, electroporation to tissues, cells and protoplasts, 10 protoplast fusion, injection into reproductive organs, injection into immature embryos and high velocity projectile introduction to cells, tissues, calli, immature and mature embryos. The choice of technique will depend largely on the type of plant to be transformed.

Cells incorporating the vector of the present invention may be selected, 15 as described above, and then cultured in an appropriate medium to regenerate transformed plants, using techniques well known in the art. The culture conditions, such as temperature, pH and the like, will be apparent to the person skilled in the art. The resulting plants may be reproduced, either sexually or asexually, using methods well known in the art, to produce 20 successive generations of transformed plants.

In a further aspect of the present invention there is provided a plant cell, plant, plant seed or other plant part, including, eg transformed with, a vector of the present invention.

The plant cell, plant, plant seed or other plant part may be from any 25 suitable species, including monocotyledons, dicotyledons and gymnosperms. In a preferred embodiment the plant cell, plant, plant seed or other plant part may be from a monocotyledon, preferably a grass species, more preferably a ryegrass (*Lolium* species) or fescue (*Festuca* species), even more preferably a ryegrass, most preferably perennial ryegrass, including forage- and turf-type 30 cultivars.

The present invention also provides a plant, plant seed or other plant part derived from a plant cell of the present invention.

The present invention also provides a plant, plant seed or other plant part derived from a plant of the present invention.

5 In a further aspect of the present invention there is provided a method of modifying lignin biosynthesis in a plant, said method including introducing into said plant an effective amount of a nucleic acid or nucleic acid fragment and/or a vector according to the present invention.

By "an effective amount" is meant an amount sufficient to result in an
10 identifiable phenotypic trait in said plant, or a plant, plant seed or other plant part derived therefrom. Such amounts can be readily determined by an appropriately skilled person, taking into account the type of plant, the route of administration and other relevant factors. Such a person will readily be able to determine a suitable amount and method of administration. See, for example,
15 Maniatis et al, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, the entire disclosure of which is incorporated herein by reference.

Using the methods and materials of the present invention, plant lignin biosynthesis may be increased, decreased or otherwise modified relative to an
20 untransformed control plant. It may be increased or otherwise modified, for example, by incorporating additional copies of a sense nucleic acid or nucleic acid fragment of the present invention. It may be decreased, for example, by incorporating an antisense nucleic acid or nucleic acid fragment of the present invention. In addition, the number of copies of genes encoding for different
25 enzymes in the lignin biosynthetic pathway may be manipulated to modify the relative amount of each monolignol synthesized, thereby leading to the formation of lignin having altered composition.

In a still further aspect of the present invention there is provided use of a nucleic acid or nucleic acid fragment according to the present invention, and/or

- 10 -

nucleotide sequence information thereof, and/or single nucleotide polymorphisms thereof, as a molecular genetic marker.

More particularly, nucleic acids or nucleic acid fragments according to the present invention, and/or nucleotide sequence information thereof, and/or 5 single nucleotide polymorphisms thereof, may be used as a molecular genetic marker for qualitative trait loci (QTL) tagging, mapping, DNA fingerprinting and in marker assisted selection, and may be used as candidate genes or perfect markers, particularly in ryegrasses and fescues. Even more particularly, 10 nucleic acids or nucleic acid fragments according to the present invention, and/or nucleotide sequence information thereof, may be used as molecular genetic markers in forage and turf grass improvement, eg. tagging QTLs for dry matter digestibility, herbage quality, mechanical stress tolerance, disease resistance, insect pest resistance, plant stature and leaf and stem colour.

In a still further aspect of the present invention there is provided a 15 substantially purified or isolated polypeptide from a ryegrass (*Lolium*) or fescue (*Festuca*) species, selected from the group consisting of the enzymes 4CL, CCR and CAD.

The ryegrass (*Lolium*) or fescue (*Festuca*) species may be of any suitable type, including Italian or annual ryegrass, perennial ryegrass, tall 20 fescue, meadow fescue and red fescue. Preferably the species is a ryegrass, more preferably perennial ryegrass (*L. perenne*).

In a preferred embodiment of this aspect of the invention, the substantially purified or isolated enzyme 4CL includes an amino acid sequence selected from the group consisting of sequences shown in Figures 2, 3 and 4 25 hereto (Sequence ID Nos: 2, 4 and 6, respectively); and functionally active fragments and variants thereof.

In a further preferred embodiment of this aspect of the invention, the substantially purified or isolated enzyme CCR includes an amino acid sequence selected from the group consisting of the sequence shown in Figure

- 11 -

10 hereto (Sequence ID No: 8); and functionally active fragments and variants thereof.

In a still further preferred embodiment of this aspect of the invention, the substantially purified or isolated enzyme CAD includes an amino acid sequence selected from the group consisting of the sequence shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 10, 12, 15 and 17, respectively); and functionally active fragments and variants thereof.

By "functionally active" in this context is meant that the fragment or variant has one or more of the biological properties of the enzymes 4CL, CCR and CAD, respectively. Additions, deletions, substitutions and derivatizations of one or more of the amino acids are contemplated so long as the modifications do not result in loss of functional activity of the fragment or variant. Preferably the fragment or variant has at least approximately 60% identity to the relevant part of the above mentioned sequence, more preferably at least approximately 80% identity, most preferably at least approximately 90% identity. Such functionally active variants and fragments include, for example, those having conservative amino acid substitutions of one or more residues in the corresponding amino acid sequence. Preferably the fragment has a size of at least 10 amino acids, more preferably at least 15 amino acids, most preferably at least 20 amino acids.

In a further embodiment of this aspect of the invention, there is provided a polypeptide recombinantly produced from a nucleic acid or nucleic acid fragment according to the present invention. Techniques for recombinantly producing polypeptides are well known to those skilled in the art.

25 In a still further aspect of the present invention there is provided a lignin or modified lignin substantially or partially purified or isolated from a plant, plant seed or other plant part of the present invention.

Such lignins may be modified from naturally occurring lignins in terms of the length, the degree of polymerisation (number of units), degree of branching

- 12 -

and/or nature of linkages between units.

In a still further aspect, the present invention provides an isolated regulatory element capable of causing expression of an exogenous gene in plant cells. Preferably the regulatory element is isolated from a nucleic acid or 5 nucleic acid fragment encoding OMT, 4CL, CCR or CAD.

The regulatory element may be a nucleic acid molecule, including DNA (such as cDNA or genomic DNA) and RNA (such as mRNA) that is single- or double- stranded, optionally containing synthetic, non-natural or altered nucleotide bases, and combinations thereof.

10 Preferably the regulatory element includes a promoter, more preferably an *O*-methyltransferase promoter, even more preferably an *O*-methyltransferase promoter from a ryegrass (*Lolium*) or fescue (*Festuca*) species, more preferably a ryegrass, most preferably perennial ryegrass (*Lolium perenne*).

15 In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a promoter from the caffeic acid *O*-methyltransferase gene corresponding to the cDNA homologue *LpOMT1* from perennial ryegrass.

20 Preferably the regulatory element includes a nucleotide sequence including the first approximately 4630 nucleotides of the sequence shown in Figure 18 hereto (Sequence ID No: 13); or a functionally active fragment or variant thereof.

25 By "functionally active" in this context is meant that the fragment or variant (such as an analogue, derivative or mutant) is capable of causing expression of a transgene in plant cells. Such variants include naturally occurring allelic variants and non-naturally occurring variants. Additions, deletions, substitutions and derivatizations of one or more of the nucleotides are contemplated so long as the modifications do not result in loss of functional

- 13 -

activity of the regulatory element. Preferably the functionally active fragment or variant has at least approximately 80% identity to the relevant part of the above sequence, more preferably at least approximately 90% identity, most preferably at least approximately 95% identity. Preferably the fragment has a 5 size of at least 100 nucleotides, more preferably at least 150 nucleotides, most preferably at least 200 nucleotides.

In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a nucleotide sequence selected from the group consisting of:

10 Nucleotides -4581 to -1

Nucleotides -4285 to -1

Nucleotides -4020 to -1

Nucleotides -2754 to -1

Nucleotides -1810 to -1

15 Nucleotides -831 to -1

Nucleotides -560 to -1

Nucleotides -525 to -1

Nucleotides -274 to -1

Nucleotides -21 to -1

20 of Figure 18 hereto (Sequence ID No: 13);

or a functionally active fragment or variant thereof.

In another preferred embodiment the regulatory element includes a 4 coumarate-CoA ligase promoter, even more preferably a 4 coumarate-CoA ligase promoter from a ryegrass (*Lolium*) or fescue (*Festuca*) species, more 25 preferably a ryegrass, most preferably perennial ryegrass (*Lolium perenne*).

- 14 -

In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a promoter from the 4 coumarate-CoA ligase gene corresponding to the cDNA homologue *Lp4CL2* from perennial ryegrass.

Preferably the regulatory element includes a nucleotide sequence 5 including the first approximately 2206 nucleotides of the sequence shown in Figure 38 hereto (Sequence ID No: 17); or a functionally active fragment or variant thereof.

By "functionally active" in this context is meant that the fragment or variant (such as an analogue, derivative or mutant) is capable of causing 10 expression of a transgene in plant cells. Such variants include naturally occurring allelic variants and non-naturally occurring variants. Additions, deletions, substitutions and derivatizations of one or more of the nucleotides are contemplated so long as the modifications do not result in loss of functional activity of the regulatory element. Preferably the functionally active fragment or 15 variant has at least approximately 80% identity to the relevant part of the above sequence, more preferably at least approximately 90% identity, most preferably at least approximately 95% identity. Preferably the fragment has a size of at least 100 nucleotides, more preferably at least 150 nucleotides, most preferably at least 200 nucleotides.

20 In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a nucleotide sequence selected from the group consisting of:

Nucleotides – 2206 to –1

Nucleotides -1546 to –1

25 Nucleotides –1186 to –1

Nucleotides –406 to –1

Nucleotides – 166 to –1

- 15 -

of Figure 38 hereto (Sequence ID No: 17);

or a functionally active fragment or variant thereof.

In another preferred embodiment the regulatory element includes a cinnamoyl-CoA reductase promoter, even more preferably a cinnamoyl-CoA reductase promoter from a ryegrass (*Lolium*) or fescue (*Festuca*) species, 5 more preferably a ryegrass, most preferably perennial ryegrass (*Lolium perenne*).

In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a promoter from the cinnamoyl-CoA reductase 10 gene corresponding to the *LpCCR1* cDNA from perennial ryegrass.

Preferably the regulatory element includes a nucleotide sequence including the first approximately 6735 nucleotides of the sequence shown in Figure 39 hereto (Sequence ID No: 18); or a functionally active fragment or variant thereof.

15 By "functionally active" in this context is meant that the fragment or variant (such as an analogue, derivative or mutant) is capable of causing expression of a transgene in plant cells. Such variants include naturally occurring allelic variants and non-naturally occurring variants. Additions, deletions, substitutions and derivatizations of one or more of the nucleotides 20 are contemplated so long as the modifications do not result in loss of functional activity of the regulatory element. Preferably the functionally active fragment or variant has at least approximately 80% identity to the relevant part of the above sequence, more preferably at least approximately 90% identity, most preferably at least approximately 95% identity. Preferably the fragment has a 25 size of at least 100 nucleotides, more preferably at least 150 nucleotides, most preferably at least 200 nucleotides.

- 16 -

In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a nucleotide sequence selected from the group consisting of:

Nucleotides - 6735 to -1
5 Nucleotides -5955 to -1
Nucleotides -5415 to -1
Nucleotides -4455 to -1
Nucleotides - 4035 to -1
Nucleotides -3195 to -1
10 Nucleotides -2595 to -1
Nucleotides -1755 to -1
Nucleotides -1275 to -1
Nucleotides -495 to -1
Nucleotides -255 to -1
15 Nucleotides -75 to -1

of Figure 39 hereto (Sequence ID No: 18);

or a functionally active fragment or variant thereof.

By an "exogenous gene" is meant a gene not natively linked to said regulatory element. In certain embodiments of the present invention the 20 exogenous gene is also not natively found in the relevant plant or plant cell.

The exogenous gene may be of any suitable type. The exogenous gene may be a nucleic acid such as DNA (e.g. cDNA or genomic DNA) or RNA (e.g. mRNA), and combinations thereof. The exogenous gene may correspond to a target gene, for example a gene capable of influencing disease resistance, 25 herbage digestibility, nutrient quality, mineral content or drought tolerance or be a fragment or variant (such as an analogue, derivative or mutant) thereof

which is capable of modifying expression of said target gene. Such variants include nucleic acid sequences which are antisense to said target gene or an analogue, derivative, mutant or fragment thereof. The transgene may code for a protein or RNA sequence depending the target condition and whether down 5 or up-regulation of gene expression is required. Preferably, the target gene is selected from exogenous coding sequences coding for mRNA for a protein, this protein may be of bacterial origin (such as enzymes involved in cell wall modification and cell wall metabolism, cytokinin biosynthesis), or eukaryotic origin (such as pharmaceutically active polypeptides) or of plant origin (such as 10 enzymes involved in the synthesis of phenolic compounds, cell wall metabolism, sugar metabolism, lignin biosynthesis). Preferably, the target gene is selected from the group comprising *O*-methyltransferase, 4 coumarate CoA-ligase, cinnamoyl CoA reductase, cinnamyl alcohol dehydrogenase, cinnamate 4 hydroxylase, phenolase, laccase, peroxidase, coniferol glucosyl 15 transferase, coniferin beta-glucosidase, phenylalanine ammonia lyase, ferulate 5-hydroxylase, chitinase, glucanase, isopentenyltransferase, xylanase.

The plant cells, in which the regulatory element of the present invention is capable of causing expression of an exogenous gene, may be of any suitable type. The plant cells may be from monocotyledons (such as grasses 20 from the genera *Lolium*, *Festuca*, *Paspalum*, *Pennisetum*, *Panicum* and other forage and turf grasses, corn, grains, oat, sugarcane, wheat and barley), dicotyledons (such as arabidopsis, tobacco, legumes, alfalfa, oak, eucalyptus and maple) and gymnosperms. Preferably the plant cells are from a monocotyledon, more preferably a grass species such as a ryegrass (*Lolium*) 25 or fescue (*Festuca*) species, even more preferably a ryegrass, most preferably perennial ryegrass (*Lolium perenne*).

The regulatory element according to the present invention may be used to express exogenous genes to which it is operatively linked in the production of transgenic plants.

30 Accordingly, in a further aspect of the present invention there is provided a vector including a regulatory element according to the present

invention.

In a preferred embodiment of this aspect of the invention, the vector may include a regulatory element according to the present invention, an exogenous gene as hereinbefore described, and a terminator; said regulatory element, exogenous gene and terminator being operatively linked, such that said regulatory element is capable of causing expression of said exogenous gene in plant cells. Preferably, said regulatory element is upstream of said exogenous gene and said terminator is downstream of said exogenous gene.

The vector may be of any suitable type and may be viral or non-viral.

10 The vector may be an expression vector. Such vectors include chromosomal, non-chromosomal and synthetic nucleic acid sequences, eg. derivatives of plant viruses; bacterial plasmids; derivatives of the Ti plasmid from *Agrobacterium tumefaciens*; derivatives of the Ri plasmid from *Agrobacterium rhizogenes*; phage DNA; yeast artificial chromosomes; bacterial artificial chromosomes; binary bacterial artificial chromosomes; vectors derived from combinations of plasmids and phage DNA. However, any other vector may be used as long as it is replicable on integrative or viable in the plant cell.

The terminator may be of any suitable type and includes for example polyadenylation signals, such as the Cauliflower Mosaic Virus 35S polyA (CaMV 35S polyA) and other terminators from the nopaline synthase (*nos*) and the octopine synthase (*ocs*) genes.

The vector, in addition to the regulatory element, the exogenous nucleic acid and the terminator, may include further elements necessary for expression of the nucleic acid, in different combinations, for example vector backbone, origin of replication (ori), multiple cloning sites, spacer sequences, enhancers, introns (such as the maize Ubiquitin Ubi intron), antibiotic resistance genes and other selectable marker genes [such as the neomycin phosphotransferase (*npt*2) gene, the hygromycin phosphotransferase (*hph*) gene, the phosphinothricin acetyltransferase (*bar* or *pat*) gene], and reporter genes (such as beta-glucuronidase (GUS) gene (*gusA*)]. The vector may also

- 19 -

contain a ribosome binding site for translation initiation. The vector may also include appropriate sequences for amplifying expression.

The regulatory element of the present invention may also be used with other full promoters or partial promoter elements.

5 As an alternative to use of a selectable marker gene to provide a phenotypic trait for selection of transformed host cells, the presence of the vector in transformed cells may be determined by other techniques well known in the art, such as PCR (polymerase chain reaction), Southern blot hybridisation analysis, histochemical GUS assays, northern and Western blot 10 hybridisation analyses.

Those skilled in the art will appreciate that the various components of the vector are operatively linked, so as to result in expression of said transgene. Techniques for operatively linking the components of the vector of the present invention are well known to those skilled in the art. Such 15 techniques include the use of linkers, such as synthetic linkers, for example including one or more restriction sites.

The vectors of the present invention may be incorporated into a variety of plants, including monocotyledons, dicotyledons and gymnosperms. In a preferred embodiment the vectors are used to transform monocotyledons, 20 preferably grass species such as ryegrasses (*Lolium* species) and fescues (*Festuca* species), more preferably perennial ryegrass (*Lolium perenne*) including forage- and turf- type cultivars.

Techniques for incorporating the vectors of the present invention into plant cells (for example by transduction, transfection or transformation) are 25 well known to those skilled in the art. Such techniques include *Agrobacterium* mediated introduction, electroporation to tissues, cells and protoplasts, protoplast fusion, injection into reproductive organs, injection into immature embryos and high velocity projectile introduction to cells, tissues, calli, immature and mature embryos. The choice of technique will depend largely

- 20 -

on the type of plant to be transformed.

Cells incorporating the vector of the present invention may be selected, as described above, and then cultured in an appropriate medium to regenerate transformed plants, using techniques well known in the art. The culture 5 conditions, such as temperature, pH and the like, will be apparent to the person skilled in the art. The resulting plants may be reproduced, either sexually or asexually, using methods well known in the art, to produce successive generations of transformed plants.

In a further aspect of the present invention there is provided a plant cell, 10 plant, plant seed or other plant part, including, eg. transformed with, a vector of the present invention.

The plant cell, plant, plant seed or other plant part may be from any suitable species, including monocotyledons, dicotyledons and gymnosperms. In a preferred embodiment the plant cell, plant, plant seed or other plant part is 15 from a monocotyledon, preferably a grass species, more preferably a ryegrass (*Lolium* species) or fescue (*Festuca* species), even more preferably perennial ryegrass (*Lolium perenne*), including forage- and turf-type cultivars.

The present invention also provides a plant, plant seed, or other plant part derived from a plant cell of the present invention.

20 The present invention also provides a plant, plant seed or other plant part derived from a plant of the present invention.

In a still further aspect of the present invention there is provided a recombinant plant genome including a regulatory element according to the present invention.

25 In a preferred embodiment of this aspect of the invention the recombinant plant genome further includes an exogenous gene operatively linked to said regulatory element.

- 21 -

In a further aspect of the present invention there is provided a method for expressing an exogenous gene in plant cells, said method including introducing into said plant cells an effective amount of a regulatory element and/or a vector according to the present invention.

5 By "an effective amount" is meant an amount sufficient to result in an identifiable phenotypic change in said plant cells or a plant, plant seed or other plant part derived therefrom. Such amounts can be readily determined by an appropriately skilled person, taking into account the type of plant cell, the route of administration and other relevant factors. Such a person will readily be able
10 to determine a suitable amount and method of administration. See, for example, Maniatis et al, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, the entire disclosure of which is incorporated herein by reference.

15 The present invention will now be more fully described with reference to the accompanying Examples and drawings. It should be understood, however, that the description following is illustrative only and should not be taken in any way as a restriction on the generality of the invention described above.

In the Figures

20 Figure 1 shows plasmid maps of the three cDNAs encoding perennial ryegrass 4CL homologues.

Figure 2 shows the nucleotide (Sequence ID No: 1) and amino acid (Sequence ID No: 2) sequences of *Lp4CL1*.

Figure 3 shows the nucleotide (Sequence ID No: 3) and amino acid (Sequence ID No: 4) sequences of *Lp4CL2*.

25 Figure 4 shows the nucleotide (Sequence ID No: 5) and amino acid (Sequence ID No: 6) sequences of *Lp4CL3*.

- 22 -

Figure 5 shows amino acid sequence alignment of deduced proteins encoded by *Lp4CL1* (Sequence ID No: 2), *Lp4CL2* (Sequence ID No: 4) and *Lp4CL3* (Sequence ID No: 6).

Figure 6 shows northern hybridisation analysis of developing perennial ryegrass using *Lp4CL1*, *Lp4CL2* and *Lp4CL3* as hybridisation probes. SR: roots from seedlings (3-5 d post-germination), SS: shoots from seedlings (3-5 d post-germination), ML: leaves from 12-week-old plants, MS: stems from 12-week-old plants. Blots were washed in 0.2 X SSPE, 0.1 % SDS at 65 °C. *Lp4CL1*, *Lp4CL2* and *Lp4CL3* do not cross hybridise at this stringency. Sizes are given in kb.

Figure 7 shows northern hybridisation analysis showing the time course of expression of 4CL mRNA in wounded perennial ryegrass leaves. Sizes are given in kb.

Figure 8 shows genomic Southern hybridisation analysis using *Lp4CL1*, *Lp4CL2* and *Lp4CL3* as hybridisation probes. 10 µg of digested perennial ryegrass genomic DNA or 20 µg of digested tall fescue genomic DNA were separated on a 1.0 % agarose gel, transferred to Hybond N⁺ membranes and then hybridised with ³²P labelled *Lp4CL1*, *Lp4CL2* or *Lp4CL3* probes. The ryegrass *Lp4CL1*, *Lp4CL2* and *Lp4CL3* genes reveal homologous sequences in tall fescue and indicate that the ryegrass 4CL genes can be used to isolate and to manipulate the expression of the tall fescue (*Festuca arundinacea*) 4CL genes.

Figure 9 shows restriction map of *LpCCR1*. An *L. perenne* seedling cDNA library constructed in Uni-ZAPTM (Stratagene) was screened in a solution containing 10xPIPES, 50% deionised formamide and 10% SDS at 42°C. Filters were washed at room temperature, three times in 0.1% SDS, 2× SSPE and then twice in 0.1% SDS, 0.2× SSPE. The location of the probe used for northern and Southern hybridisation analyses is indicated by the black line labelled *LpCCR531*.

- 23 -

Figure 10 shows the nucleotide (Sequence ID No: 7) and amino acid (Sequence ID No: 8) sequences of *LpCCR1*.

Figure 11 shows Southern hybridisation analysis of DNA from double haploid (DH) perennial ryegrass using *LpCCR1* as hybridisation probe. 10 μ g of DH genomic DNA was digested with DraI, BamHI, EcoRI, EcoRV, HindIII or XbaI, separated on a 1% agarose gel and then capillary blotted onto nylon membrane (Amersham Hybond-N). The membrane was probed with the digoxigenin (DIG) labelled *LpCCR531* fragment at 25ng/ml in the hybridisation solution. Hybridisation was in 4x SSC, 50% formamide, 0.1% N-Lauroyl-sarcosine, 0.02% SDS, 2% Blocking solution at 42°C. The membrane was washed twice for five minutes in 2x SSC, 0.1% SDS at room temperature, then twice for fifteen minutes in 0.5x SSC, 0.1% SDS at 68°C. Molecular weight was determined by comparison to a DIG-labelled marker (Roche Molecular Biochemicals).

Figure 12 shows northern hybridisation analysis of RNA samples from different organs and developmental stages of perennial ryegrass using *LpCCR1* probe. Roots from seedlings (3-5 d post-germination), shoots from seedlings (3-5 d post-germination), roots from seedlings (7-10 d post-germination), leaves from seedlings (7-10 d post-germination), roots from 6 and 10 week old plants, leaves from 6 and 10 week old plants, stems from 6 and 10 week old plants, whole seedling from 11 day old *Phalaris* and 7 day old *Festuca*.

Total RNA was isolated using Trizol (GibcoBRL) and 15 μ g was separated on a 1.2% Agarose gel containing 6% formamide and then capillary blotted onto nylon membrane (Amersham Hybond-N). The membrane was stained with 0.2% methylene blue/0.3M sodium acetate to visualise the marker and ensure that RNA was evenly loaded. 50 ng *LpCCR531* was random-labelled with 32 P-dCTP (Amersham Megaprime) and hybridisation conditions were 4x SSC, 50% formamide, 0.5% SDS, 5x denhardt solution, 5% dextrane sulphate, 0.1% Herring sperm DNA at 42°C over-night. The ryegrass *LpCCR1*

- 24 -

gene reveal homologous transcripts in tall fescue and *Phalaris*, thus indicating that the ryegrass CCR gene can be used to manipulate the expression of the tall fescue (*Festuca arundinacea*) and *Phalaris* CCR endogenous genes.

5 Figure 13 shows the nucleotide (Sequence ID No: 9) and amino acid (Sequence ID No: 10) sequences of *LpCAD1*.

Figure 14 shows the nucleotide (Sequence ID No: 11) and amino acid (Sequence ID No: 12) sequences of *LpCAD2*.

Figure 15 shows a plasmid map of a cDNA clone encoding perennial ryegrass CAD homologue *LpCAD1*.

10 Figure 16 shows northern hybridisation analysis of RNA samples from different organs and developmental stages of perennial ryegrass using A) *LpCAD1* and B) *LpCAD2* as hybridisation probes. Roots from seedlings 3-5 d post-germination, 7-10 d post-germination, 6 weeks and 10 weeks, Shoots from seedlings 3-5 d post-germination and 7-10 d post-germination, Leaves from 6 week old and 10 week old plants, stem tissue from 6 and 10 week old plants. RNA isolated from *Phalaris* and *Festuca* 11 and 7 day old seedlings. The ryegrass CAD genes reveal homologous transcripts in tall fescue and *Phalaris*, thus indicating that the ryegrass CAD gene can be used to manipulate the expression of the tall fescue and *Phalaris* CAD endogenous genes.

15

20

Figure 17 shows genomic Southern hybridisation analysis. 10 µg of perennial ryegrass genomic DNA digested with a range of restriction enzymes was separated on a 0.8% agarose gel, transferred to Hybond N and then hybridised with a DIG labelled A) *LpCAD1*, and B) *LpCAD2* hybridisation probe.

25 Figure 18 shows the nucleotide sequence of the *LpOmt1* promoter (Sequence ID No: 13).

- 25 -

Figure 19 shows a plasmid map of plant transformation vector carrying the reporter β -glucuronidase (GUS) gene (*gusA*) under control of the perennial ryegrass *LpOmt1* promoter.

Figure 20 (upper image) shows PCR analysis of transgenic tobacco plants containing the *gusA* gene under the control of the perennial ryegrass *LpOMT1* promoter (upper figure). PCR reactions using *gusA*-specific primers were performed. Figure 20 (lower images) show histochemical GUS assays, demonstrating xylem-specific *gusA* expression (A and B) and *gusA* expression in glandular leaf trichomes (C and D) in transgenic tobacco plants containing the *gusA* gene under the control of the perennial ryegrass *LpOMT1* promoter.

Figure 21 shows the isolation of the *LpCCR1* genomic clone 1. A) Southern hybridization analysis of CCR genomic clone λ Lp6.1.1a digested with *Xba*I, *Nco*I, *Sac*I, *Xho*I, *Xho*I/*Sac*I DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with a DIG labelled CCR1 probe. B) Map showing the genomic gene organisation of *LpCCR1* clone 1 based on sequence results. C) Comparison of plant CCR exon size and number in different plant species (*Lolium perenne*, *Lp.*, *Eucalyptus gunni*, *Eg.*, *Eucalyptus saligna*, *Es.*, *Populus balsamifera*, *Pb.*)

Figure 22 shows the isolation of the *LpCCR1* genomic clone 2. A) Southern hybridization analysis of CCR genomic clone λ Lp6.1.1a digested with *Xba*I, *Nco*I, *Sac*I, *Xho*I, *Xho*I/*Sac*I DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with 200bp of the CCR1 promoter (Figure 21B). B) Map showing the promoter region of *LpCCR1* clone 2 based on sequence results.

Figure 23 shows the isolation of an *Lp4CL* genomic clone. A) Southern hybridisation analysis of 4CL genomic clone λ Lp4CL2 digested with *Bam*HI, *Kpn*I or *Sac*I. DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with a DIG labelled 4CL1 hybridisation probe. B) 10 μ l of a standard PCR reaction using forward and reverse oligonucleotides

- 26 -

designed to positions outlined on C). The PCR products were separated on a 0.8% agarose gel and stained with ethidium bromide. C) Map showing the genomic gene organisation of $\lambda Lp4CL2$ based on sequence and PCR results.

Figure 24 shows the isolation of an *Lp4CL* genomic clone. A) Southern hybridisation analysis of 4CL genomic clone $\lambda Lp4CL2$ digested with BamHI, KpnI, Sall. DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with a DIG labelled 4CL1 probe. B) Map showing the genomic gene organisation of *Lp4CL2* clone 1 and the promoter region of clone 2.

Figure 25 shows plasmid map of plant transformation vector carrying the *gusA* gene under control of the perennial ryegrass *Lp4CL2* promoter (*Lp4CL2::gusA*).

Figure 26 shows nucleotide (Sequence ID No: 14) and amino acid (Sequence ID No: 15) sequences of genomic clone CAD2 cv Barlano (Intron 1 and first 111 bp of the coding region are missing).

Figure 27 shows nucleotide (Sequence ID No: 16) and amino acid (Sequence ID No: 15) sequences of coding sequence deduced from genomic clone CAD2 cv Barlano (region in bold is missing from the genomic clone).

Figure 28 shows the isolation of *LpCAD2* genomic clone. A) Southern hybridization analysis of CAD genomic clone $\lambda LpCAD2$ digested with BamHI, EcoRI, KpnI, Sall or XbaI. DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with a DIG labelled CAD2 hybridisation probe. B) Map showing the genomic gene organisation of $\lambda LpCAD2$ based on sequence results.

Figure 29 shows A) Sense and antisense *Lp4CL1*, *Lp4CL2* and *Lp4CL3* transformation vectors under control of the CaMV 35S promoter; B) Sense and antisense *Lp4CL1*, *Lp4CL2* and *Lp4CL3* transformation vectors under control

of the maize ubiquitin promoter.

Figure 30 shows A) Sense and antisense *LpCCR1* transformation vectors under control of the CaMV 35S promoter; B) Sense and antisense *LpCCR1* transformation vectors under control of the maize ubiquitin promoter.

5 Figure 31 shows A) Sense and antisense *LpCAD1* transformation vectors under control of the CaMV 35S promoter; B) Sense and antisense *LpCAD1* transformation vectors under control of the maize ubiquitin promoter.

Figure 32 shows molecular analysis of *Lp4CL1*-transgenic tobacco. A) Plasmid map of transformation vector carrying a chimeric sense *Lp4CL1* gene. 10 B) PCR analysis of independent transgenic tobacco clones using *Lp4CL1* specific primers. C) Southern hybridization analysis of independent transgenic tobacco plants using an *Lp4CL1* specific probe. D) Northern hybridization analysis of independent transgenic tobacco plants using an *Lp4CL1* specific probe.

15 Figure 33 shows molecular analysis of *LpCCR1*-transgenic tobacco. A) Plasmid map of transformation vectors carrying a chimeric sense and antisense *LpCCR1* gene. B) PCR analysis of independent sense transgenic tobacco clones using *LpCCR1* specific primers.

Figure 34 shows protocol for suspension culture-independent 20 production of transgenic perennial ryegrass plants. A) Isolated zygotic embryos, plated on MSM5 medium, day 0; B) Embryogenic callus formation and proliferation, 6 - 8 weeks after embryo isolation; C) Embryogenic calli arranged on high osmotic MSM3Plus medium prior to biolistic transformation; D) Histochemical GUS assay showing GUS expressing foci 3 – 4 days post-25 bombardment of chimeric *gusA* gene; E) Selection of embryogenic calli on MSM3 medium containing 100 mg/l paromomycin (Pm), 2 weeks after microprojectile bombardment; F) Regeneration of Pm resistant shoots on MSK medium containing 100 mg/l Pm, 4 weeks after microprojectile bombardment; G) *In vitro* plant regeneration from PM resistant embryogenic calli, 6 weeks

- 28 -

after microprojectile bombardment; H) Transgenic perennial ryegrass plants 28 weeks after embryo isolation.

Figure 35 shows molecular analysis of transgenic perennial ryegrass plants carrying sense and antisense *LpOmt1* transgenes. Plasmid maps of 5 vectors used for the co-transformation of perennial ryegrass embryogenic calli; pHp23 carrying a chimeric neomycin phosphotransferase (*npt2*) selectable marker gene; pUbiomt1 carrying a maize ubiquitin promoter driven sense *LpOmt1* gene; pUbitmo1 carrying a maize ubiquitin promoter driven antisense *LpOmt1* gene (top). PCR analysis using *npt2*-specific primers of 5 independent 10 transgenic perennial ryegrass plants from biolistic transformation with sense and antisense *LpOmt1* vectors (upper centre). Southern hybridization analysis with an *omt1* hybridization probe of 7 independent perennial ryegrass plants co-transformed with sense (lanes 1-3) and antisense (lanes 4-7) *LpOmt1* vectors (lower centre left). Southern hybridisation analysis with an *npt2* 15 hybridisation probe of independent perennial ryegrass plants (lower centre right). Northern hybridisation analysis of perennial ryegrass plants co-transformed with antisense *LpOmt1* vector (bottom). C = negative control untransformed perennial ryegrass; P = positive plasmid control.

Figure 36 shows biochemical analysis of *LpOmt1*-transgenic perennial 20 ryegrass. OMT activity of leaf samples from selected independent *LpOmt1*-transgenic perennial ryegrass plants (Ell8, Ell11, Ell14 and Ell15) was determined and compared to untransformed perennial ryegrass negative control plant *L. perenne* cv. Ellett (wild type). Mean values and standard deviations of replicate assays are shown.

25 Figure 37 shows PCR screening of transgenic ryegrass plants. PCR analysis using *npt2*-specific primers of 8 independent transgenic perennial ryegrass plants from biolistic transformation with antisense *LpUbi4CL2* vector.

Figure 38 shows the nucleotide sequence of genomic clone 4CL2 from 28 perennial ryegrass (Sequence ID No: 17).

- 29 -

Figure 39 shows the nucleotide sequence of genomic clone CCR1 from perennial ryegrass (Sequence ID No: 18).

Figure 40 shows the map location of *Lp4CL1*, *Lp4CL3*, *LpCAD1*, *LpCAD2*, *LpCCR1*, *LpOMT1* and *LpOMT2* (in bold) within the genetic linkage 5 map of perennial ryegrass.

EXAMPLE 1

Isolation and characterisation of three 4-Coumarate CoA-Ligase (4CL) cDNAs from *Lolium perenne*

Materials and Methods

10 ***Plant material***

Plants and embryogenic cell suspensions of perennial ryegrass (*Lolium perenne* L.) cv Ellet and tall fescue (*Festuca arundinacea* Schreb.) cv Triumph were established and maintained as previously described (Heath et al., 1998). Wounding experiments were performed with 10-day-old seedlings of perennial 15 ryegrass (cv Ellet) as previously described (Heath et al., 1998).

Screening of a cDNA library

A cDNA library prepared with RNA isolated from perennial ryegrass seedlings (Heath et al., 1998) was screened with a [³²P]dCTP-labelled rice partial 4CL probe. The rice 4CL probe and consisted of a 844 bp 4CL specific 20 sequence inserted into PUC119. This insert has 93 % sequence identity with a rice 4CL cDNA sequence (Genbank, L43362, bases 453-1300). cDNA inserts were excised and recircularized using the ExAssist helper phage with SOLR strain (Stratagene) as described by the manufacturer.

DNA sequencing

25 cDNA clones were digested with 8 restriction enzymes (*Bam*H_I, *Eco*R_I, *Kpn*I, *Not*I, *Pst*I, *Sall*, *Xba*I, *Xho*I) and selected clones were sequenced on both

- 30 -

strands by the dideoxy chain termination method using M13 forward and reverse primers. For sequencing the internal regions of *Lp4CL1*, *Lp4CL2* and *Lp4CL3* synthetic oligonucleotide primers were designed from the DNA sequences previously determined. Sequencing was performed using the ABI 5 dye terminator kit and automatic sequencer. Nucleotide sequences were aligned using the SeqEd program (ABI) and further analysis was performed using the HIBIO DNASIS vs2 program (Hitachi Software Engineering).

Genomic DNA blot analysis

Genomic DNA was isolated from single genotype-derived cell 10 suspensions of perennial ryegrass and tall fescue according to Lichtenstein and Draper (1985). Ten µg of perennial ryegrass DNA and 20 µg of tall fescue DNA was digested with each of the restriction enzymes *Hind*III and *Xba*I, separated on 1 % agarose gels, and transferred to Hybond N⁺ membranes according to the manufacturer's instructions (Amersham). Probes consisted of 15 *Bam*HI/*Kpn*I fragments of *Lp4CL1* (1771 bp), *Lp4CL2* (2034 bp) or *Lp4CL3* (2080 bp) labelled using the Megaprime labelling kit (Amersham) and [³²P]dCTP. Hybridization was performed at 65 °C in 5 X SSPE, 5 X Denhardt's solution, 0.5 % (w/v) SDS, and 200 µg/mL denatured herring sperm DNA. Membranes were washed three times in 2 X SSPE, 0.1 % SDS for 10 min at 20 25 °C and then twice in 0.1 X SSPE, 0.1 % SDS for 20 min at 65 °C.

RNA blot analysis

Total RNA (10 µg) was separated on 1.2 % formaldehyde gels and transferred to Hybond N (Amersham) membranes according to the manufacturers instructions. Membranes were stained with 0.2 % methylene 25 blue to confirm correct loading and transfer of RNA. Hybridisation was performed at 42 °C in 5 X SSPE, 5 X Denhart's solution, 0.5 % SDS, 50 % deionized formamide, 200 µg/mL denatured herring sperm DNA. Preparation of probes and washing of membranes was as for DNA blot analysis except for the tall fescue Northern blot when the final two washes were performed with 30 0.1 X SSPE, 0.1 % SDS for 10 min at 42°C.

Results

Isolation and sequence analysis of perennial ryegrass 4CL cDNAs

A cDNA library prepared from RNA extracted from perennial ryegrass seedlings was screened with a rice 4CL hybridization probe and ten cDNAs 5 were isolated from 2×10^5 pfu. The cDNAs were characterised by restriction analysis with 8 restriction enzymes. All clones were full length (approximately 2.0-2.2 kb) with poly(A) tails and could be separated into three groups: *Lp4CL1* (four clones) *Lp4CL2* (five clones) and *Lp4CL3* (one clone). Plasmid maps for 10 *Lp4CL1*, *Lp4CL2* and *Lp4CL3* are shown (Figure 1). *Lp4CL1*, *Lp4CL2* and *Lp4CL3* were fully sequenced (Figures 2, 3 and 4, respectively).

Lp4CL1 is 2284 bp long with an open reading frame (ORF) of 1710 bp, a 5' noncoding region of 322 bp and a 3' noncoding region of 252 bp including a poly(A) tail. *Lp4CL2* is 1992 bp long with an ORF of 1668 bp, a 5' noncoding region of 61 bp and a 3' noncoding region of 263 bp including a poly(A) tail. 15 *Lp4CL3* is 2038 bp long with an ORF of 1671 bp, a 5' noncoding region of 112 bp and a 3' noncoding region of 255 bp including a poly(A) tail.

Within the coding region, *Lp4CL1* has 70 % nucleic acid sequence identity with both *Lp4CL2* and *Lp4CL3*, while *Lp4CL2* has 79 % sequence identity with *Lp4CL3*. There is little sequence homology in the 3' noncoding 20 regions between clones (52-55 %).

Amino acid sequence comparisons

The putative proteins encoded by the three cDNAs consist of 570 amino acids [60290 u (Da)] for *Lp4CL1*, 556 amino acids (59238 u) for *Lp4CL2* and 557 amino acids (59735 u) for *Lp4CL3*. The deduced amino acid sequences of 25 *Lp4CL1*, *Lp4CL2* and *Lp4CL3* are shown (Figure 5). *Lp4CL2* and *Lp4CL3* share 79 % amino acid sequence identity, *Lp4CL1* and *Lp4CL2* have 61 % amino acid sequence identity, while *Lp4CL1* and *Lp4CL3* have only 58 % amino acid sequence identity. Regions of high sequence homology are more prevalent in the central and c-terminal regions of the enzyme. For example the

- 32 -

sequence identity between amino acids 208 to 568 of each enzyme is 85 % for *Lp4CL2* and *Lp4CL3*, 72 % for *Lp4CL1* and *Lp4CL2* and 67 % for *Lp4CL1* and *Lp4CL3*.

5 *Lp4CL1*, *Lp4CL2* and *Lp4CL3* share several common regions with other plant 4CLs. In particular, they contain the putative AMP-binding domain and the conserved GEICIRG motif, except for *Lp4CL3* where the second isoleucine has been replaced with valine (Figure 5). It has been proposed that domain II is associated with the catalytic activity of 4CL. Also, four Cys residues conserved in plant 4CLs are conserved in *Lp4CL1*, *Lp4CL2* and *Lp4CL3*

10 (Figure 5). These results suggest that the *L. perenne* cDNAs encode three divergent 4CL enzymes that are likely to have originated from three different 4CL genes.

Expression of perennial ryegrass 4CL genes

15 *Lp4CL1*, *Lp4CL2* and *Lp4CL3* were used as hybridization probes in Northern blots with RNA prepared from different organs of perennial ryegrass at two developmental stages. All three probes hybridized to a single mRNA species of approximately 2.2 - 2.3 kb. *Lp4CL1*, *Lp4CL2* and *Lp4CL3* were expressed at both seedling and mature stages of development and in all organs tested. For *Lp4CL2* and *Lp4CL3* the strongest signal was found in RNA 20 samples from seedling roots and mature stems (Figure 6).

25 *Lp4CL1*, *Lp4CL2* and *Lp4CL3* were also used as hybridization probes in Northern blots with RNA prepared from tall fescue. All three probes hybridized to a similar mRNA species (2.3 kb) as that in perennial ryegrass (Figure 6). The strongest signal was found in RNA samples from mature stems with weaker signals in RNA from roots and seedling shoots. No expression of *Lp4CL1*, *Lp4CL2* or *Lp4CL3* was observed in leaves. The three probes varied in their ability to hybridize to the corresponding homologues in tall fescue, with *Lp4CL3* resulting in the highest signal and *Lp4CL1* hybridizing only weakly.

To determine whether 4CL could be induced under stress conditions,

- 33 -

leaves of perennial ryegrass seedlings were wounded. No increase in the transcript level upon wounding was observed with *Lp4CL1*, *Lp4CL2* or *Lp4CL3* (Figure 7).

Genomic organization of perennial ryegrass 4CL genes

5 Perennial ryegrass DNA was digested with two restriction enzymes, *Hind*III or *Xba*I. Restriction sites for these enzymes are not present in the cDNA sequence of *Lp4CL1*, *Lp4CL2* or *Lp4CL3*. When *Lp4CL1*, *Lp4CL2* or *Lp4CL3* was used as a probe, several DNA hybridizing fragments of varying intensity were revealed (Figure 8). Each probe hybridized to a unique set of
10 fragments, suggesting that *Lp4CL1*, *Lp4CL2* and *Lp4CL3* represent three different genes. Furthermore, *Lp4CL1* and *Lp4CL2* hybridized to 2 to 3 major fragments per digest which may represent either alleles of the same gene or indicate the presence of more than one gene in each class. The *Lp4CL1*,
15 *Lp4CL2* and *Lp4CL3* probes also revealed several different size hybridizing DNA fragments in genomic Southern blots from tall fescue under high stringency conditions (Figure 8), suggesting that three similar 4CL genes are present in *F. arundinacea*.

EXAMPLE 2

Isolation and characterisation of a Cinnamoyl CoA Reductase (CCR)

20 ***cDNA from *Lolium perenne****

A total of 500,000 phage were screened from a cDNA library constructed from ten-day-old etiolated *L. perenne* seedlings using a maize CCR probe. Ninety-three positive plaques were observed in the primary screen and five were subsequently analysed by restriction enzyme digestion.
25 Four out of the five were identical. One of the four identical cDNAs, *LpCCR1*, was selected for further analysis (Figure 9).

Nucleic acid sequence analysis of perennial ryegrass CCR cDNA

The full nucleotide sequence of *LpCCR1* was obtained and the amino

- 34 -

acid sequence predicted (Figure 10). *LpCCR1* is a 1395 bp cDNA with 149 bp of 5' non-coding region and 160 bp of 3' non-coding region. An open reading frame of 1086 bp encodes a protein of 362 amino acids. The composition of the coding region was found to be 68% G+C rich. Codon usage was also
5 examined and found to be biased towards XXC/G codons (94%), with XCG and XUA codons accounting for only 9% and 0.55% respectively. G+C richness and bias towards G and C in the third position of a codon triplet are previously reported characteristics of monocot genes.

Genomic organization of perennial ryegrass CCR gene

10 The number of CCR genes present in the ryegrass genome was determined by Southern blot analysis of genomic DNA from double haploid plants, using as probe a fragment of the *LpCCR1* cDNA (LpCCR531, Figure 9). Double haploid DNA reduces the complexity associated with allelic variation. Genomic DNA was cut with enzymes that do not cut the cDNA
15 internally; *Dra*I, *Bam*HI, *Eco*RI, *Eco*RV, *Hind*III and *Xba*I, and the membrane was hybridised and washed under medium-stringency conditions. A single strongly hybridising band was evident in each lane (Figure 11) indicating that there is a single copy of the *LpCCR1* gene in the perennial ryegrass genome.

Expression of perennial ryegrass CCR gene

20 To investigate the expression profile of the CCR gene in ryegrass, northern hybridisation analysis was carried out with total RNA extracted from roots and shoots at seedling growth stages (0.5-1cm and 4-6cm shoots) and roots, stem and leaves at mature growth stages (6 and 10 weeks). Seedlings were grown on filter paper in the dark at 25°C and then transferred to soil and
25 glasshouse conditions (25°C) until the 6 and 10-week stages. Whole seedling total RNA from *Festuca* and *Phalaris* was included in the northern analysis. Hybridisation with *LpCCR531* (Figure 9) was performed at medium-stringency and the membrane was then washed at high-stringency. A transcript of approximately 1.5 kb was detected in all tissues, the level of expression
30 varying with maturity and from one tissue type to another (Figure 12). The *LpCCR1* transcript appears to be more abundant in roots and stem than

- 35 -

shoots and leaves. In the stem, transcript abundance increases from 6-weeks to 10-weeks; indicating that transcription in stem tissue is up-regulated as the plant matures. Expression was found predominantly in tissues such as stems and roots that are forming secondary cell walls indicating that *LpCCR1* is 5 constitutively involved in lignification.

EXAMPLE 3

Isolation and characterisation of Cinnamyl Alcohol Dehydrogenase (CAD) cDNAs from *Lolium perenne*

A 558 bp cinnamyl alcohol dehydrogenase (CAD) fragment was 10 amplified from cDNA synthesised from total RNA prepared from perennial ryegrass seedlings. The conserved amino acid domains between *Pinus radiata*, *Medicago sativa*, *Aralia cordata*, *Eucalyptus botryoides* and *Arabidopsis thaliana* CADs were used to design oligonucleotides for the 15 amplification of the perennial ryegrass CAD. The forward oligonucleotide was designed to the conserved amino acid domain CAGVTVYS and the reverse oligonucleotide to the conserved domain DVRYRFV. The 551 bp PCR fragment was cloned and sequenced to confirm that it corresponded to a perennial ryegrass CAD PCR fragment. A cDNA library prepared from RNA extracted from perennial ryegrass seedlings was screened with the 551bp 20 PCR fragment specific for perennial ryegrass CAD. Eight cDNAs were isolated and separated into six groups by restriction digest analysis. One representative clone each from two groups (*LpCAD1*, *LpCAD2*) were selected for further characterisation.

Nucleic acid sequence analysis of perennial ryegrass CAD cDNAs

25 The complete sequence of the perennial ryegrass CAD homologue *LpCAD1* was determined (Figure 13). The 1325 bp clone had a poly (A) tail, typical start and stop codons and the open reading frame (ORF) of this clone coded for a putative protein of 408 amino acids.

- 36 -

The complete nucleotide sequence of the perennial ryegrass CAD homologue *LpCAD2* was also determined (Figure 14).

Expression of perennial ryegrass CAD genes

A northern hybridisation analysis with RNA samples isolated from 5 perennial ryegrass at different developmental stages hybridised with the full length *LpCAD1* 1325 bp cDNA (Figure 15) was performed to determine patterns of organ and developmental expression. The probe hybridised to a single mRNA species of approximately 1.6 kb. The *LpCAD1* transcript was expressed in all tissue tested: roots, shoots, stem and leaves (Figure 16A). 10 The *LpCAD1* transcript was most abundant in root tissue and the mature stem, this expression pattern is typical of a gene involved in the lignification of plant cell walls. Intergeneric homologies were revealed in *Festuca* and *Phalaris*.

A similar northern hybridisation analysis was performed with *LpCAD2* (Figure 16B), however the transcript was found to be most abundant in mature 15 stem tissue and the shoots.

Genomic organization of perennial ryegrass CAD genes

A Southern hybridisation analysis using DNA samples isolated from a perennial ryegrass double haploid plant digested with *Dra*I, *Bam*HI, *Eco*RI, *Eco*RV, *Hind*III and *Xba*I and hybridised with a 500 bp *LpCAD1* probe was 20 performed. The hybridisation pattern at high stringency revealed the presence of two prominent bands for most digests indicating that *LpCAD1* belongs to a small gene family and exists a multicity gene in perennial ryegrass (Figure 17A).

A similar Southern hybridization analysis was performed with *LpCAD2* 25 (Figure 17B) the hybridisation pattern at high stringency revealed the presence of one or two prominent bands for most digests indicating that *LpCAD2* exists as a single copy gene or a member of a small gene family in perennial ryegrass (Figure 17B).

EXAMPLE 4

Isolation and characterisation of genomic clones and promoters for *O*-methyltransferase (OMT), cinnamoyl-CoA reductase (CCR), 4 coumarate CoA-ligase (4CL) and cinnamyl alcohol dehydrogenase (CAD) from

5

Lolium perenne

Genomic clones and promoters of *O*-methyltransferase (OMT), cinnamoyl-CoA reductase (CCR), 4 coumarate CoA-ligase (4CL) and cinnamyl alcohol dehydrogenase (CAD) were isolated from a perennial ryegrass genomic library using the corresponding cDNAs as hybridisation probes.

10 ***Isolation and characterisation of genomic clones and promoters for perennial ryegrass O-methyltransferase (OMT)***

A perennial ryegrass genomic library was screened with the cDNA clone, *LpOmt1*, (Heath *et al.* 1998) encoding *O*-methyltransferase (OMT). The sequence of the 5' untranslated region and the coding region was found to be 15 identical to that of the *LpOmt1* cDNA previously isolated. The entire 4.8 kb genomic clone was fully sequenced (Figure 18).

To further characterise the promoters, transcriptional fusions of the promoter sequence to the β -glucuronidase (GUS) coding sequence (*gusA*) 20 have been generated (Figure 19). Direct gene transfer experiments to tobacco protoplasts were performed with the corresponding chimeric genes to transgenically express them in a heterologous system for *in planta* expression 25 pattern analysis by histochemical GUS assays. A set of transgenic tobacco plants carrying a chimeric *gusA* gene under the control of the 5' regulatory region of the *LpOmt1* promoter was generated to assess the potential use of the *LpOmt1* promoter for xylem-specificity and targeted downregulation of genes encoding key lignin biosynthetic enzymes.

The transgenic tobacco plants generated using the *LpOmt1* promoter driven chimeric *gusA* transformation vector were screened by PCR and histochemical GUS assays.

A PCR screening was undertaken using *gusA* specific primers for the initial identification of transgenic tobacco plants (Figure 20). PCR positive tobacco plants were screened by histochemical GUS assays for *in planta* expression pattern analysis (Figure 20).

5 ***Isolation and characterisation of genomic clones and promoters for perennial ryegrass cinnamoyl-CoA reductase (CCR)***

A CCR genomic clone from perennial ryegrass was isolated containing 6.5 kb of promoter and the entire gene organisation (intron/exon boundaries). The CCR promoter can be used for targeted expression of foreign genes in 10 transgenic plants.

A perennial ryegrass genomic library was screened with the cDNA clone *LpCCR1* which codes for the lignin biosynthetic enzyme, cinnamyl-CoA reductase (CCR). Four different genomic clones were identified based on restriction digest analysis. Clone 6.1.1a was selected for further analysis. A 15 6.42 kb *Xhol* fragment from clone 6.1.1a, which hybridized strongly to the *LpCCR1* cDNA probe, was subcloned into pBluescriptSK (Figure 21A). Sequence analysis revealed that the 6.42 kb *Xhol* fragment contained the entire *LpCCR1* gene and 200 bp of promoter region. The intron/exon boundaries are illustrated in figure 21B, the location and the size of the exons 20 appear to be conserved in other CCRs from different species (Figure 21C).

To isolate the promoter region of *LpCCR1*, the Southern blot containing digested phage genomic DNA isolated from clone λ Lp6.1.1a was reprobed with the 200bp promoter region. The probe hybridized strongly to a 6.5 kb *Sall* fragment. This genomic fragment *LpCCR1* clone 2, was subcloned into 25 pBluescriptSK and sequenced (Figure 22A). Sequence results revealed that the 6.5 kb *Sall* fragment contained 6.5 kb of promoter (Figure 22B). The full sequence of *LpCCR1* genomic clone containing the promoter and entire gene sequence (exons and introns) was obtained and is shown on Figure 39.

Isolation and characterisation of genomic clones and promoters for perennial ryegrass 4 coumarate CoA-ligase (4CL)

A 4CL2 genomic clone from perennial ryegrass was isolated containing 2.5 kb of promoter and partial gene organisation (intron/exon boundaries). The 5 4CL2 promoter can be used for targeted expression of foreign genes in transgenic plants. The 2.5 kb promoter has been fused to the reporter gene *gusA* for expression analysis.

A perennial ryegrass genomic library was screened with an *Lp4CL* cDNA probe. After tertiary screening positive 4CL genomic clones were 10 obtained and characterised by restriction digest and Southern hybridisation analysis (Figure 23A).

Sequence analysis revealed that the isolated 4CL genomic clone (4CL2) from perennial ryegrass had 100% nucleotide identity to the *Lp4CL2* cDNA clone. To further characterise this 5 kb λ *Lp4CL2* genomic clone and to 15 confirm that it corresponds to the cDNA of *Lp4CL2*, a number of PCR reactions using primers designed to the cDNA were used. PCR results confirmed that the 5 kb genomic fragment was a partial genomic clone corresponding to the *Lp4CL2* cDNA (Figure 23B). Using primer combinations F1 and R1 the entire 4.8kb genomic fragment was amplified. To determine the 20 location of introns additional PCR reactions using the primer combinations F1 / R2 and F2 / R1 were performed, a 1 kb and 3.5 kb bands were amplified respectively. The location and size of the introns could be determined from these results, and further confirmed by sequence analysis. This large 5 kb genomic fragment contains 4 small exons representing the coding sequence of 25 *Lp4CL2* between 508 bp and 1490 bp (Figure 23C).

The genomic clone 1, *Lp4CL2* contained no promoter region. To isolate the promoter region of *Lp4CL2*, the Southern blot containing digested phage genomic DNA isolated from clone λ *Lp4CL2* was reprobed with a 300 bp EcoRI/BgII isolated from the 5' end of the cDNA clone *Lp4CL2*. The 300 bp 30 probe hybridised strongly to a 2.5 kb BamHI fragment. This genomic fragment

- 40 -

5 *Lp4CL2* clone 2, was subcloned into pBluescriptSK and sequenced (Figure 24A). Sequence results revealed that the 2.5 kb *Bam*HI fragment contained the 508 bp of the 5' ORF of *Lp4CL2* missing from genomic clone 1 and 2.0 kb of promoter region (Figure 24B). The full sequence of the *Lp4CL2* genomic clone containing the promoter and partial gene sequence (exons and introns) was obtained and is shown on Figure 39.

The promoter from *Lp4CL2* was thus isolated and used for the production of a chimeric *gusA* reporter gene (Figure 25).

10 ***Isolation and characterisation of genomic clones and promoters for perennial ryegrass cinnamyl alcohol dehydrogenase (CAD)***

A CAD genomic clone from perennial ryegrass was isolated containing the gene organisation (intron/exon boundaries) minus intron 1 containing the first 111 bp of the CAD coding region. The genomic clone has allowed the identification of a G at position 851 bp in the coding region of the CAD2 15 genomic clone isolated from perennial ryegrass cv. Barlano which is absent in the CAD2 cDNA clone isolated from perennial ryegrass cv. Ellett. The SNP (single nucleotide polymorphism) found to exist between the 2 cultivars has the potential utility as a molecular marker for herbage quality, dry matter digestibility, mechanical stress tolerance, disease resistance, insect pest 20 resistance, plant stature and leaf and stem colour.

25  Results below show the isolation of the genomic clone and sequence analysis of deduced coding sequence from the genomic clone CAD2 from perennial ryegrass cv. Barlano compared to the truncated cDNA CAD2 from the cv Ellett. The missing G in the perennial ryegrass cv. Ellett has been highlighted (Figures 26 and 27).

A perennial ryegrass genomic library was screened with a probe corresponding to the 5' end of the *LpCAD2* cDNA clone, which codes for the lignin biosynthetic enzyme cinnamyl alcohol dehydrogenase. Ten positive plaques were identified and isolated in the primary library screening. After a 30 secondary and tertiary screening, two positive plaques were obtained and

- 41 -

corresponding positive genomic clones were further characterised by restriction digest and Southern hybridization analyses. Both genomic clones were found to be identical based on restriction digest analyses. One clone, named $\lambda LpCAD2$ was chosen for further Southern hybridization analyses. A 5 4.5 kb *Bam*HI fragment which hybridized strongly to the *LpCAD2* cDNA probe was subcloned into pBluescriptSK and sequenced (Figure 28A). Sequence analysis revealed that the 4.5 kb *Bam*HI fragment was a partial genomic clone of *LpCAD2*. This large 4.5 kb genomic fragment contains 4 small exons representing the coding sequence of *LpCAD2* between 213 bp and the stop 10 codon at 1213 bp, and the location of the intron/exon boundaries are illustrated in Figure 28B.

EXAMPLE 5

Development of transformation vectors containing chimeric genes with 4CL, CCR and CAD cDNA sequences from perennial ryegrass

15 To alter the expression of the key enzymes involved in lignin biosynthesis 4CL, CCR and CAD, through antisense and/or sense suppression technology and for over-expression of these key enzymes in transgenic plants, a set of sense and antisense transformation vectors was produced. Transformation vectors containing chimeric genes using perennial ryegrass 20 4CL, CCR and CAD cDNAs in sense and antisense orientations under the control of either the CaMV 35S or the maize ubiquitin promoter were generated (Figures 29, 30 and 31).

EXAMPLE 6

Production and characterisation of transgenic tobacco plants expressing 25 chimeric 4CL, CCR and CAD genes from perennial ryegrass

A set of transgenic tobacco plants carrying chimeric 4CL, CCR and CAD genes from perennial ryegrass were produced and analysed.

- 42 -

Transformation vectors with *Lp4CL1*, *Lp4CL2* and *Lp4CL3* full length cDNA sequences in sense and antisense orientations under the control of either the CaMV 35S or the maize ubiquitin promoters were generated. Transformation vectors with *LpCCR1* cDNA in both sense and antisense orientation under the control of either the CaMV 35S and maize ubiquitin promoters were generated. Transformation vectors with 1325 bp full length *LpCAD1* cDNA in sense and 1051 bp partial *LpCAD1* cDNA in antisense orientation under the control of either the CaMV 35S and maize ubiquitin promoters were generated.

10 Direct gene transfer experiments to tobacco protoplasts were performed using these transformation vectors.

The production and molecular analysis of transgenic tobacco plants carrying the perennial ryegrass *Lp4CL1* and *LpCCR1* cDNAs under the control of the constitutive CaMV 35S promoter is described here in detail.

15 A set of transgenic tobacco plants generated using the *Lp4CL1* sense transformation vector was screened by PCR and subjected to Southern and northern hybridization analyses.

20 A PCR screening was undertaken using *npt2* and *Lp4CL1* specific primers for the initial identification of transgenic tobacco plants. Independent transgenic tobacco plants were identified to be co-transformed with both the selectable marker *npt2* and the *Lp4CL1* chimeric genes (Figure 32).

25 Southern hybridisation analysis was performed with DNA samples from PCR positive transgenic tobacco plants to demonstrate the integration of the chimeric *Lp4CL1* transgene in the tobacco plant genome. Independent transgenic tobacco plants carried between 1 and 5 copies of the *Lp4CL1* transgene. No cross-hybridization was observed between the endogenous tobacco 4CL gene and the perennial ryegrass hybridization probe used (Figure

- 43 -

32).

Northern hybridization analysis using total RNA samples prepared from the transgenic tobacco plants carrying the chimeric sense *Lp4CL1* transgene and probed with the *Lp4CL1*-specific hybridization probe revealed the 5 presence of a 1.2 kb *Lp4CL1* transcript strongly expressed in one *Lp4CL1*-transgenic tobacco plant analysed (Figure 32).

The sense and antisense transformation vectors of *LpCCR1* under the control of the CaMV 35S promoter were introduced into tobacco protoplasts via direct gene transfer. A set of transgenic tobacco plants was generated and 10 screened by PCR with specific primers to identify transgenic tobacco plants carrying chimeric *LpCCR1* transgene. The molecular analysis of *LpCCR1*-transgenic tobacco plants is shown (Figure 33).

EXAMPLE 7

Production and characterisation of transgenic perennial ryegrass 15 plants expressing chimeric OMT, 4CL, CCR and CAD genes from perennial ryegrass

An improved transformation method was developed for the production of transgenic perennial ryegrass plants by biolistic transformation of embryogenic cells. Transgenic perennial ryegrass plants were generated using 20 chimeric OMT, 4CL, CCR and CAD genes from perennial ryegrass and the improved transformation method.

Improved method for the production of transgenic perennial ryegrass plants

This improved procedure utilises embryogenic calli produced from 25 mature seed-derived embryos as direct targets for biolistic transformation without requiring the establishment of embryogenic cell suspensions. The protocol relies on a continuous supply of isolated zygotic embryos for callus induction. Transgenic ryegrass plants can be regenerated 24 – 28 weeks after

embryo isolation (Fig. 34). Isolated embryos are plated onto MSM5 medium to produce embryogenic calli suitable as targets for biolistic transformation within 8 weeks. The embryogenic calli, treated on high-osmoticum medium MSM3 Plus prior to microprojectile bombardment, are selected on MSM3 medium 5 containing 100 mg/l paromomycin (Pm) for 2 weeks before being transferred onto MSK with 100 mg/l Pm for further 4 weeks until differentiation of Pm resistant shoot appear. Regenerated shoots are transferred on to fresh selective media MSK with 100 mg/l Pm for a further 4 weeks (Figure 34).

10 ***Production of transgenic perennial ryegrass plants expressing chimeric OMT, 4CL, CCR and CAD genes from perennial ryegrass***

Transgenic perennial ryegrass (*Lolium perenne*) plants were generated using chimeric ryegrass OMT, 4CL, CCR and CAD genes by biolistic transformation of embryogenic calli. Examples of the production and detailed molecular analysis of these transgenic ryegrass plants are described.

15 Transgenic perennial ryegrass plants for OMT down-regulation were produced using biolistic transformation of embryogenic calli and plant transformation vectors pUbiomt1 and pUbitmo1 carrying *LpOmt1* cDNA sequence in sense and antisense orientation under control of the constitutive maize ubiquitin promoter. These transgenic perennial ryegrass plants for 20 down-regulated OMT activity were regenerated from paromomycin resistant calli obtained from biolistic transformation using microprojectiles coated with two plasmids; pHp23 (carrying the chimeric *npt2* gene as the selectable marker) and either the sense or antisense *LpOmt1* transformation vector driven by the maize *Ubi* promoter.

25 Transgenic perennial ryegrass plants were subjected to a polymerase chain reaction (PCR) screening using *npt2*-specific primers. Independent *npt2* PCR-positive transgenic perennial ryegrass plants obtained from biolistic transformation of embryogenic calli – generated from approximately 60,000 isolated mature seed-derived embryos - using *LpOmt1* sense (pUbiomt1) and 30 *LpOmt1* antisense (pUbitmo1) transformation vectors were identified [16 pUbiomt1 transgenic plants and 27 pUbitmo1 transgenic plants] (Figure 35).

- 45 -

Southern hybridization analysis was performed with undigested and *Hind*III-digested DNA samples prepared from the PCR positive transgenic perennial ryegrass plants, to demonstrate their transgenic nature and the integration of the chimeric *npt2* and *LpOmt1* transgenes. Independent 5 transgenic perennial ryegrass plants co-transformed with both, the selectable marker *npt2* gene and *LpOmt1* chimeric genes, were identified (Figure 35). In most instances, the transgenic perennial ryegrass plants recovered contained multiple copies of the selectable marker gene including rearranged transgene 10 copies. No *npt2*-hybridizing bands were detected in the untransformed negative control.

Samples of *Hind*III-digested genomic DNA were included in the analysis when the *LpOmt1* gene-specific hybridization probe (*omt1*) was used. The *omt1* probe hybridized to a number of bands in DNA samples corresponding to both, the transgenic plants and the untransformed negative control. The *omt1*- 15 hybridizing bands shared in all samples correspond to endogenous *LpOmt1* gene sequences represented as a small multigene family in the perennial ryegrass genome (Heath et al. 1998). The different *omt1*-hybridizing bands evident in the samples from the transgenic plants and absent in the untransformed negative control sample correspond to antisense (*tmo1*) and 20 sense (*omt1*) *LpOmt1* transgene integration events (Figure 35).

Northern hybridization analysis using strand-specific *LpOmt1* probes allowed the identification of transgenic perennial ryegrass plants expressing the antisense *LpOmt1* transgene (Figure 35).

The OMT activity of selected antisense and sense *LpOmt1* transgenic 25 perennial ryegrass plants was determined. Biochemical assays for OMT activity were initially established in untransformed plants (such as tobacco and perennial ryegrass). The assays utilise radiolabelled S-adenosylmethionine as the methyl donor for the OMT-catalysed conversion of caffeic acid into ferulic acid. The production of radioactive ferulic acid is measured and allows the 30 OMT activity to be determined.

- 46 -

The OMT activity of selected *LpOmt1*-transgenic perennial ryegrass plants (*L. perenne* cv. Ellett) was determined. Significantly altered OMT activity in individual transformation events was observed (Figure 36). The manipulation of OMT activity in transgenic perennial ryegrass plants due to the 5 expression of the chimeric ryegrass *LpOmt1* gene was thus demonstrated.

Transgenic perennial ryegrass plants were recovered, using biolistic transformation of embryogenic calli, for the manipulation of the expression of genes encoding the key lignin biosynthetic enzyme, 4CL. The plant transformation vectors pUbi4CL2 and pUbi2LC4 carrying chimeric *Lp4CL2* 10 cDNA sequences in sense and antisense orientation, respectively, driven by the constitutive maize ubiquitin (*Ubi*) promoter were used. Perennial ryegrass plants for 4CL manipulation were regenerated from Pm-resistant calli obtained from biolistic transformation of embryogenic calli using microprojectiles coated with the plasmids pHp23, carrying a chimeric *npt2* gene as selectable marker 15 gene and the antisense pUbi2LC4.

Transgenic perennial ryegrass plants were subjected to a polymerase chain reaction (PCR) screening using *npt2*-specific primers. Independent *npt2* PCR-positive transgenic perennial ryegrass plants were obtained from biolistic transformation of embryogenic calli (Figure 37).

20 Transgenic perennial ryegrass plants were also recovered, using biolistic transformation of embryogenic calli, for the manipulation of the expression of genes encoding the key lignin biosynthetic enzymes, CCR and CAD.

EXAMPLE 8

25 **Genetic mapping of perennial ryegrass OMT, 4CL, CCR and CAD genes**

Lp4CL1, *Lp4CL3*, *LpCAD1*, *LpCAD2*, *LpCCR1*, *LpOMT1* and *LpOMT2* clones were PCR amplified and radio-labelled for use as probes to detect

- 47 -

restriction fragment length polymorphisms (RFLPs). RFLPs were mapped using 110 progeny individuals of the p150/112 perennial ryegrass reference population restricted with the enzymes described in the table below.

Clones	Polymorphic in p150/112	Enzyme mapped with	Locus	Linkage group
<i>Lp4CL1</i>	Y	<i>Dra</i> l	<i>Lp4CL1</i>	2
<i>Lp4CL3</i>	Y	<i>Eco</i> RV	<i>Lp4CL3</i>	6
<i>LpCAD1</i>	Y	<i>Eco</i> RV	<i>LpCAD1</i>	2
<i>LpCAD1.2.1</i>	Y	<i>Eco</i> RI	<i>LpCAD2a</i> <i>LpCAD2b</i> <i>LpCAD2c</i>	7 - 2
<i>LpCCR1</i>	Y	<i>Eco</i> RI	<i>LpCCR1</i>	7
<i>LpOMT1</i>	Y	<i>Dra</i> l	<i>LpOMT1</i>	7
<i>LpOMT2</i>	Y	<i>Eco</i> RV	<i>LpOMT2</i>	6

5

Lp4CL1, *Lp4CL3*, *LpCAD1*, *LpCAD2*, *LpCCR1*, *LpOMT1* and *LpOMT2* loci mapped to the linkage groups as indicated in the table and in Figure 40. These gene locations can now be used as candidate genes for quantitative trait loci for lignin biosynthesis associated traits such as herbage quality, dry 10 matter digestibility, mechanical stress tolerance, disease resistance, insect pest resistance, plant stature and leaf and stem colour.

REFERENCES

Heath *et al* (1988) cDNA cloning and differential expression of three caffeic acid O-methyltransferase homologues from perennial ryegrass (*Lolium perenne*). *Journal of Plant Physiology* 153:649-657

Lichtenstein, C, And J. Draper (1985) Genetic engineering of plants. *In:* D.M. Glover (ed.), *DNA Cloning*, Vol. 2, pp. 67-119, IRL Press, Washington.

Finally, it is to be understood that various alterations, modifications and/or additions may be made without departing from the spirit of the present 20 invention as outlined herein.

- 48 -

It will also be understood that the term "comprises" (or its grammatical variants) as used in this specification is equivalent to the term "includes" and should not be taken as excluding the presence of other elements or features.

5 Documents cited in this specification are for reference purposes only and their inclusion is not an acknowledgement that they form part of the common general knowledge in the relevant art.

CLAIMS

1. A substantially purified or isolated nucleic acid or nucleic acid fragment encoding an enzyme selected from the group consisting of 4 coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR) and cinnamyl alcohol dehydrogenase (CAD), from a ryegrass (*Lolium*) or fescue (*Festuca*) species.
5
2. A nucleic acid or nucleic acid fragment according to claim 1 wherein said ryegrass or fescue species is perennial ryegrass (*Lolium perenne*).
- 10 3. A nucleic acid or nucleic acid fragment according to claim 1 encoding 4CL and including a nucleotide sequence selected from the group consisting of (a) sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 1, 3 and 5; respectively) (b) complements of the sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 1, 3 and 5, respectively); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).
15
4. A nucleic acid or nucleic acid fragment according to claim 1 encoding CCR and including a nucleotide sequence selected from the group consisting of (a) the sequence shown in Figure 10 hereto (Sequence ID No: 7); (b) the complement of the sequence shown in Figure 10 hereto (Sequence ID No: 7); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).
20
- 25 5. A nucleic acid or nucleic acid fragment according to claim 1 encoding CAD and including a nucleotide sequence selected from the group consisting of (a) the sequences shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 9, 11, 14 and 16, respectively); (b) complements of the sequences shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 9,

- 50 -

11, 14 and 16, respectively); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

6. A vector including a nucleic acid or nucleic acid fragment
5 according to claim 1.

7. A vector according to claim 6 further including a promoter and a terminator, said promoter, nucleic acid or nucleic acid fragment and terminator being operatively linked.

8. A plant cell, plant, plant seed or other plant part, including a
10 vector according to claim 6.

9. A method of modifying lignin biosynthesis in a plant, said method including introducing into said plant an effective amount of a nucleic acid or nucleic acid fragment according to claim 1 and/or a vector according to claim 6.

15 10. Use of a nucleic acid or nucleic acid fragment according to claim 1, and/or nucleotide sequence information thereof, and/or single nucleotide polymorphisms thereof as a molecular genetic marker.

11. A substantially purified or isolated polypeptide from a ryegrass (*Lolium*) or fescue (*Festuca*) species, selected from the group consisting of the
20 enzymes 4CL, CCR and CAD.

12. A polypeptide according to claim 11 wherein said ryegrass or fescue species is perennial ryegrass (*Lolium perenne*).

13. A polypeptide according to claim 11 wherein said polypeptide is 4CL and includes an amino acid sequence selected from the group consisting of sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 2, 4 and 25 6, respectively); and functionally active fragments and variants thereof.

- 51 -

14. A polypeptide according to claim 11 wherein said polypeptide is CCR and includes an amino acid sequence selected from the group consisting of the sequence shown in Figure 10 hereto (Sequence ID No: 8); and functionally active fragments and variants thereof.

5 15. A polypeptide according to claim 11 wherein said polypeptide is CAD and includes an amino acid sequence selected from the group consisting of the sequence shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 10, 12, 15 and 17, respectively); and functionally active fragments and variants thereof.

10 16. A lignin or modified lignin substantially or partially purified or isolated from a plant, plant seed or other plant part according to claim 8.

15 17. An isolated regulatory element capable of causing expression of an exogenous gene in plant cells, wherein said regulatory element is isolated from a nucleic acid or nucleic acid fragment encoding a protein selected from the group consisting of: O-methyl transferase (OMT), 4 coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR) and cinnamyl alcohol dehydrogenase (CAD).

18. A regulatory element according to claim 17 wherein said regulatory element includes an O-methyltransferase promoter.

20 19. A regulatory element according to claim 17 wherein said regulatory element includes a 4 coumarate CoA-ligase promoter.

20. A regulatory element according to claim 17 wherein said regulatory element includes a cinnamoyl CoA-reductase promoter.

25 19. A regulatory element according to claim 17 from a ryegrass (*Lolium*) or Fescue (*Festuca*) species.

20. A regulatory element according to claim 17 including the first

- 52 -

approximately 4630 nucleotides of the sequence shown in Figure 18 hereto (Sequence ID No: 13); or a functionally active fragment or variant thereof.

21. A regulatory element according to claim 17 including the first approximately 2206 nucleotides of the sequence shown in Figure 38 hereto 5 (Sequence ID No: 17); or a functionally active fragment or variant thereof.

22. A regulatory element according to claim 17 including the first approximately 6735 nucleotides of the sequence shown in Figure 39 hereto (Sequence ID No: 18); or a functionally active fragment or variant thereof.

23. A vector including a regulatory element according to claim 17.

10 24. A vector according to claim 23 further including an exogenous gene and a terminator, said regulatory element, exogenous gene and terminator being operatively linked, such that said regulatory element is capable of causing expression of said exogenous gene in plant cells.

15 25. A plant cell, plant, plant seed or other plant part, including a vector according to claim 23.

26. A method for expressing an exogenous gene in plant cells, said method including introducing into said plant cells an effective amount of a regulatory element according to claim 17 and/or a vector according to claim 23.

1/76

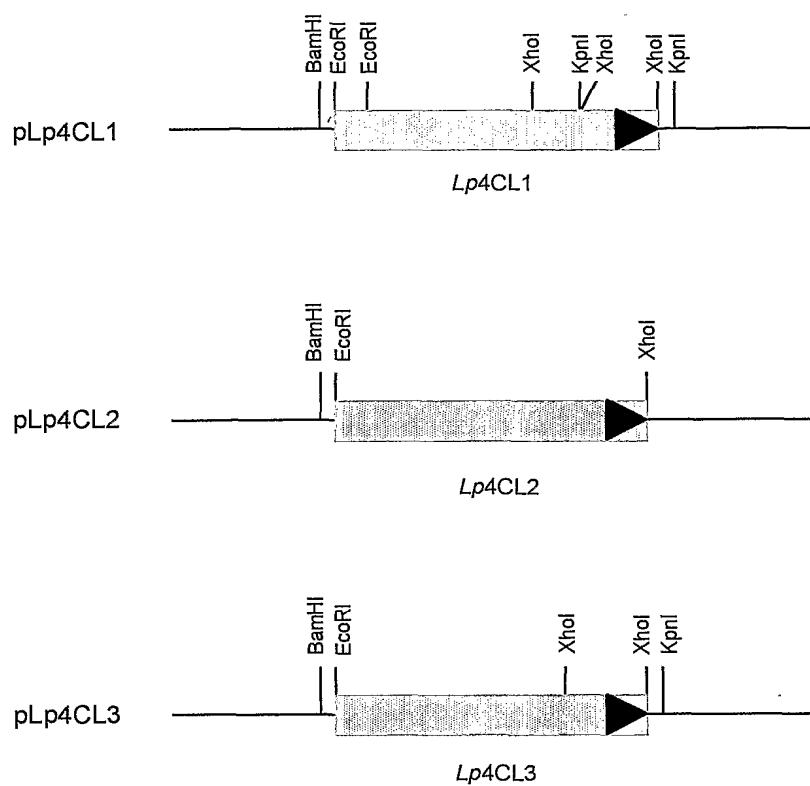


FIGURE 1

1	CGGCACGAGTGGACTTTCCGACGCCGGAGTCGCCGATGATGACCGCCTGAGGAGGTAGT	60
61	CGTAGTCGTCCCTCGCCCTGTACGCGCGCTGCCGCCATTTCCTTCCTCGCCTCGCGG	120
121	TCCTCCTCCCCGACCTGCGCTAGGCTCTGGATCTCGCGGGTTGGCGCGCGTCCCG	180
181	CTGTGAGCTCGTGCCTGAATTGGCACGCCACCTTCGAGGCGTGCAGTGGTACGAGCTC	240
241	GCGAGCCATTGTCAGTGCAGTGAGGCTCTGCTACTCGTTGCCATTCCAAGAAGCTCTC	300
301	TGCTCCCTGAAACCAGAGGATCATGATCACGGTGGCGGCCGAGGTGCAGCAGCCGA	360
	M I T V A A P E V Q Q P Q	
361	GATCGCGGGCGCTGCTGCGCCGTGGAGGCGGCACCGGAGGCACGACGATCTCCG	420
	I A A A A A V E A A A P E A T T I F R	
421	GTCCAGGCTCCGGACATCGACATCCGACCCACATGCCCTGCACGACTATTGCTTCGC	480
	S R L P D I D I P T H M P L H D Y C F A	
481	GACGGCAGCCTCGGCCCCGACGCCGTGCCTCATCACCGCGGCCACGGGGAAAGACCTA	540
	T A A S A P D A P C L I T A A T G K T Y	
541	CACGTTGCCGAGACGCACCTGCTGTGCCGCAAGGCCGGCGCTGCACGGGCTCGG	600
	T F A E T H L L C R K A A A A A L H G L G	
601	CGTGGCCACGGGGACCGGATCATGCTGCTGCCAGAACTCCGTGGAGTCGCGCTCGC	660
	V R H G D R I M L L Q N S V E F A L A	
661	CTTCCTGCCGCGTCCATGCTGCCGCGTCAGCACGGCGCGAACCGTTCTGCACGCC	720
	F F G A S M L G A V S T A A N P F C T P	
721	GCAGGAGATCCACAAGCAGCTCGTGGCTCCGGCGGAAGCTGGTCGTACGCAGTCCGC	780
	Q E I H K Q L V A S G A K L V V T Q S A	

FIGURE 2

781	CTACGTCGACAAGCTCCGGCACGAGGCCTCCCCGAATCGGCAGGGCCCTCACCGTGAT	840
	Y V D K L R H E A F P R I G E A L T V I	
841	CACCATCGACGAGGACGACGGCACCCCGAACGGCTGCCAGCCTTCTGGGCCCTCGTGTC	900
	T I D E D D G T P D G C Q P F W A L V S	
901	AGCCGCCGACGAGAACAGCGTCCCGGAGTCTCCATCTGCCGGACGACGGTGGCGCT	960
	A A D E N S V P E S P I S P D D A V A L	
961	GCCCTACTCGTCGGGCACGACGGGCTGCCAACGGCGTGGTGCACGCACGGGGCT	1020
	P Y S S G T T G L P K G V V L T H G G L	
1021	GGTGTGAGCGTGGCGCAGCAGGTGGACGGCGAGAACCGAACCTGCACATGCCGGCGGG	1080
	V S S V A Q Q V D G E N P N L H M R A G	
1081	GGAGGACGTGGTGCCTGCGTGCCTGCCACATCTCTCGCTCAACTCGGTGCT	1140
	E D V V L C V L P L F H I F S L N S V L	
1141	GCTGTGCGCGCTGCCGGCGCCGCGCCGTGATGCTGATGCCATGGTCAGATGGGGC	1200
	L C A L R A G A A V M L M P R F E M G A	
1201	CATGCTGGAGGGCATCGAGCGGTGGCGCGTCACGGTGGCGCCGTGGTGCCGCCGCTGGT	1260
	M L E G I E R W R V T V A A V V P P L V	
1261	GCTCGCGCTCGCCAAGAACCCGGGGTGGAGAACGACGACCTCAGCTCCATTGGATCGT	1320
	L A L A K N P G V E K H D L S S I R I V	
1321	GCTCTCCGGCGCCGCGCCGCTCGCAAGGAGCTCGAGGACGCCCTACGTGGCCGCCCTGCC	1380
	L S G A A P L G K E L E D A L R G R L P	
1381	GCAGGCCATCTCGGACAGGGCTACGGGATGACGGAGGCCGGCGGTGCTGTCCATGTG	1440
	Q A I F G Q G Y G M T E A G P V L S M C	
1441	CCCGCGTTCGCGCGGGAGCCGACGCCGGCAAGTCCGGCTCGTGCACCGTGGTGC	1500
	P A F A R E P T P A K S G S C G T V V R	

FIGURE 2 CONTINUED

1501	CAACGCCAGCTCAAGGTGGTCGACCCGACACCGCGTCTCCCTCGGCCGCAACCTCCC -----+-----+-----+-----+-----+-----+-----+-----+ N A Q L K V V D P D T G V S L G R N L P	1560
1561	CGCGAGATCTGCATCCGGCCCGCAGATCATGAAAGGATACTTGAATGATCCGTGGC -----+-----+-----+-----+-----+-----+-----+ G E I C I R G P Q I M K G Y L N D P V A	1620
1621	CACGGCCGCGACCATCGACGTCGAGGGTGGCTCCACACCGCGACATCGGCTACGTCGA -----+-----+-----+-----+-----+-----+-----+ T A A T I D V E G W L H T G D I G Y V D	1680
1681	CGACGACGAGGTCTTCATCGTCGACCGCGTCAAGGAGCTCATCAAGTTCAAGGGCTT -----+-----+-----+-----+-----+-----+-----+ D D D E V F I V D R V K E L I K F K G F	1740
1741	CCAGGTACCGCCGGCCGAGCTCGAGGCTCTGCTCATCGCGATCCGTCCATGCCGACGC -----+-----+-----+-----+-----+-----+-----+ Q V P P A E L E A L L I A H P S I A D A	1800
1801	GGCCGTCGTCCCGCAAAAGGATGATGCCGCCGGCGAGGTCCCGTTGCCTCGTGGTCCG -----+-----+-----+-----+-----+-----+-----+ A V V P Q K D D A A G E V P V A F V V R	1860
1861	CGCCGCCACTCCGACATGCCGAGGAGGCCATCAAGGAGTTCTATCCAAGCAGGTGGT -----+-----+-----+-----+-----+-----+-----+ A A D S D I A E E A I K E F V S K Q V V	1920
1921	GTTCTACAAGAGGCTGCACAAGGTCTACTTCACCCACGCGATAACCAAGTCGGCGTCGGG -----+-----+-----+-----+-----+-----+-----+ F Y K R L H K V Y F T H A I P K S A S G	1980
1981	GAAGATACTCAGGAAAGAACTCAGAGCTAAACTCGCCGCCGGCCACTGCCTGAAGAGT -----+-----+-----+-----+-----+-----+-----+ K I L R K E L R A K L A A P A T A * R V	2040
2041	GGTCATGGCTTCATGCTAACATTCGATCAGAAAGGCACTCTAGCATATATGTTCCA -----+-----+-----+-----+-----+-----+-----+ V H G F M L I I S I R K A L L A Y M F H	2100
2101	CCTTTGTTTCATGGAAAGATTGTATTCCAGCTAGTGGCCAGTGACTGAGTAAGGGATG -----+-----+-----+-----+-----+-----+-----+ L L F H L E D C I P A S G Q *	2160
2161	GGGATAAAAGTTTGTCTACGTTTCTTTACGCTACTCTCCATTGGGAGTACAATG -----+-----+-----+-----+-----+-----+-----+ -----	2220
2221	TATCAGGGATTCTGTATTGAAGTTAACAGATTGGTTCAATTATAAAAAAAAAAAAAA -----+-----+-----+-----+-----+-----+-----+ -----	2280
2281	AAAA 2284	

FIGURE 2 CONTINUED

1	CGGCACGAGCGCCATTCCCTCACCTTCAGCTCCGGCCAAAGATTCCATCCGGCGAGATC	60
61	CATGGGCTCCATCGCGCGGACGCGCCTCCCGGGAGCTGGTGTCCGGTCCAAGCTCCC M G S I A A D A P P A E L V F R S K L P	120
121	GGACATCGAGATCCCGACCCACCTGACGCTGCAGGACTACTGCTTCCAGCGCCTGCCGGA D I E I P T H L T L Q D Y C F Q R L P E	180
181	GCTCTCCGCGCGCCTGCCTCATCGACGGCGCCACGGGCGCCGCTCACCTACGGCGA L S A R A C L I D G A T G A A L T Y G E	240
241	GGTGGACGCCCTGTCCC GCCGCTGCGCCGCGGGCTGCGCCGCCCTGGCGTCGGCAAGGG V D A L S R R C A A G L R R L G V G K G	300
301	CGACGT CGTCATGGCGCTCCCGCAACTGCCCGAGTTCGCCCTCGTGTTCCTCGGC D V V M A L L R N C P E F A F V F L G A	360
361	GGCCCGGCTCGGCGCCGCCACCACCGCCAACCCGTTCTACACGCCAACGAGATCCA A R L G A A T T T A N P F Y T P H E I H	420
421	CCGCCAGGCCACCGCCGCCGGGCCAGGGTCATCGTCACCGAGGGCTGCGCCGTCGAGAA R Q A T A A G A R V I V T E A C A V E K	480
481	GGTGC GCGCCTTCGCCGCCAGAGAGAGGGATTCCCGTCGTCTCCGTCGACGAGGGCGTCGA V R A F A A E R G I P V V S V D E G V D	540
541	CGGGCGCTGCCTCCGTTGCCGAGACTCTGCTCGGGGAAGAAAGCGGGAGCGGTTCGT G G C L P F A E T L L G E E S G E R F V	600
601	CGACGAGGCCGGTCGACCCGACGACGTGGTGGCGCTGCCGTACTCGTCCGGCACCCGG D E A V D P D D V V A L P Y S S S G T T G	660
661	CCTGCCCAAGGGCGTCATGCTCACCCACCGCAGCCTCGTCACCAGCGTCGCCAGCAGGT L P K G V M L T H R S L V T S V A Q Q V	720
721	GGACGGTGAGAACCGAACCTGCACTTCAGCTCGTCGGACGTGCTGCTGTGCGTGCTGCC D G E N P N L H F S S S D V L L C V L P	780

FIGURE 3

781	GCTGTTCCACATCTACTCGCTCAACTCGGTGCTGCTCGCCGGTCTCCGCGCCGGGTGC	840
	-+-----+-----+-----+-----+-----+-----+-----+-----+-----+	
	L F H I Y S L N S V L L A G L R A G C A	
841	GATCGTGATCATGCGCAAGTTGACCGACGGCGCGCTGGTGGACCTGGTGCGCACGCACGG	900
	-+-----+-----+-----+-----+-----+-----+-----+-----+-----+	
	I V I M R K F D H G A L V D L V R T H G	
901	CGTCACCGTGGCGCCATTGCGGCCATCGTGGTGGAGATCGCCAAGAGCGCGCGGGT	960
	-+-----+-----+-----+-----+-----+-----+-----+-----+-----+	
	V T V A P F V P P I V V E I A K S A R V	
961	GACCGCCGCGGACCTGGCGTCCATCCGGCTGGTCATGTCGGGGCGGCCATGGCAA	1020
	-+-----+-----+-----+-----+-----+-----+-----+-----+-----+	
	T A A D L A S I R L V M S G A A P M G K	
1021	GGAGCTGCAGGACCGCGTTCATGGCCAAGATCCCCAACGCCGTGCTCGGCCAGGGATATGG	1080
	-+-----+-----+-----+-----+-----+-----+-----+-----+-----+	
	E L Q D A F M A K I P N A V L G Q G Y G	
1081	GATGACCGAGGCCGGCCCTGTGCTGGCGATGTCCTGGCTTCGCCAAGGAGGCCGTTCGC	1140
	-+-----+-----+-----+-----+-----+-----+-----+-----+-----+	
	M T E A G P V L A M C L A F A K E P F A	
1141	GGTCAAGTCCGGTTCTGGCACCCTCGTCAGGAACGCCGAGCTCAAGATCGTCGACCC	1200
	-+-----+-----+-----+-----+-----+-----+-----+-----+-----+	
	V K S G S C G T V V R N A E L K I V D P	
1201	CGACACCGCCGCTCCCTCGGCCAACCTGCCGGGGAGATCTGCATCCGGCAAGCA	1260
	-+-----+-----+-----+-----+-----+-----+-----+-----+-----+	
	D T G A S L G R N L P G E I C I R G K Q	
1261	GATCATGAAAGGTTACCTAAATGATCCGGTGGCCACAAAGAACACCATTGACAAGGACGG	1320
	-+-----+-----+-----+-----+-----+-----+-----+-----+-----+	
	I M K G Y L N D P V A T K N T I D K D G	
1321	TTGGCTGCATACTGGTGACATTGGTTATGTCGATGATGACGACGAGATCTTATTGCGA	1380
	-+-----+-----+-----+-----+-----+-----+-----+-----+-----+	
	W L H T G D I G Y V D D D D E I F I V D	
1381	CAGACTGAAGGAGATAATTAAATATAAGGGATTCCAAGTACCTCCGGCGGAACTTGAAGC	1440
	-+-----+-----+-----+-----+-----+-----+-----+-----+-----+	
	R L K E I I K Y K G F Q V P P A E L E A	
1441	CCTTCTCATTACACACCCCTGAAATCAAGGATGCTGCTGCGTATCGATGCAAGACGA	1500
	-+-----+-----+-----+-----+-----+-----+-----+-----+-----+	
	L L I T H P E I K D A A V V S M Q D E L	
1501	TGCTGGTGAAGTTCCGGTTGCGTTGTTGTGCGGACTGAGGGTTCAGAGATCAGCGAAA	1560
	-+-----+-----+-----+-----+-----+-----+-----+-----+-----+	
	A G E V P V A F V V R T E G S E I S E N	

FIGURE 3 CONTINUED

```

1561 CGAGATCAAGCAGTTGTTGCAAAAGAGGTTGTTCTACAAGAGGATCTGCAAAGTGTT 1620
-----+-----+-----+-----+-----+-----+
E I K Q F V A K E V V F Y K R I C K V F

1621 CTTCGCGGATTCCATTCAAAGAGTCCATCTGGCAAGATCCTCAGGAAGGACCTGAGAGC 1680
-----+-----+-----+-----+-----+-----+-----+
F A D S I P K S P S G K I L R K D L R A

1681 AAAGCTCGCCGCAGGCATTCCCAGCAGTAATACCACACAGTCCAAAAGCTAACAGATA 1740
-----+-----+-----+-----+-----+-----+-----+
K L A A G I P S S N T T Q S K S *

1741 TATTGTTCCAACCTTACACACCTCTGTCCAACACCATGTAATGTTCTTAATATAAACG 1800
-----+-----+-----+-----+-----+-----+-----+
GAAATTATTACATATAGAAGGGCTGATTCTTTACTAGATGTGTCCAACATATGATATG 1860
-----+-----+-----+-----+-----+-----+-----+
CTTGTAGGCCGATGTGTAACCTGTCATGTATAGATAACGCCCTTTTGACAAGAA 1920
-----+-----+-----+-----+-----+-----+-----+
AGGCTGATTATAATGTATACCGTGAACGTGAATATTGTTAGGGAGATCAAAAAAAAAA 1980
-----+-----+-----+-----+-----+-----+-----+
AAAAAAAAAAAAA 1992
-----+-----+

```

FIGURE 3 CONTINUED

1 CGGCACGAGATCTCCCACGACTAATTAGAAGAAGATTTACTTAGTCTCTGCTTCTCGCT 60
 1 CGATCGCCGGCCGGTAGGTTAGCTAGCTACTCGTACTAGACCATTACCATGGGTTCT 120
 61 M G S
 121 CGTGCCGGAGGAGTCAGTGGTGGCGGTGGCACCGCGGAGACGGTGTCCGGTCGAAGCT 180
 121 V P E E S V V A V A P A E T V F R S K L
 181 CCCCCACATCGAGATCAACAACGAGCAGACGCTGCAGAGCTACTGCTTCGAGAAGATGGC 240
 181 P D I E I N N E Q T L Q S Y C F E K M A
 241 CGAGGTCGCGTCCCGCCCTGCATCATCGACGGCCAGACGGCGCCTCCTACACCTACAC 300
 241 E V A S R P C I I D G Q T G A S Y T Y T
 301 GGAGGTCGACTCCCTGACCCGTCGCGCCGCGGGCTGCGCCGCATGGCGTGGGAA 360
 301 E V D S L T R R A A A G L R R M G V G K
 361 GGGCGACGTGGTGTGATGAACCTGCTGCGCAACTGCCGGAGTTCGCCTCTCCTCCTGGG 420
 361 G D V V M N L L R N C P E F A F S F L G
 421 CGCGCGCGGCTGGCGCCACCACCGCCAACCCGTTCTACACCCCGCACGAGAT 480
 421 A A R L G A A T T T A N P F Y T P H E I
 481 CCACCGCCAGGCGGAGGCGCGGGCGCCAAGCTGATCGTACCGAGGCCTGCGCCGTGGA 540
 481 H R Q A E A A G A K L I V T E A C A V E
 541 GAAGGTGCTGGAGTTCGCGCGGGGGCGGGCGTGGTACCGTCGACGGAGCG 600
 541 K V L E F A A G R G V P V V T V D G R R
 601 CGACGGGTGCGTGGACTTCGCGGAGCTGATCGCCGGCGAGGAGCTGCCGAGGCGAACGA 660
 601 D G C V D F A E L I A G E E L P E A D E
 661 GGCGGGGGTCTCCCCGACGACGTCGTCGCCCTGCCCTACTCCTCCGGCACCCACCGGGCT 720
 661 A G V L P D D V V A L P Y S S G T T G L
 721 CCCCCAAGGGCGTCATGCTACCCACCGCAGCCTCGTCACCAGCGTCGCCAGCTGGTCGA 780
 721 P K G V M L T H R S L V T S V A Q L V D

FIGURE 4

781	CGGGTCGAACCTAACGTGTGCTCAACAAGGACGACCGCTGCTGTGCCTGCTGCCGCT G S N P N V C F N K D D A L L C L L P L	840
841	CTTCCACATCTACTCGCTGCACACGGTGTGCTGGCGGGCTCCGCGTCGGCGCCGCGAT F H I Y S L H T V L L A G L R V G A A I	900
901	CGTCATCATGCGCAAGTTCGACGTGGCGCTGGTGGACCTCGTCCGCGCGACCGCAT V I M R K F D V G A L V D L V R A H R I	960
961	CACCATCGCGCCATTGTGCCGCCATCGTCGTGGAGATCGCCAAGAGCGACCGCGTCGG T I A P F V P P I V V E I A K S D R V G	1020
1021	CGCCGACGACCTCGCATCCGCATGGTGCTCTCCGGCGCCGCCATGGGCAAGGA A D D L A S I R M V L S G A A P M G K D	1080
1081	CCTCCAGGACGCCCTCATGGCCAAGATCCCCAACGCCGTGCTGGACAGGGGTACGGGAT L Q D A F M A K I P N A V L G Q G Y G M	1140
1141	GACCGAGGCTGGCCGGTGGCATGTGCCTGGCGTCGCCAAGGAGCCGTTCAAGGT T E A G P V L A M C L A F A K E P F F K V	1200
1201	CAAGTCCGGGTCGTGCGAACCGTGGTGCGAACGCCGAGCTCAAGGTGTCGACCCGA K S G S C G T V V R N A E L K V V D P D	1260
1261	CACCGGCGCATCCCTGGCCGGAACCGCTGGCGAGATTTGCGTCCGGGGAAAGCAGAT T G A S L G R N Q P G E I C V R G K Q I	1320
1321	CATGATAGGTTACCTGAACGACCCAGAGTCGACCAAGAACACCATCGACAAGGACGGCTG M I G Y L N D P E S T K N T I D K D G W	1380
1381	GCTGCACACCGGAGACATCGGCTGGATGACCGACGAGATCTTCATCGTCGACAG L H T G D I G L V D D D E I F I V D R	1440
1441	GCTCAAGGAGATCATCAAGTACAAGGGCTTCCAAGTGGCGCCGGAGCTCGAGGCCCT L K E I I K Y K G F Q V A P A E L E A L	1500
1501	CCTCCTCACGAACCGGAGGTCAAGGACGCCGCCGTGTAAGGGTGAAGGATGATCTCTG L L T N P E V K D A A V V G V K D D L C	1560

FIGURE 4 CONTINUED

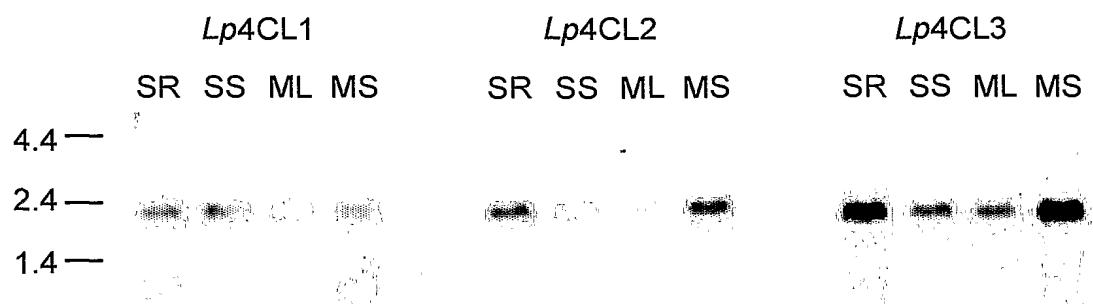
1561	CGGCGAAGTCCGGTCGCCTTCATTAAGAGGATCGAAGGATCTGAGATCAACGAGAACGA -----+-----+-----+-----+-----+-----+-----+-----+ G E V P V A F I K R I E G S E I N E N E	1620
1621	GATCAAGCAATT CGTCTCAAAGGAGGTGTTCTACAAGAGGATCAACAAGGTCTACTT -----+-----+-----+-----+-----+-----+-----+ I K Q F V S K E V V F Y K R I N K V Y F	1680
1681	CACCGACTCCATTCCCAAGAACCCCTCCGGCAAGATCCTAAGGAAGGACTTGAGAGCCAG -----+-----+-----+-----+-----+-----+-----+ T D S I P K N P S G K I L R K D L R A R	1740
1741	GCTCGCCGCTGGCATCCCCACCGAAGTTGCCGCGCCGAGAACGCTAAGGGCCGTTCTCAG -----+-----+-----+-----+-----+-----+-----+ L A A G I P T E V A A P R S *	1800
1801	GAACGCAGTCACCCATGGT GCTGTTAGGTGCTTTATAGACCAACACCAAATGGGAAAG -----+-----+-----+-----+-----+-----+-----+ -----	1860
1861	AAACTACGGGAGGGGATCATATTATTGTTGCAGGAGATATCAGTTGTTGATTGCCCTG -----+-----+-----+-----+-----+-----+-----+ -----	1920
1921	CTTGTGTAATGTTGATAAAATGAAATGATATAATAGATGTGTTGTTTATTTTGACCA -----+-----+-----+-----+-----+-----+-----+ -----	1980
1981	TGTAAGAACAAAGGCTGTTTATACACTTATTTTGAAAAA -----+-----+-----+-----+-----+-----+-----+ -----	2038

FIGURE 4 CONTINUED

FIGURE 5

	10	20	30	40	50	60
<i>Lp4CL1</i>	MITVAAPENVQQPQIAAAAAAVEAAAPEATTIFRSRSLPDIQIPTHMPILHDYCFATAASAPD					
<i>Lp4CL2</i>	MGSIAADAPPAL..VFRSKLPDIEIPTHLTLQDYCFQRLPELSA					
<i>Lp4CL3</i>	MGSVPEESVVAVAPAETVFRSKLPDIEINNEQTLQSYCFEKMAEVAS					
	70	80	90	100	110	120
<i>Lp4CL1</i>	APCLITAATGKTYTFAETHLLCRKAAAALHGLGVRHGDRIMLLQNSVEFALAEFGASML					
<i>Lp4CL2</i>	RACLIDGATGAALTYGEVDALSRCAAGLRRRLGVGKGDVVMALLRNCPEFAFVFLGAARL					
<i>Lp4CL3</i>	RPCIIDGQGTGASYTYTEVDSLTRRAAGLRRMGVGKGDVVMNLLRNCPEFAFSFLGAARL					
	130	140	150	160	170	180
<i>Lp4CL1</i>	GAVSTAANPFCCTPOEIHKOLVASGAKLIVVTQSAYVOKLRHEAFPRIGEALTVITIDEDDG					
<i>Lp4CL2</i>	GAATTTANPFYTPHEIHRQATAAGARVIVTEACAVEKVRAFAAERGIPVVSV.....DE					
<i>Lp4CL3</i>	GAATTTANPFYTPHEIHRQAAAGAKLIVTEACAVEKVLEFAAGRGIPVVSV.....DG					
	190	200	210	220	230	240
<i>Lp4CL1</i>	TPDGCQPFWALVSAADENSVPESPIS..PDDAVALPYSSGTTGLPKGVVLTHGLVSSVA					
<i>Lp4CL2</i>	GVDGGCLPFAETLLGEESGERFVDEAVDPDDVVALPYSSGTTGLPKGVMLTHRSLVTSVA					
<i>Lp4CL3</i>	RRDGCVDF.AELIAGEELPEADEAGVL.PDDVVALPYSSGTTGLPKGVMLTHRSLVTSVA					
	250	260	270	280	290	300
<i>Lp4CL1</i>	QQVDGGENPNLHMRAGEDDVVLCLVPLFHFISLNSVLLCALRAGAAVMLMPREEMGAMLEGI					
<i>Lp4CL2</i>	QQVDGGENPNLHFSS.SDVLLCLVPLFHIYSLNSVLLAGLRAAGCAIVIMRKEDHGALVLDLV					
<i>Lp4CL3</i>	QIVDGSNPNVCFNK..DDALLCLLPLFHIYSLHTVLLAGLRRVGAIVIMRKEDVGALVLDLV					
	310	320	330	340	350	360
<i>Lp4CL1</i>	ERWRVTVAAVVPPLVLALAKNPGVEKHDLSSIRIIVLSGAAPIGKELEDALRGRLPQAIIG					
<i>Lp4CL2</i>	RTHGVTVPFVPPIVVETIAKSARVTAADLASIRLVMMSGAPVKGKELODAFMAKIPNAVIG					
<i>Lp4CL3</i>	RAHRTTIAPEFVPPIVVETIAKSDRVGADDLASIRMLSGAAPVKGKDLODAFMAKIPNAVIG					
	370	380	390	400	410	420
<i>Lp4CL1</i>	QGYGMTEAGPVLSMCPAFAREPPTPAKSGSGCTVVRNAOLKVVDPDTGVSLGRNLPGEICL					
<i>Lp4CL2</i>	QGYGMTEAGPVLMACLAFAKEPEFAVKSGSGCTVVRNAELKIVDPDTGASLGRNLPGEICL					
<i>Lp4CL3</i>	QGYGMTEAGPVLMACLAFAKEPEFKVKSGSGCTVVRNAELKVVDPDTGASLGRNQPGECIV					
	430	440	450	460	470	480
<i>Lp4CL1</i>	RGPQIMKGYLNDPVATAATIDVEGWLHTGDIGYVDDDDDEVFIVDRVKELIKFKGFQVPEA					
<i>Lp4CL2</i>	RGKQIMKGYLNDPVATAKNTIDKDGWLHTGDIGYVDDDDDEVFIVDRVKELIKFKGFQVPEA					
<i>Lp4CL3</i>	RGKQIMKGYLNDPESTKNTIDKDGWLHTGDIGYVDDDDDEVFIVDRVKELIKFKGFQVPEA					
	490	500	510	520	530	540
<i>Lp4CL1</i>	ELEALLLTAAHESIADAADVVPQKDDAAGEGPVAFVVRAADSIAABEAIKEFVSKOVVFYKRL					
<i>Lp4CL2</i>	ELEALLLTTHPEIKDAAVVSMQDELAEGEVPVAFVVRTEGSEISENEIKAOFVAKEVVFYKRI					
<i>Lp4CL3</i>	ELEALLLTNPVEVKDAAVVGVKDDLCGEVPVAFIKRTEGSEINENEIKAOFVSKEVVFYKRI					
	550	560	570			
<i>Lp4CL1</i>	HKVYFTHAIPKSASGKILRKELRAKLAAPATA					
<i>Lp4CL2</i>	CKVVFADSIPKSPSGKILRKDLRAKLAAGIPSSNTTQSKS					
<i>Lp4CL3</i>	NKVYFTDSIPKNPSGKILRKDLRARLAAQIPTEVAAPRS					

FIGURE 6



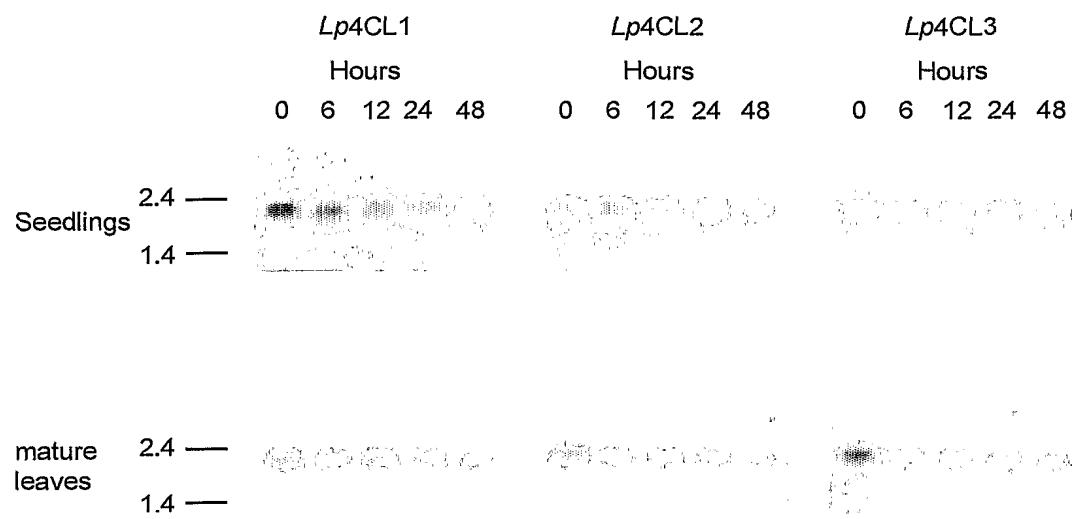


FIGURE 7

FIGURE 8

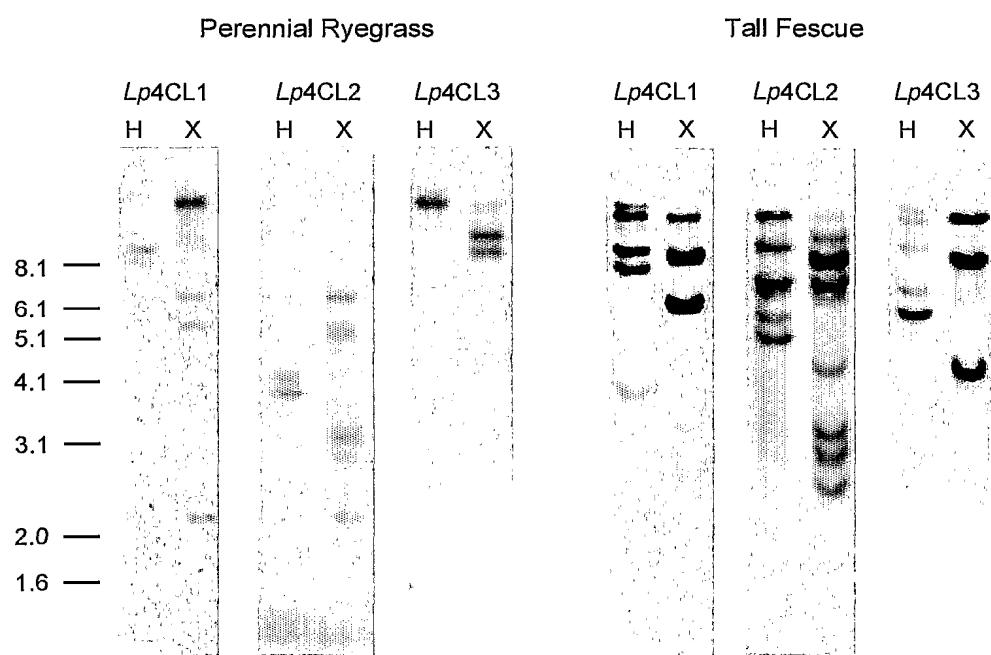
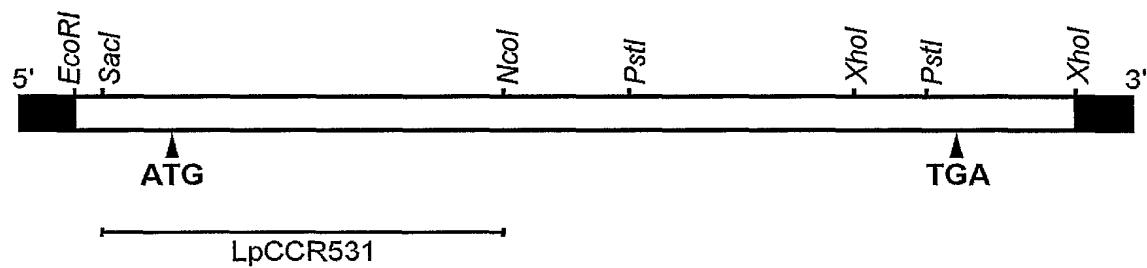


FIGURE 9



1	GGCACGAGGAATCCTACCAAACCGAGCTACCAGATCCTCTCTACTAATCGAGCTCCCTA	60
61	CGCTGCTCCGCCCTGTCTTCGTTCCGCCTCACCGCCGGCCGGTCTCCGCTCCAAGCTAC	120
121	GTCCGTCCGTCCACATATATAGCATCGACATGACCATCGCCGAGGTCGTGGCTGCCGGAG M T I A E V V A A G D	180
181	ACACCGCCGCCGCCGTGGTGCAGCCGCCGGAAACGGGCAGACCGTGTGCGTGACCGCG T A A A V V Q P A G N G Q T V C V T G A	240
241	CCGCCGGGTACATCGCGTGTGGCTCGTCAAGCTGCTGGAGAAGGGGTACACCGTCA A G Y I A S W L V K L L L E K G Y T V K	300
301	AGGGCACCGTCAGGAACCCAGACGACCCGAAGAACGCGCACCTGAGGGCGCTCGACGGCG G T V R N P D D P K N A H L R A L D G A	360
361	CCGCCGACCGGCTGGTCCTCTGCAAGGCCGACCTCCTCGACTACGACGCCATCCGCCCG A D R L V L C K A D L L D Y D A I R R A	420
421	CCATCGACGGCTGCCACGGGTCTTCCACACCGCGTCCCCCGTCACCGACGACCCCCGAGC I D G C H G V F H T A S P V T D D P E Q	480
481	AAATGGTGGAGCCGGCGGTGAGGGGCACGCAGTACGTATAGACGCGGCGGGAGGCCG M V E P A V R G T Q Y V I D A A A E A G	540
541	GCACGGTGGCGGGATGGTGCTCACCTCCATCGCGCCGTCACCATGGACCCCAACC T V R R M V L T S S I G A V T M D P N R	600
601	GCGGGCCGGACGTGGTCGACGAGTCGTGCTGGAGCGACCTCGACTTCTGCAAGAAAA G P D V V V D E S C W S D L D F C K K T	660
661	CCAGGAACTGGTACTGCTACGGGAAGGCCGGTTGGGAGCAGGGCGCATCGGAGTTGGCGC R N W Y C Y G K A V A E Q A A S E L A R	720
721	GGCAGCGCGGCGTGGACCTTGTGGTGGTGAACCCGGTGTGGTGATCGGCCCCCTGCTGC Q R G V D L V V V N P V L V I G P L L Q	780

FIGURE 10

781	AGCCGACGGTGAACGCCAGCATCGGCCACATCCTCAAGTACCTGGACGGGTCGGCCAGCA -----+-----+-----+-----+-----+-----+-----+ P T V N A S I G H I L K Y L D G S A S K	840
841	AGTTCGCCAACGCCGTGCAGGCGTACGTGGACGTCCGCGACGTGGCGACGCCACCTCC -----+-----+-----+-----+-----+-----+ F A N A V Q A Y V D V R D V A D A H L R	900
901	GCGTCTTCGAGTGCGCCGCGTCCGGCCGCCACCTCTGCCGAGCGCGTCCCTCCACC -----+-----+-----+-----+-----+-----+ V F E C A A A S G R H L C A E R V L H R	960
961	GCGAGGACGTGCGCATCCTCGCCAAGCTCTTCCCCGAGTACCCGTCCCCACCAAGGT -----+-----+-----+-----+-----+-----+ E D V V R I L A K L F P E Y P V P T R C	1020
1021	GCTCTGATGAGACGAACCGAGGAAGCAGCCATACAAGATGTCGAACCAGAAAGCTCCAGG -----+-----+-----+-----+-----+-----+ S D E T N P R K Q P Y K M S N Q K L Q D	1080
1081	ACCTCGGACTCGAGTTCAGGCCGGTGAGCCAGTCCTGTACGAGACGGTGAAGAGCCTCC -----+-----+-----+-----+-----+-----+ L G L E F R P V S Q S L Y E T V K S L Q	1140
1141	AGGAGAAGGGCCACCTTCCGGTGCTCAGCGAGCAGGCAGAGGGGGACAAGGAAACCTTAG -----+-----+-----+-----+-----+-----+ E K G H L P V L S E Q A E A D K E T L A	1200
1201	CTGCCGAGCTGCAGGCAGGGTTACCATCCGAGCATGAGGAACAAGAAATCAACCATGTC -----+-----+-----+-----+-----+-----+ A E L Q A G V T I R A *	1260
1261	CATACTGCTACTGTCATGTAAACCAGCTGTTGAATGCCTAAATCTAAGTTCTTGTAAATA -----+-----+-----+-----+-----+-----+ 	1320
1321	CTGTGTTGTTCATGTGGACTAGATTGATCGAATAAACATCTCTACACAAGGTTGCTAAA -----+-----+-----+-----+-----+-----+ 	1380
1381	AAAAAAAAAAAAAA -----+----- 1395	

FIGURE 10 CONTINUED

FIGURE 11

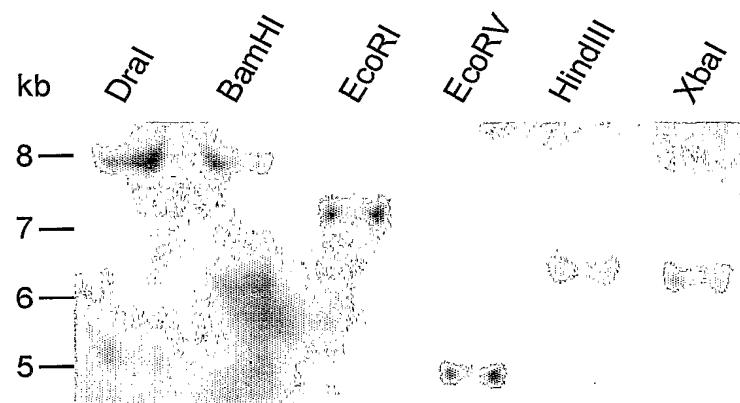
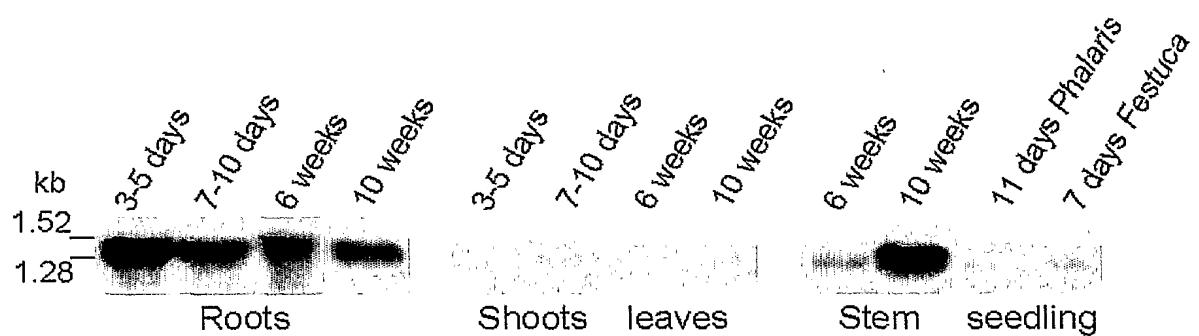


FIGURE 12



1 GGCACGAGCAACAAGTCATCAATGGCGGAAGGCTTGCCGGCCTCGGTTGGGCTGCGAGG
 60 M A E G L P A L G W A A R
 61 GACGCCCTCCGGTCACCTCTCCCTTACAGCTCTCGAGAACGCTTCCGAAGGACGACgAT
 120 D A S G H L S P Y S F S R S V P K D D D
 121 GTGACGATCAAGGTGCTTCTGCCACACTGACCTCCACATCATCAAGAAC
 180 V T I K V L F C G I C H T D L H I I K N
 181 GACTGGGGCAACGCCCTACCCATCGTCCCAGGGCATGAGATCGTGGCGTCGCGCC
 240 D W G N A L Y P I V P G H E I V G V V A
 241 AGCGTCGGCAGCGGCGTCAGCAGCTCAAGGCCGGCgACACGGTGGCGTGGCTACTTC
 300 S V G S G V S S F K A G D T V G V G Y F
 301 CTCGACTCCTGCCGCACCTGCTACAGCTGCAGCAAGGGTACGAGAACTTCTGCCACC
 360 L D S C R T C Y S C S K G Y E N F C P T
 361 CTGACGCTCACCTCAAACGGCGTCAGGCCGGCGGCCACCACCCAGGGCGGCTCTCC
 420 L T L T S N G V D G G G A T T Q G G F S
 421 GACGTCTCGTCGTAACAAGGACTACGTCATCCGCGTCCCGACAACTGCCCTGGCC
 480 D V L V V N K D Y V I R V P D N L P L A
 481 GCGCGGCACCTCTCCCTGCGCCGGCGTCACAGTCTACAGCCATGGGGAGTACGGC
 540 G A A P L L C A G V T V Y S P M V E Y G
 541 CTCAACGCCCCCgGGAAGCACyTCGGcGTCGTCGGCCTGGCGGGCTGGCCACGTCGcC
 600 L N A P G K H X G V V G L G G L G H V A
 601 GTCAAGTTGGCAAGGCCTCGGGATGACCGTCACCGTCATCAGCTCTGGACAGGAAG
 660 V K F G K A F G M T V T V I S S S D R K
 661 CGCGACGAGGCCTCGGCCGCTCGGCGCCGACGCGTTCTCGTCAGCAGCGACCCCCGAG
 720 R D E A L G R L G A D A F L V S S D P E

FIGURE 13

CAGATGAAGCGGGCGGGCACCATGGACGGCATCATCGACACGGTGTCCGCGGGCCAC 721
 Q M K A A A G T M D G I I D T V S A G H 780

 CCGATCGTCCGCTGCTGACCTGCTCAAGCCCATGGGCAGATGGTCGTGGTGGCGCG 781
 P I V P L L D L L K P M G Q M V V V G A 840

 CCCAGCAAGCCGCTCGAGCTCCGGCCTTCGCCATCATCGGCGGCCAAGCGCCTCGCC 841
 P S K P L E L P A F A I I G G G K R L A 900

 GGGAGCGGCACCGGCAGCGTCGCACACTGCCagGCCATGCTCGACTTCGCGGGCAAGCAC 901
 G S G T G S V A H C Q A M L D F A G K H 960

 GGCATCACCGCCGACGTCGAGGTGCGTCAAGATGGACTACgGTCAACACCGCCATCGAGCG 961
 G I T A D V E V V K M D Y G Q H R H R A 1020

 GCTAGAGAAGAACGACGTCAGGTACCGCTTCGTCATCGACGTCGCCAGGCCACCTGCA 1021
 A R E E R R Q V P L R H R R R Q P P A 1080

 GGGCACCGCCGCTTAACTTGTGCTACACAATGTGGACGCGCGCTCGTTGGTCCAGAAAA 1081
 G H R R L T C A T Q C G R A L V W S R K 1140

 AGGTTCGCCGGCTCACAGCCACATGAACAAAGTCAATGAGTCGTTGGTGTGTTATCT 1141
 R F A G S Q P H E Q V N E S L V C C L S 1200

 TCATTCCACATATGGGACGCAGTTCCAGATTTCATGTCAAATAATTGCGTCGTGCGG 1201
 S F H I W D A V P D F H V K 1260

 TTGTCAAGACTCAAATAGGAGAAAAAAAGACTCGTGATTGCGTTTGCAAAAAAA 1261
 AAAAAA 1320

 1321 1325

FIGURE 13 CONTINUED

1	GGCACGAGTCGCCTCCAACGTCTTCCCTTAACCGGCCGTCCCTACGC	tTGCA	CCACCACCACC	60
61	ACGCACAGACAGAGCAGTTCCCAGCCCCGCCGGAACCGGATGGC	ACCCACGGCGCGG		120
	M A P T A A E			
121	AGCAGACGGAGCACCACCAAGCACACCAGGAAGGCGGTGGGCTGGCGCGCGACGACG			180
	Q T E H H Q H T R K A V G L A A R D D A			
181	CCGGCCACCTCTCCCCGCTGCCATCACACGGAGGAGCACAGGAGACGACGATGTGGTGA			240
	G H L S P L A I T R R S T G D D D V V I			
241	TAAAGATTTGACTGCGGAATCTGCCACTCTGACCTGCACGCCCTGAAGAACGACTGGA			300
	K I L Y C G I C H S D L H A L K N D W K			
301	AGAACTCAAGGTACCCGATGATCCCCGGCACGAGATGCCGGCGAGGTACGGAGGTGG			360
	N S R Y P M I P G H E I A G E V T E V G			
361	GCAAGAACGTGAGCAAGTTCAAGGCCGGCACCGCGTGGCGTCGGTGCATGGTAACT			420
	K N V S K F K A G D R V G V G C M V N S			
421	CGTGCCGGTCGTGCGAGAGCTGCGACAAGGGCTTCGAGAACCACTGCCCGGCATGATCC			480
	C R S C E S C D K G F E N H C P G M I L			
481	TCACCTACAACTCGGTCGACGTCGACGGCACCGTCACCTACGGCGTACTCCAGCATGG			540
	T Y N S V D V D G T V T Y G G Y S S M V			
541	TGGTGGTGCACGAGCGGTTCGTGGTCCGGTCCCGACGCCATGCCGTGGACAAGGGCG			600
	V V H E R F V V R F P D A M P L D K G A			
601	CGCCGCTGCTGCGCCGGCATACCGTGTACAGCCCCATGAAGTACCAACGGGCTCAACG			660
	P L L C A G I T V Y S P M K Y H G L N V			
661	TTCCCGGGCTGCACCTCGCGTGTGGGCTGGCGGGCTGGCCACGTTGCGGTCAAGT			720
	P G L H L G V L G L G G L G H V A V K F			
721	TCGGCAAGGCCTCGGAATGAAAGTGACGGTATCAGCTCGTCGCCGGGAAGAAGGAGG			780
	G K A F G M K V T V I S S S P G K K E E			

FIGURE 14

781	AGGCCCTGGGGCGGCTGGCGCCGACCGTTCATCGTCAGCAAGGACGCCGACGAGATGA -----+-----+-----+-----+-----+-----+-----+ A L G R L G A D A F I V S K D A D E M K	840
841	AGGCTGTGATAGCACCATGGATGGCATCANTAAACACGGTATCTGCAAACATCCCCCTGA -----+-----+-----+-----+-----+-----+-----+ A V I A P W M A S X N T V S A N I P L T	900
901	CCCCCTCTCTTCGGGCTGCTCAAGCCCAACGGCAAGATGATCATGGTCGGCCTCCCCGAGA -----+-----+-----+-----+-----+-----+-----+ P L F G L L K P N G K M I M V G L P E K	960
961	AGCCCATCGAGATTCCCTCCCTTCGCTCTAGTTGCCACGAATAAGACCCTGGCCGGGAGCA -----+-----+-----+-----+-----+-----+-----+ P I E I P P F A L V A T N K T L A G S I	1020
1021	TCATCGGGCATGAGCGACACGCAGGAGATGCTGGACCTCGCGCGAAGCACGGCGTGA -----+-----+-----+-----+-----+-----+-----+ I G G M S D T Q E M L D L A A K H G V T	1080
1081	CGGCCGACATCGAGGTGGTCGGCGCGGAGTATGTGAACACGGCCTGGAGCGCCTGCCA -----+-----+-----+-----+-----+-----+-----+ A D I E V V G A E Y V N T A L E R L A K	1140
1141	AGAACGACGTCAAGGTATCGCTTCGTATCGACATCGCAACACCCCTCGACAATGTTGG -----+-----+-----+-----+-----+-----+-----+ N D V R Y R F V I D I G N T L D N V A A	1200
1201	CCACCACCGAGTGAACGTACTCAGCACTGCTTACGATCTACGTTGTTCCACTGTTAGTGC -----+-----+-----+-----+-----+-----+-----+ T T E *	1260
1261	TCCGTAGTAAACAATAACGATCAAAACTCTTGTATCTGGTGCATTGGTAGACATGG -----+-----+-----+-----+-----+-----+-----+ TTGTTGCGAGGAAACTGAGTTGAAGGATGGATGGATAAAAAAAAAAAAAAA	1320
1321	-----+-----+-----+-----+-----+-----+-----+ 1378	1378

FIGURE 14 CONTINUED

FIGURE 15

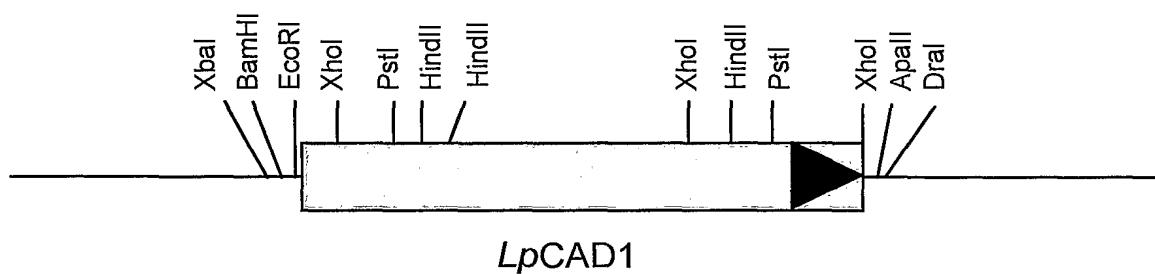
*LpCAD1*

FIGURE 16

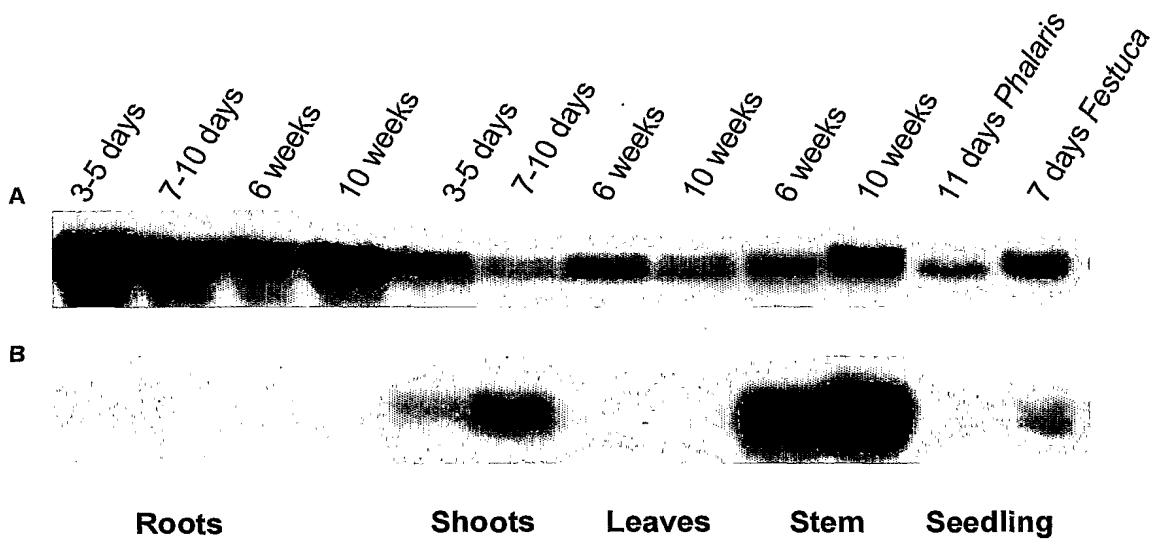
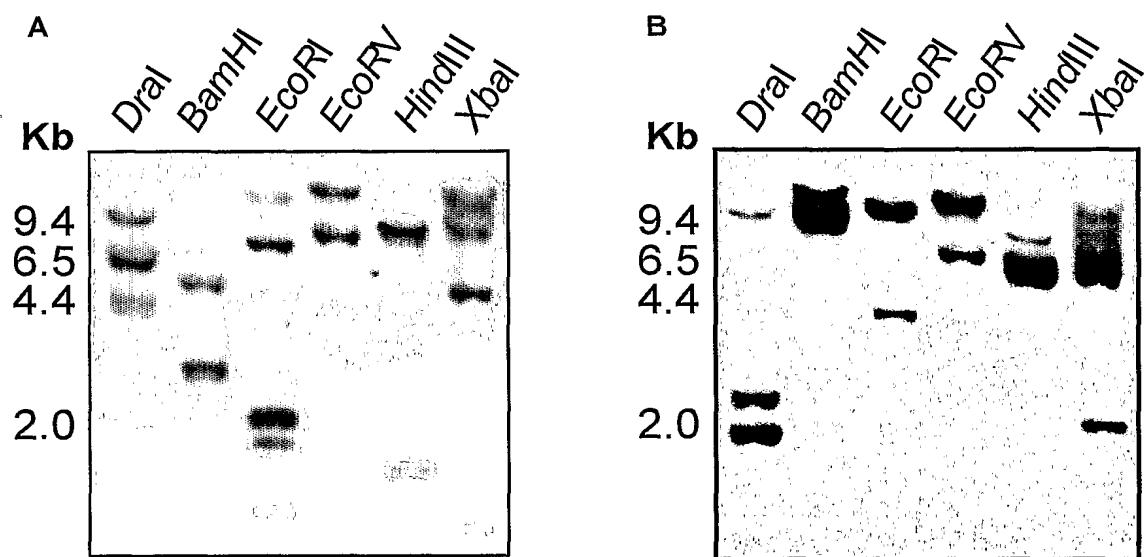


FIGURE 17



pBluescript	
GC	GGCCGCTCTAAACTAGTGGATCCCCGGCTGCAGGAATTGATATCAAGCTTATnG
SaI	
ATACCGTCGACAGCGGT	TnCAAATCCCGGTCCTGGGTGGAAGTnAGCAGTGGAAAGA
-4580	TGTGTGCGAGGGGTTGTGTTGGATGnAAGACAGGCGGCCAGTGGAGAACAGAGAGA
-4520	ACGCGAGAGGCCAAAGTATCCGCAGCCCCGCAAACAAGGCCTAGATTGGGTTAAGTTG
-4460	GGTCGTCTCAGACACCGCGGCCATCCTTTAGGTGGTCCGCGCTGGACCGTATTTTA
-4400	TCTGAGTTGACCCATTCAAGACGCGCAGACACGAGATGGATGGTGCAGTwAgAGATGACCT
-4340	AAGTACAArAACCTCTCCCCGA . GCTGCCGCCATC _c GTCACCTACCGAGCGAcAAAGcTT
-4280	CCCACTTCATCACACTCAGCCCAGCAAGCATACTGATGGTGAGGCCACTCGGGCTGTGC
-4220	CCACCGACCCCCACGCCATCCAAAACCAACTCTACTTTTCACCAmCACCAACAAAAGACAAA
-4160	ATATGGGATTTGTGATGAGATGGAAGCGGAGCTTGTCAAGAATGGAAACGCATAAAT
-4100	CGAGAACACGTATACAGTGCTGGAAATTGGATGACTAAGCCCCAAGGGTTAGAAAAAAA
-4040	XbaI TnAGACCATGTCTAGATGGAATTAGACATTTTTGATATAATAGAAGCGGGACTTGGCGC
-3980	GACAATTCTCAAACCTCGTCCCTAACAGGTATCGAAC _t TTCGA _t AGTTAGCGTGTGCTACT
-3920	GCggAcCCCCAACCAACTGTGTTAAGGCCACATC _g GTTAAGGCCAAGGGTTAGATGAAA
-3860	GTACCAATTCTCACTCATTTGCGACTAGCTACAAAACCTGCTTTCACATGTACGGTCATA
-3800	CTACAATTGACCTGGTAACGTAAGTATGGACTGTATGGTGTGCTAAGGTGTGTTGGC
-3740	AGCTCAAATAAACCAAAATTCAACACACGTCAACCATGAAC _t GAGATTACACACCAAC
-3680	GGCTGAGCCGTCTCCTTAAAAGATAGAGGGAGAAACCATAATCACCATTGGTGGTCAT
-3620	GTGTGAGTGTGCAAGCAAAAAAAATGGAGAACGCAACCCGTTGAGAGAGTGCAGAGAG
-3560	CATACAAGAACACCAACACAAAGTGTGAAGGGAGAAAAGAATATGAGATAAGATTTCGGA

FIGURE 18

-3500	AATACTTTGCACACCCATGCATGGGTGTGGGTGTTCCGTACCGTCTATGTATTCCTC	-3441
-3440	GAAATTCACTGCCACCATGGTAGATAAAAATATTTTTCTCTCCTCTTTTATTCAA	-3381
-3380	ATCTCAAAGCATAAkrArTGGTGACAGAACGATAAGATTCTACCTAGCTTCTGAGATC	-3321
-3320	CCACTAGTTATCTCAACCTGGTATTGAAGGATTAAACCATGCTTGAATTAGATTGGCT	-3261
-3260	TCAAACCTGGTAGTAGCTTGTTCATACTTGATTACTTGGTATGGTAGTTGGTTGA	-3201
-3200	GATTTGGTCAATGTAGAACATCAGATTTGAGAGCGATTGTCAGCTTGAATTGCCGCAGTT	-3141
-3140	TAGCACATACTAGTTGGATAGATGAACAGTTGGAGAGACAAATAATGCTATACGAGC	-3081
-3080	TCATCGGATAATATTAGTCTATGGCTTGTGCTCGGTGTCCTCTGCAAACCTTACCCC	-3021
-3020	TCTGTAGATGGTAGGATTTCTGATATCCTTCACTGGTTAAGGGTGTGCGTGTAAAGGAA	-2961
-2960	CGGGAGATACCGGATCACACCTTTCGTCTACACTTACAAGCATGTAACACCTAACGATT	-2901
-2900	GATTGATATCTAGGCTTACACCCATGGAGGTAAACTAATATTATTGAAATGCGACTTT	-2841
-2840	TCAAAAGTCCAATATAACCTTGACGATGATCTTACAACACTCGCGCCAGTCTGTATG	-2781
-2780	<i>KpnI</i> ATATCAGATTGGCCGAGGATCGTGGTACCTTGTAGTGGACTATGATGCTCATGGAGGTT	-2721
-2720	GTATGGACATGTTGTAATGCTGGTTCTCTAGGTTTTCTAATCAACTTGGCATTCTT	-2661
-2660	CTCCTAACACATAATAAGAGGGAACCTCCATACATTATTCTGAAAAAAGCATGGCCA	-2601
-2600	ACAATGAAACAGAAAAGTACGACAGTCTATACCCGACCCAAACAATGGCTCAGGTCTT	-2541
-2540	TCACGATGCATAGTTGTTAGCATGTATTTATAGTAGGAACAAAATTAAAGACAAC	-2481
-2480	TGcnAAAACAATTGTCCTTGAGTGTGTTTAAGGATGCGGCAATTATCGATTATAACA	-2421
-2420	TTACATATGTGATTGGATTAGCCAACTTTTGTCCTTCCgATGATCATATGAAAGGGTTGT	-2361
-2360	ATCTTAGGGCATCTCCAATGGGnAGACTCAAATGCAAAAAAAATnGTCCGTTGGGTCTC	-2301
-2300	CnGGACAAAACCTGCTCCCAACGGGGCAACCCAACTTAAACGGACAGGTGCAAGCGTCC	-2241

28 / 76

-2240	GGCnTGACCCAAAAGTGCAGCAAATTGGnAnATTTTGGGGCnAGCCAGACGAACGCGG	-2181
-2180	GCGTCCACTGTATCCGACTATGTCCGCATCCTGGCCCCTCTGACAGTGACACAAAATACA	-2121
-2120	ACCACATGCGCCCCCACCCTCTCTCCCTCCGTTGCCCTTCCCAGGAAAnCnGTCC	-2061
-2060	TCGCTCCTCGCCGGAATTGATCTCGCTAACCATGCTCCGCCACCCCTCGcCTkAAGG	-2001
-2000	CCCCAgCCGCCGCTACCCTCCCTTTGTCAGCCATTgGAAGTCGCCGgAGTTGAAACGA	-1941
-1940	GCGCCGCCAGCCTcGACACCGCCGAGCAAGACGAAGACTGGCCGGAGCTGCCGAGACGG	-1881
-1880	GACGGGGACGGAGCTGCCATGCGTGCCTCGCAGGGGCGCGATGGGGCGGAGCTGCCG	-1821
-1820	<i>PstI</i> TGGCTGGCTGCAcCCTCGCGGAGCTACCTCGTCGCGCCAGGGCCGCCCGCCCTGCCGACCGgCGg	-1761
-1760	ACGCCGCCCCGTGCTACCTCGTCGCGCCAGGGCCGCCCGCCCTGCCGACCGgCGg	-1701
-1700	CGgAgACGCGAcCTTCGCGGAGCTGCCGGCGAGAGACGCCGCTTCGCGACAGCGCC	-1641
-1640	CTCCTCGATCTCGTCGAGCCGATACGCgGcTAgGAgGGACGCCGGCTCCCGGTGTC	-1581
-1580	GGCCTCCGTGTGGCGCATCGCGGCCCTCCGTCGAGGCGCCGAGCTTCCTCGCCGGCG	-1521
-1520	TCGTGGCGCAGCCTGCCCTGATTGGCTCTGAGGCGCGGCGAGCTTCCTCGCCGGCG	-1461
-1460	GCGGGCGGAGCCTCCGCTGCCGCGACCTGCTCTGCCGGTCCGAGACGCCGGCG	-1401
-1400	GGCAGAGCTCCTCGCGCGCTCGGCGCGCTCCCTCGCCGGCGATGGCGCTCCAGGC	-1341
-1340	TCGCACGCCCTCCGGGTGGCGCAGCGAGAGCGCAGCCCTCCGGTGAGTTAGGCACAGG	-1281
-1280	CGCGACACGACATCCCCGGCTCGGCGAGCGCAGCGCAGCGCAGCGCAGCGTACGCC	-1221
-1220	TAGGTTGGCAACTAGTaCTACGAGGAAGAAAGAGGAGAAACAATTATTGGGTACAGCG	-1161
-1160	TTGGCGTACTGTGCGATCCAAACGGACACCCGAGCGCAGCGAAGCGTCCGCG	-1101
-1100	GTGGcGACCCAAACGACCCGAAACGGACGTCCGTTGGGTCCGGTGCCTGGAGATGCCCT	-1041
-1040	TACTCCCCATCCTCAAATGAGTCTAATTATATCTTGTGTAAGTTAAAAAGTTAA	-981

FIGURE 18 CONTINUED

-980 ACTTTGATCAACATTAGTAATGATAGTAGCAACGAATACAAAATTAAATTGTAATAATAT
 -920 ATTATGAAACTTTATTAAAGATGGATCTAGTTATACTAATTCTGGGATGGAGGAAG
 -860 TAGCTAAATATTGTTAATTCTAAATAAAATTAAACTTAACTTAAACAAAGTTA
 -800 Putative Myb Binding domains CAAGCATAATTATCTGtGGATGGAGGAAGtAGCTAAGATAACACCAATCCTCTCTACAT
 -740 TACCTAGCATGCCACATCAGGAAACTATTAGGATAAGCTCCAAGGAACCACCCAGAAC
 -680 ACAATTTACATGGCCTGGCTACCTAATGACAATTCCGAGCAACTGGTGGTGGTAC
 -620 GCGTTCCCTGTTCAATTGTCCTATTACAAGAGTGGCCCTGTATAGGTAAAAAAATAA
 -560 *HindIII* *PstI* CAAGCTTCAAGGACGGCCATGTTCCCTGTTCTCAGGCTGCACGTACTCACGACGAAG
 -500 TGTATCTCGTGTCTGGACATTGTCTCGCGCATTGTAAACCAGAAATTAAAAATGTG
 -440 GTGGCCTGCTATATCTGTATGGGGGTATCATGCACTCCTCGCAGAGGAATCCAGACGAC
 -380 GATTTACACGTGTTCCACCTTAGCTTTTTAAGTGTGTGTAAGGAACGATCATATA
 -320 *XbaI* ACTGCCCTGAATGCTGCATATATATAACCGACTCCATCATGTACTCGAGACAAGGTG
 -260 TCAAGAAAAACAAACTATGCCTATCTCACTAGCAATGATTGAGAGTACAGCTTTCCGG
 -200 TGCCATATTTTCCTATATATCTTTCTGAAGAACAAAGAAAAAAACAGTTGGTGT
 -140 GGTGGTTGGTGAAGCGAGAAAGCCCCATATAAGCCCTGCTCACCCCTCCCCGCAAAGCACA
 -80 *PvuI* ACTCATAGCTCGGGCTCTCGCTCACACCAAAATGCCAACAGCACCAGCATCTCGA
 -20 TCGGCAGACGCATAGATCGATGGGCTCCACCGCCGCCGACATGGCCGCGTCCGGACGA
 40 GGACGCGTGCATGTTGCCCTCCAGCTCGCTTCTCGTGGCTCCCGATGACGCTGAA
 100 GAACGCCATCGAGCTTGGCCTCCTGGAGATCCTGGTGGCCGCCGGCAAGTCGCTGAC
 160 CCCGACCGAGGTGGCCGCCAAGCTCCCGTCCGGCGGAACCCGGAAAGCCGGACATGGT
 39 M G S T A A D M A A S A D E
 99 D A C M F A L Q L A S S S V L P M T L K
 159 N A I E L G L L E I L V A A G G K S L T
 219 P T E V A A K L P S A A N P E A P D M V

FIGURE 18 CONTINUED

220	GGACCGCATACTCCGGCTGCTCGCGTCGTACAACGTCGTGACGTGCCTGGTGGAGGAGGG +-----+-----+-----+-----+-----+-----+-----+-----+-----+----- D R I L R L L A S Y N V V T C L V E E G	279
280	CAAGGACGGCCGCCCTCCCGGAGCTACGGCGCCGCGCCCGTGTGCAAGTCCCTACCCCC +-----+-----+-----+-----+-----+-----+-----+-----+-----+----- K D G R L S R S Y G A A P V C K F L T P	339
340	CAACGAGGACGGCGCTCCATGGCGGGCGCTCGCGTCATGAACCAAGGACAAGGTCCCTCAT +-----+-----+-----+-----+-----+-----+-----+-----+-----+----- N E D G V S M A A L A L M N Q D K V L M	399
Intron/exon boundary		
400	GGAGAG <u>CTG</u> GTGAGTCTCTCAGTGGAGCTAGTTACTGTAGATCCGAATTGTTCCCTTA +-----+-----+-----+-----+-----+-----+-----+-----+-----+----- E S	459
460	<i>SalI</i> pBluescript GTGAGGGTTAATTCCGGCGGCC <u>CTCGAC</u> CTCGAGGGGGGCCGGTACCCATTGCC +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	
TATA GTGAGTCGTATTACGCGCGCTCACTGGCGCGTGTACAAACGTCGTGACTGGGAA AACCTGGCGTTACCCAACTTAACCGACATCCCGCTTCGCCAGCTGGCGT AATAGCGAAGAGGCCCGCACCGATGCCCTTCCAACAGTTGCGCAGCCGTGAATGGCGAA TGGGACGCGCCCTGTAGCGCGCATTAAAGCGCGCGGGTGTGTG 744		

FIGURE 18 CONTINUED

FIGURE 19

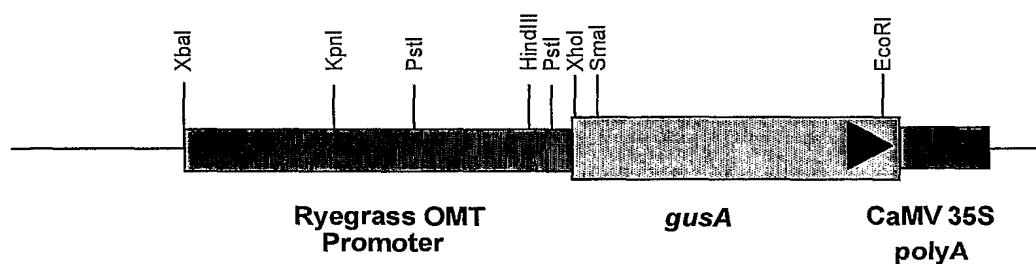


FIGURE 20

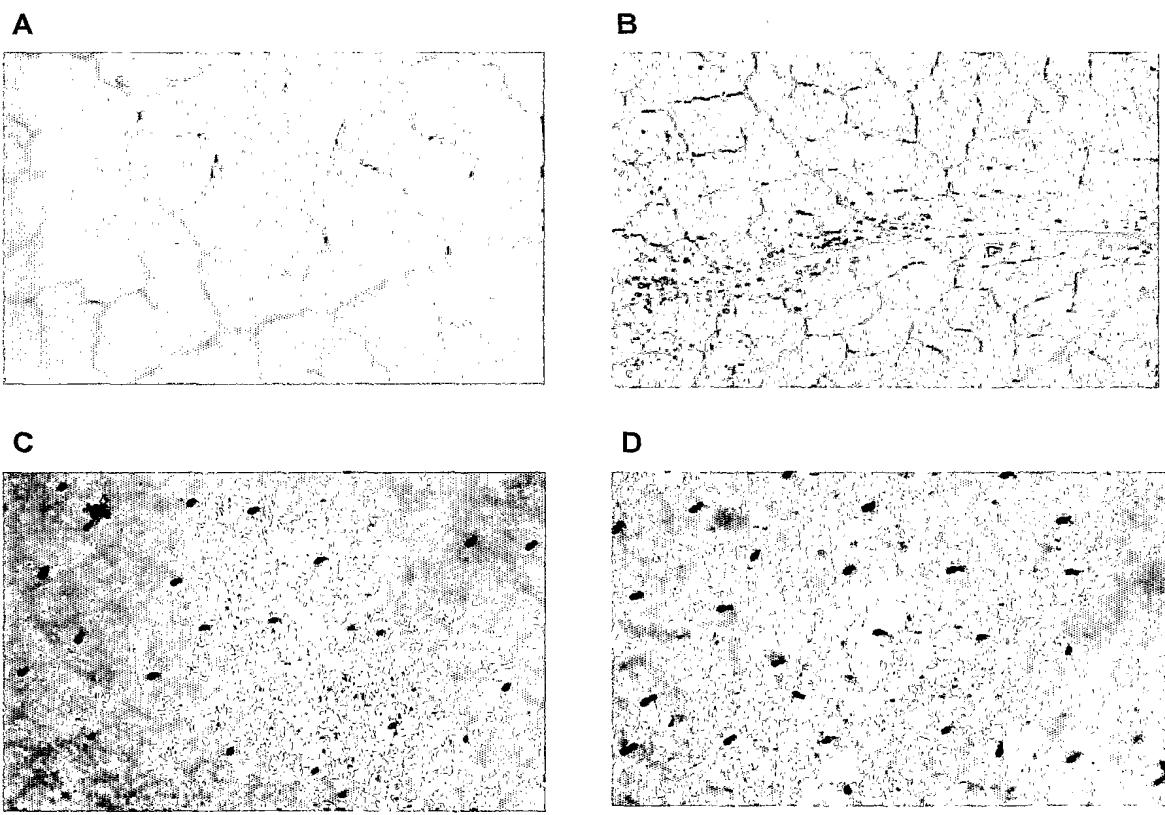
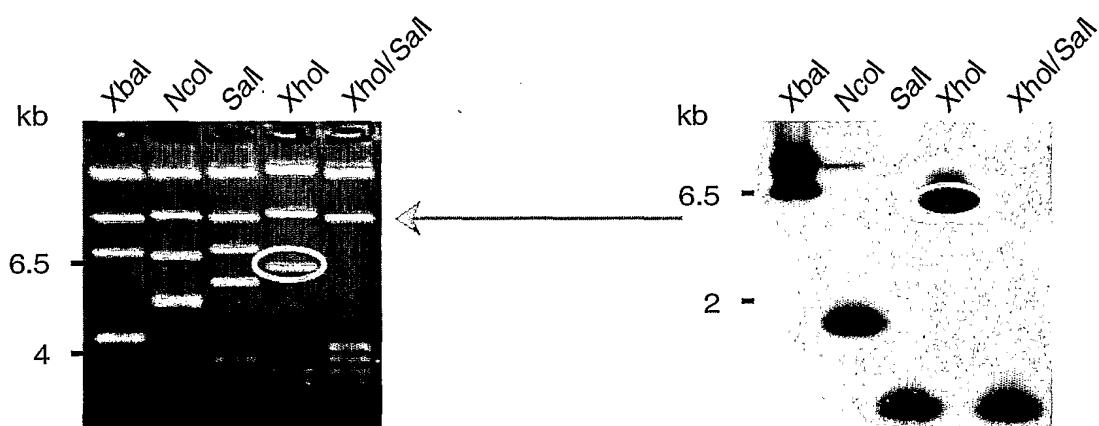
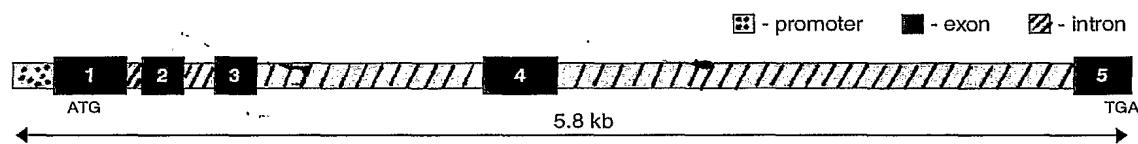


FIGURE 21

A**B****C**

Exon	<i>LpCCR1</i>	<i>EgCCR1</i>	<i>EsCCR1</i>	<i>PbCCR1</i>
1	173 bp	133 bp	133 bp	139 bp
2	155 bp	155 bp	155 bp	155 bp
3	189 bp	186 bp	186 bp	186 bp
4	353 bp	353 bp	353 bp	353 bp
5	220 bp	218 bp	184 bp	184 bp

FIGURE 22

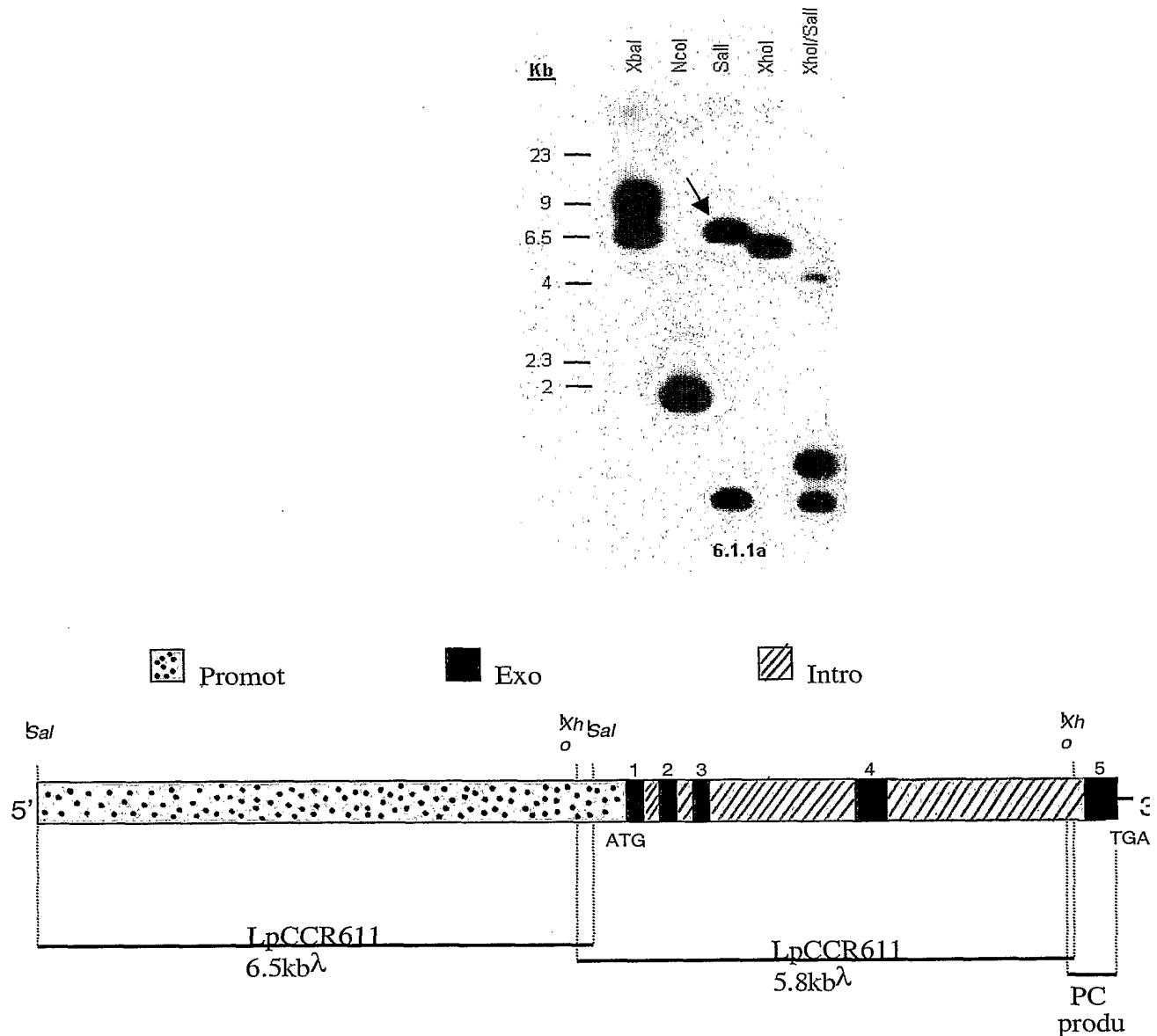
A

FIGURE 23

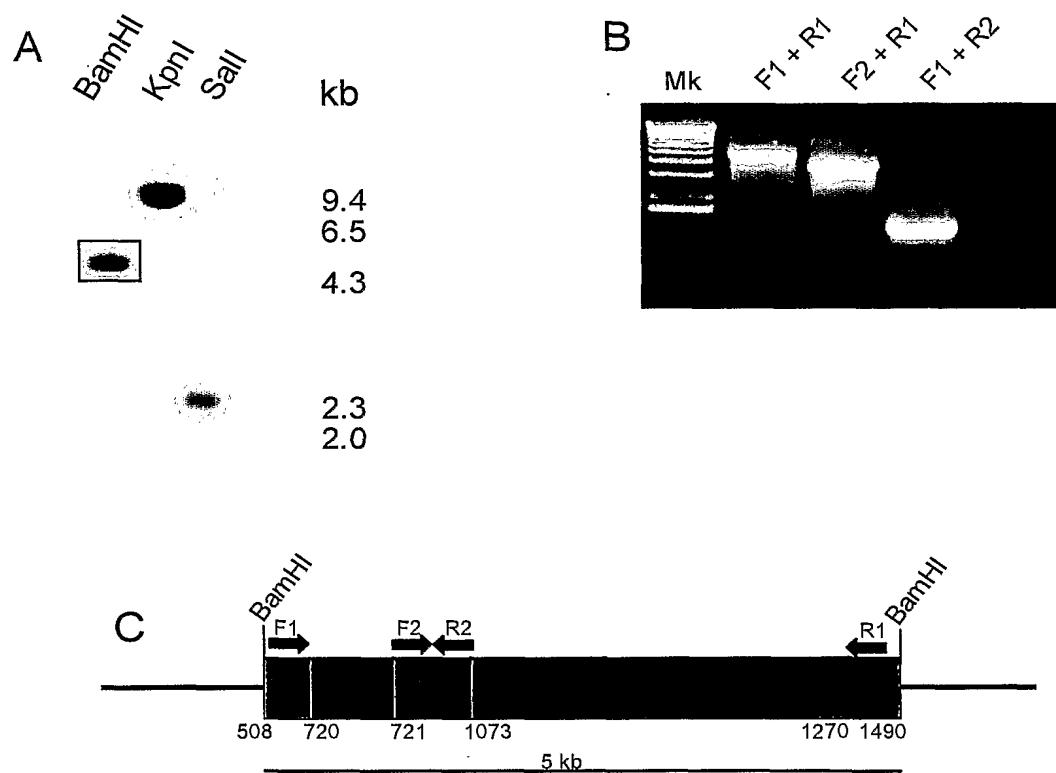


FIGURE 24

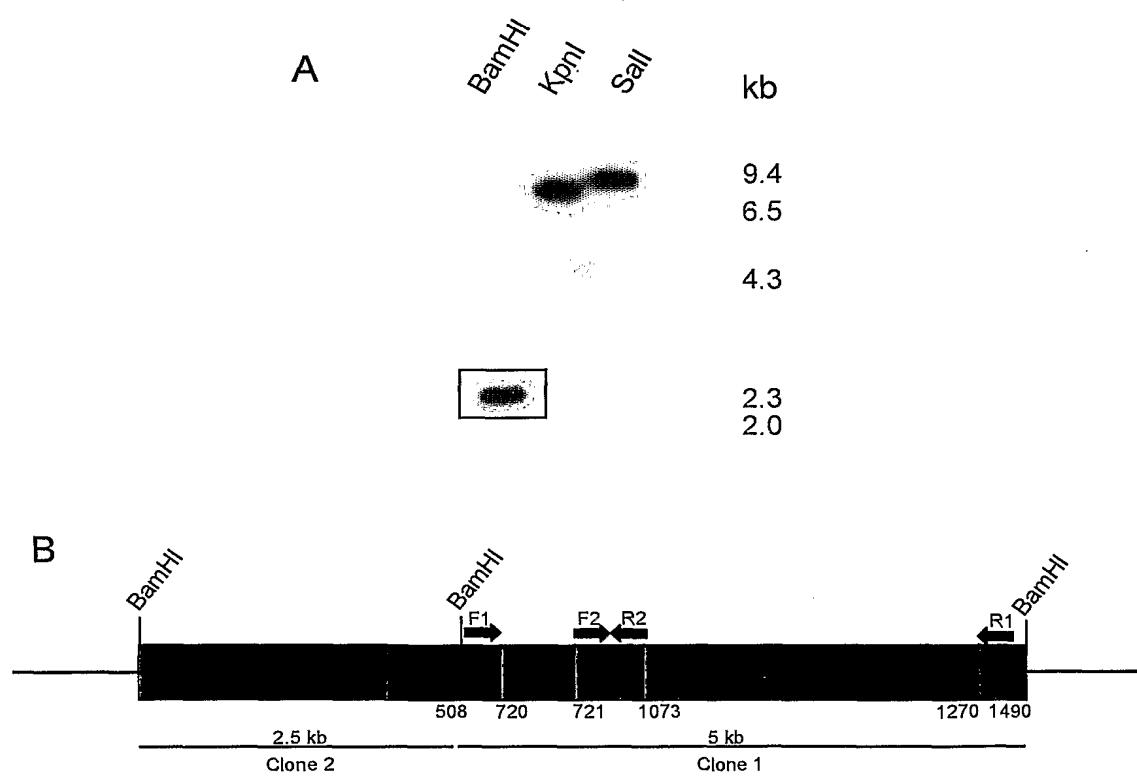
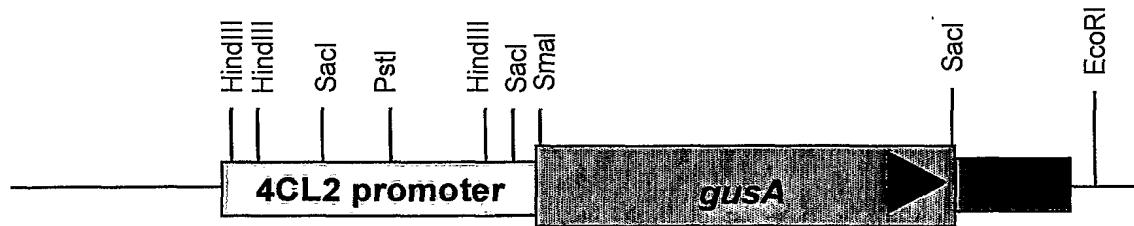


FIGURE 25



1	TCCCGTATCTTCAACGTGACACCCCTACACTTCCTGCTTGTCTGGAGATTTACACACAC	60
61	ACGGCAATTACCAGGAGTATCTCCTAGATTATTTTTTCGATAAGGATCTTCCAGATAT	120
121	AGCATGTGAATCTCTGTACTACTGTTGTCAAGCAAAATTAAACATTGACATCAGTGT	180
181	TTTTGTTGGGGCAGCGGAATCTTGACGCCCTTGCCTCTCAAGACATGTCACCC	240
241	CACTAGTTAGTGTGCCAGCTGGTAGTACTACGTACGATGCTCCCTCCCTCCGTAATTATT	300
301	CAACCTTTTGCTCTCTTTATAAAGTCAAACCTTTAAATCTGACCAGATATCTGC	360
361	TAAAAAATTAGCAGACATGCATACATCAAAGCAGTAGTCCTCCCTCCGTTAAAATTACC	420
421	TGGGTTTATTCAAATAAAGTCAAACTCTGTAAAATTCAATTAAATATTAGAAAAATCTA	480
481	ACAGCACCTGTAGTATAAAAGTATGCTCCCTCTGTTGTAAAAAGCTAAGCAACTTTT	540
541	TGAGATACGGATAAATCTTAGCTAAAACATGTCTATATACCTTGTATCTAGATAAAGT	600
601	TGGAAAGCTTTTAGAAACAGACAAAGTATGTGTTGACATTATGAATGTTGAGTATT	660
661	TTCCTCTAATCTTGATCAAATTACAAATTGGCTTGAATAGAGGGACCATTATTAGT	720
721	ATGAAACTACATAAATTGTAAAACACTCAACATAATTACGATGGTCAGTGATAGCAC	780
781	TAACCTAGCTTTCATAAATGCCACTGCTTCAATAGAGCATGAAGCAGGACAAATTAA	840
841	TTCGTGTGACTTGAATAGAGGGAGCCTGTTCTGGTTCAACTCACCCCTGCATGTGTGTCTT	900
901	CATCCCTTTGCTCTTCCATCTGTGGTGTCAATTGAGTGTCCCACGTGCATGTGGCGA	960

FIGURE 26

961	AACTTGAACTAGAAATTGACATGCTCCACTGCCGGAGCGGAGTATCTTGTGCTTG	1020
1021	TTACCCATTATTGTTGCTACGTACTACAGTGTAGATTGAACTTCATAATCAAAGAAC	1080
1081	TTAGTTCTACAATTGGCTAACATATAATGAGCAATCAAACCTCTATATCTGTG	1140
1141	GCAAATAACTAACATTAGTTACAGTTAGATGCCAGACGCCAGTGTCTTCCCCCTT	1200
1201	TTCGGAAAAAGCTATTCCATAATAAGTGTGGAAATTAAATAATGGTACTACGAATT	1260
1261	TGAAAAAAAAGTGTCAAAAATTCACTAACGAAAGTACGTAGTACAAATTAAACTAAGAT	1320
1321	TCCGACACTTATTAGGATCGGAGAGAGTAAGTAGCAAACACTACTCCATCCACCTAAAA	1380
1381	CACGTGATTAACTTTGTCTAGATACGGATAGAAAGTTGGGATACATCCGTATCTTAAAA	1440
1441	AAAAACGCACTTATTAGACGAAGGAGGGAGTATTCAACCTTGATTAAACGGAATC	1500
1501	TACAAAGGAAATACATGGATTGTACAAGTGGCTGACCGTATCCATTATGTACTCGTACT	1560
1561	TTGCAGTTGAAAGCAAAGGCTAGTGTAAATTGTAGGTGGTCTAGGCGTCTAGCTGTT	1620
1621	CATGGCGTTATCACAGCCGTGCCAGTGTGCTCAGGGCCGTACATAAGTTGCTTGGGTAT	1680
1681	GTGTCGATCTAGGATTGCGTCTTACAATTGCTTCCAATTATTCTGTAAAGAG	1740
1741	ATCGATGTGAACTTCTCTGCGAGTAAACTGAAATTGTCTGAATAATATAACTCGGCAG	1800
1801	ATTATGTTTATCGTTGCGTAAACAGGCTACACAAATTGCTCGAGTCAGCAGCGAG	1860
1861	TTGAGCTACAACGAATCCATCAGAAAAACTATACTATAGTAGCACATCGTTCTT	1920

FIGURE 26 CONTINUED

40/76

1921	TTTCATGACGTTCTGTTCTTCCCTAACTTCCAGGAGCACCGGAGACGACGATGTGGTG	R S T G D D D V V	1980
1981	ATAAAGATTTGTAUTGCCACTCTGACCTGCACGCCCTGAAGAACGACTGG	I K I L Y C G I C H S D L H A L K N D W	2040
2041	AAGAACTCAAGGTACCCGATGATCCCCGGCACGAGATCGCCGGCGAGGTCACGGAGGTG	K N S R Y P M I P G H E I A G E V T E V	2100
2101	GGCAAGAACGTGAGCAAGTTCAAGGCCGGCACCGCGTGGCGTCGGGTGCATGGTGAAC	G K N V S K F K A G D R V G V G C M V N	2160
2161	TCGTGCCGGTCTGCGAGAGCTGCGACAAGGGCTTCGAGAACCACTGCCCGGGCATGATC	S C R S C E S C D K G F E N H C P G M I	2220
2221	CTCACCTACAACCTGGTCGACGTCGACGGCACCGTCACCTACGGCGCTACTCCAGCATG	L T Y N S V D V D G T V T Y G G Y S S M	2280
2281	GTGGTGGTGCACGAGCGGTTCTGGTCCGGTCCCCGACGCCATGCCGCTGGACAAGGGC	V V V H E R F V V R F P D A M P L D K G	2340
2341	GCGCCGCTGCTGTGCGCCGGCATCACCGTGACAGCCCCATGAAGTACCAACGGGCTAAC	A P L L C A G I T V Y S P M K Y H G L N	2400
2401	GTTCCCGGGCTGCACCTCGGCGTGCTGGGCTGGCGGGCTGGGCCACGTTGCGGTCAAG	V P G L H L G V L G L G G L G H V A V K	2460
2461	TTCGGCAAGGCCTTCGGAATGAAAGTGACGGTGATCAGCTCGTCGCCGGGAAGAAGGAG	F G K A F G M K V T V I S S S P G K K E	2520
2521	GAGGCCCTGGGCGGCTGGCGCCGACCGTTCATCGTCAGCAAGGACGCCGACGAGATG	E A L G R L G A D A F I V S K D A D E M	2580
2581	AAGGTAGGCGGACCCGCTGGTTCAAGTTACTTCCCTGTCGGTGCAGAACGAAAGAGGAA	K	2640

FIGURE 26 CONTINUED

G at 851 bp (coding sequence) missing from cDNA in cv Ellett

2641 CTTGAGGGTTCATGTTGTTGCGTTGGTATGTCAGGCTGTGAGCACCAT 2700
 2641 -----+-----+-----+-----+-----+-----+
 A V M S T M

2701 GGATGGCATCATAAACACGGTATCTGCAAACATCCCCCTGACCCCTCTTCGGGCTGCT 2760
 2701 -----+-----+-----+-----+-----+-----+
 D G I I N T V S A N I P L T P L F G' L L

2761 CAAGCCCAACGGCAAGATGATCATGGTCGGCCTCCCCGAGAACGCCATCGAGATTCC 2820
 2761 -----+-----+-----+-----+-----+-----+
 K P N G K M I M V G L P E K P I E I P P

2821 CTTCGCTCTAGTTGCCAGTAAGTCTTAGGATCTCTTGAATAAGGAGAAATCATGCACTG 2880
 2821 -----+-----+-----+-----+-----+-----+
 F A L V A

2881 ATCGATCAGAGAAATGAGATAGCATCCTGATGAACATTGTACGTGTGCAGCGAATAAG 2940
 2881 -----+-----+-----+-----+-----+-----+
 N K

2941 ACCCTGGCCGGGAGCATCATCGGCGGATGAGCGACACGCAAGGAGATGCTGGACCTCG 3000
 2941 -----+-----+-----+-----+-----+-----+
 T L A G S I I G G M S D T Q E M L D L A

3001 GCGAACGACGGCGTGACGGCGACATCGAGGTGGTCGGCGGGAGTATGTAAACACGGCC 3060
 3001 -----+-----+-----+-----+-----+-----+
 A K H G V T A D I E V V G A E Y V N T A

3061 TTGGAGCGCCTTGCCAAGAACGACGTCAGGTATCGCTTCGTATCGACATCGAACACC 3120
 3061 -----+-----+-----+-----+-----+-----+
 L E R L A K N D V R Y R F V I D I G N T

3121 CTCGACAAGGTTGCGGCCACCACCGAGTGAAACGTACTCAGCACTGCTTACGATCTACGTT 3180
 3121 -----+-----+-----+-----+-----+-----+
 L D K V A A T T E *

3181 GTTCCACTGTTAGCTCCGTAGTAAACAATAACGATCAAACCTTGTATCTGGTC 3240
 3181 -----+-----+-----+-----+-----+-----+
 .

3241 ATTGGTGTAGACATGGTTGGCGAGGAAACTGAGTTGAAGGATGGATGGATAAGTTG 3300
 3241 -----+-----+-----+-----+-----+-----+
 .

3301 CTTCTGCCGTGTTAATGGATTACCTACTTAGCTCACTGCAATTAAACAAATTAAAGAAC 3360
 3301 -----+-----+-----+-----+-----+-----+
 .

3361 GACACACCCAAAAGACTTTCGTCAGTTCTGGATTATACAAGTCGTTATGGTTGGGTG 3420
 3361 -----+-----+-----+-----+-----+-----+
 .

FIGURE 26 CONTINUED

FIGURE 26 CONTINUED

4381 GAATTTGTACCCCTCCGAGAAATTGCTAAAATGATGGAGTGACCTACAACGAGCCTGGAT 4440

4441 ATGTGAGTTCTTCTTGCCCCATTGCACAAAAATTGTAAATATTAGGGTTACTGGATCCA 4500

A) CTAGTTCTAGAGCGGCCACCGCGGGGAGCTCCAGCTTTGTTCCCTTAGTA 4555

FIGURE 26 CONTINUED

1	GGCACGAGTCGCCTCCAACGTCTTCCCTAACCGGCCGTCCCTACGCtTGCA	CCACCACC	60
61	ACGCACAGACAGAGCAGTTCCCAGCCCCGCCGGAACCGGATGGCACCCACGGCGGG		120
		M A P T A A E	
121	AGCAGACGGAGCACCACCAAGCACACCAGGAAGGCAGTGGGCTGGCGCGCGACGACG		180
	Q T E H H Q H T R K A V G L A A R D D A		
181	CCGGCCACCTCTCCCCGCTCGCCATCACACGGAGGAGCACAGGAGACGACGATGTGGTGA		240
	G H L S P L A I T R R S T G D D D D V V I		
241	TAAAGATTTGACTCGGAAATCTGCCACTCTGACCTGCACGCCCTGAAGAACGACTGGA		300
	K I L Y C G I C H S D L H A L K N D W K		
301	AGAACTCAAGGTACCCGATGATCCCCGGGCACGAGATCGCCGGCGAGGTACGGAGGTGG		360
	N S R Y P M I P G H E I A G E V T E V G		
361	GCAAGAACGTGAGCAAGTTCAAGGCCGGCACCGCGTGGCGTCGGTGCATGGTGAAC		420
	K N V S K F K A G D R V G V G C M V N S		
421	CGTCCGGTCTGCGAGAGCTGCGACAAGGGCTTCGAGAACCACTGCCGGGCATGGTCA		480
	C R S C E S C D K G F E N H C P G M I L		
481	TCACCTACAACCTCGGTCGACGTCGACGGCACCGTCACCTACGGCGGCTACTCCAGCATGG		540
	T Y N S V D V D G T V T Y G G Y S S M V		
541	TGGTGGTGCACGAGCGGTTCTGGTCCGGTCCCGGACGCCATGCCGCTGGACAAGGGCG		600
	V V H E R F V V R F P D A M P L D K G A		
601	CGCCGCTGCTGTGCGCCGGCATCACCGTGTACAGCCCCATGAAGTACCAACGGGCTCAACG		660
	P L L C A G I T V Y S P M K Y H G L N V		
661	TTCCCGGGCTGCACCTCGGCGTGGGCTGGCGGGCTGGGCCACGTTGCGGTCAAGT		720
	P G L H L G V L G L G G L G H V A V K F		
721	TCGGCAAGGCCTCGGAATGAAAGTGACGGTGATCAGCTCGTCGCCGGGAAGAAGGAGG		780
	G K A F G M K V T V I S S S P G K K E E		

FIGURE 27

781 AGGCCCTGGGGCGGCTGGCGCCGACCGTTCATCGTCAGCAAGGACGCCGACGAGATGA 840
 -----+-----+-----+-----+-----+-----+
 A L G R L G A D A F I V S K D A D E M K
 G at 851 bp (coding sequence) missing from cDNA in cv Ellett
 ▼
 841 AGGCTGTGATGAGCACCATGGATGGCATCATAAACACGGTATCTGCAAACATCCCCCTGA 900
 -----+-----+-----+-----+-----+-----+
 A V M S T M D G I I N T V S A N I P L T
 901 CCCCTCTCTCGGGCTGCTCAAGCCAACGGCAAGATGATCATGGTCGGCTCCCCGAGA 960
 -----+-----+-----+-----+-----+-----+
 P L F G L L K P N G K M I M V G L P E K
 961 AGCCCATCGAGATTCCCTCCCTCGCTCTAGTTGCCACGAATAAGACCCCTGGCCGGAGCA 1020
 -----+-----+-----+-----+-----+-----+
 P I E I P P F A L V A T N K T L A G S I
 1021 TCATCGGCGGATGAGCGACACG CAGGAGATGCTGGACCTCGCGCGAAGCACGGCGTGA 1080
 -----+-----+-----+-----+-----+-----+
 I G G M S D T Q E M L D L A A K H G V T
 1081 CGGCCGACATCGAGGTGGTCGGCGCGGAGTATGTGAACACGGCTTGGAGCGCCTGCCA 1140
 -----+-----+-----+-----+-----+-----+
 A D I E V V G A E Y V N T A L E R L A K
 1141 AGAACGACGTCAGGTATCGCTTCGTATCGACATCGGCAACACCCCTGACAATGTTGCGG 1200
 -----+-----+-----+-----+-----+-----+
 N D V R Y R F V I D I G N T L D N V A A
 1201 CCACCACCGAGTGAACGTACTCAGCACTGCTTACGATCTACGTTGTTCCACTGTTAGTGC 1260
 -----+-----+-----+-----+-----+-----+
 T T E *
 1261 TCCGTAGTAAACAATAAACGATCAAAACTCTTGTATCTGGTGCATTGGTGTAGACATGG 1320
 -----+-----+-----+-----+-----+-----+
 A) TTGTTTGCAGGAAACTGAGTTGAAGGATGGATGGATAAAAAAAAAAAAAAAA
 1 GGCACAGAGTCGCCTCCAACGTCTTCCCTTAACCGGCCGTCCCTACGCTTGCAACCACCC 1378
 -----+-----+-----+-----+-----+-----+
 61 ACGCACAGACAGAGCAGTTCCAGCCCCCGCCGGAACCGGATGGCACCCACGGCGCGG 120
 -----+-----+-----+-----+-----+-----+
 M A P T A A E
 121 AGCAGACGGAGCACCAACCAGCACACCAGGAAGGCGGTGGGCTGGCGCGCGACGACG 180
 -----+-----+-----+-----+-----+-----+
 Q T E H H Q H T R K A V G L A A R D D A

FIGURE 27 CONTINUED

181 CCGGCCACCTCTCCCCGCTCGCCATCACACGGAGGAGCACAGGAGACGACGATGTGGTGA 240
 G H L S P L A I T R R S T G D D D D V V I

 241 TAAAGATTTGTAUTCGCGAATCTGCCACTCTGACCTGCACGCCCTGAAGAACGACTGGA 300
 K I L Y C G I C H S D L H A L K N D W K

 301 AGAACTCAAGGTACCCGATGATCCCCGGGCACGAGATCGCCGGCGAGGTACGGAGGTGG 360
 N S R Y P M I P G H E I A G E V T E V G

 361 GCAAGAACGTGAGCAAGTTCAAGGCCGGCACCGCGTGGCGTCGGGTGCATGGTGAAC 420
 K N V S K F K A G D R V G V G C M V N S

 421 CGTGCCGGTCTGCGAGAGCTGCGACAAGGGCTTCGAGAACCACGTGCCGGCATGATCC 480
 C R S C E S C D K G F E N H C P G M I L

 481 TCACCTACAACCTCGGTCGACGTCGACGGCACCGTCACCTACGGCGGCTACTCCAGCATGG 540
 T Y N S V D V D G T V T Y G G Y S S M V

 541 TGGTGGTGCACGAGCGGTTCTGGTCCGGTCCCGACGCCATGCCGCTGGACAAGGGCG 600
 V V H E R F V V R F P D A M P L D K G A

 601 CGCCGCTGCTGTGCGCCGGCATCACCGTGACAGCCCCATGAAGTACCAACGGGCTCAACG 660
 P L L C A G I T V Y S P M K Y H G L N V

 661 TTCCCGGGCTGCACCTCGGCGTGCTGGGGCTGGGGCTGGCGGGCTGGGCCACGTTGCGGTCAAGT 720
 P G L H L G V L G L G G L G H V A V K F

 721 TCGGCAAGGCCTCGGAATGAAAGTGACGGTGATCAGCTCGCCGGGAAGAACGGAGG 780
 G K A F G M K V T V I S S S P G K K E E

 781 AGGCCCTGGGGCGGCTGGCGCCGACCGTTCATCGTCAGCAAGGACGCCACGAGATGA 840
 A L G R L G A D A F I V S K D A D E M K

 G missing at 851 bp in the cDNA isolated from cv Ellett
 resulted in a premature stop codon (truncated CAD2)

 841 AGGCTGTGATAGCACCATGGATGGCATATAAACACGGTATCTGCAAACATCCCCCTGAC 900
 A V I A P W M A S *

FIGURE 27 CONTINUED

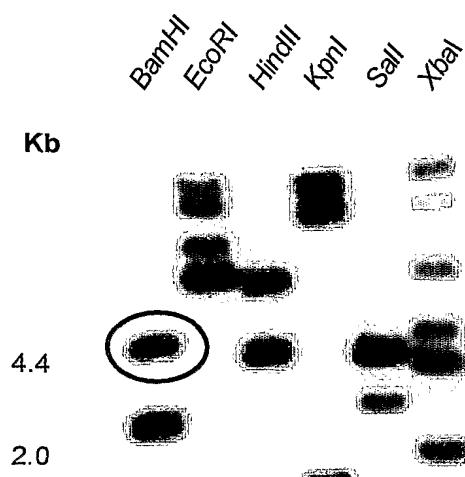
47/76

901	CCCTCTTCTGGCTGCTCAAGCCAAACGGCAAGATGATCATGGTCGGCTCCCCGAGAA	960
961	GCCCCATCGAGATTCCCTCCCTCGCTCTAGTTGCCACGAATAAGACCCCTGGCCGGAGCAT	1020
1021	CATCGGCGGCATGAGCGACACGCAGGAGATGCTGGACCTCGCGCGAAGCACGGCGTGAC	1080
1081	GGCCGACATCGAGGTGGTCCGCGCGAGTATGTGAACACGGCCTGGAGGCCCTGCCAA	1140
1141	GAACCGACGTCAGGTATCGCTTCGTATCGACATCGCAACACCCCTCGACAATGTTGCC	1200
1201	CACCAACCGAGTGAACGTACTCAGCACTGCTTACGATCTACGTTGTTCCACTGTTAGTGCT	1260
1261	CCGTAGTAAACAATAAACGATCAAACCTTGTATCTGGTGCAATTGGTAGACATGGT	1320
A)	TGTTTGCAGGAAACTGAGTTGAAGGATGGATGGATAAAAAAAAAAAAAAA	1377

FIGURE 27 CONTINUED

FIGURE 28

A



B

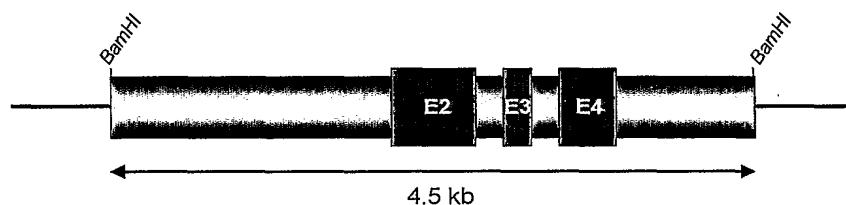
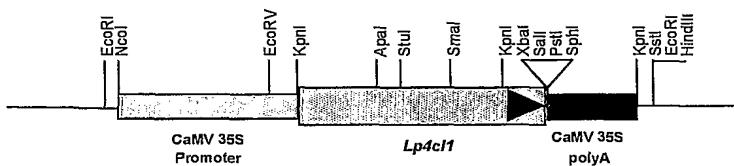


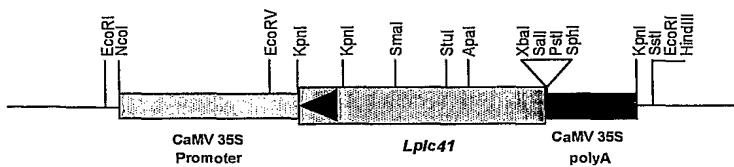
FIGURE 29A

A)

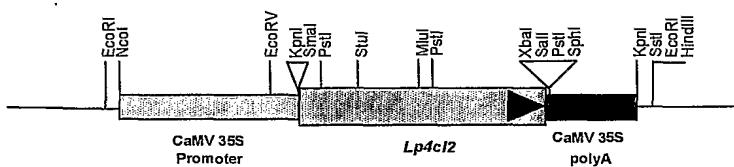
p35S4cl1



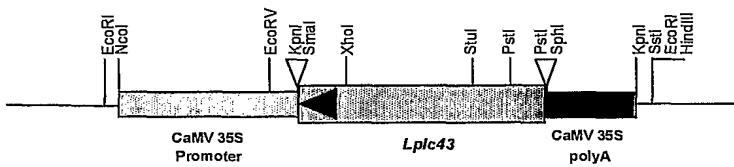
p35S4cl1



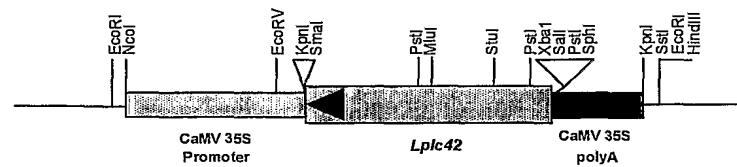
p35S4cl2



p35S4cl3



p35S4cl42



p35S4cl3

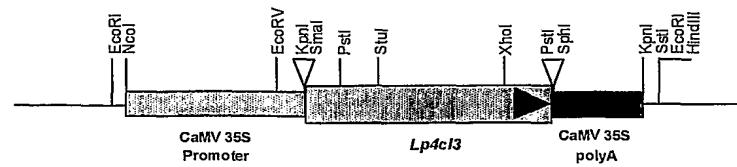
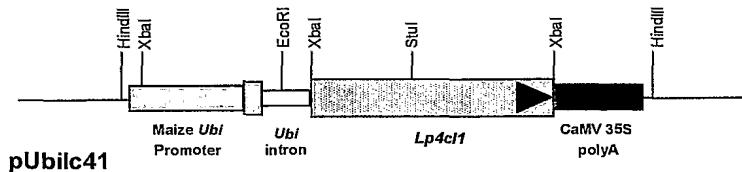


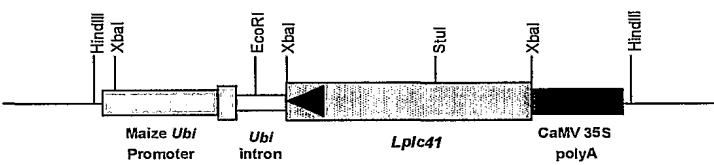
FIGURE 29B

B)

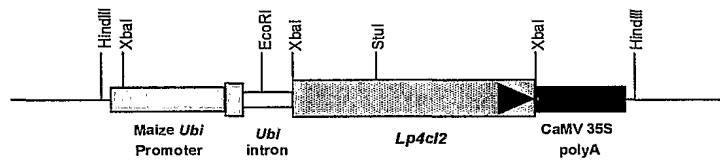
pUbi4cl1



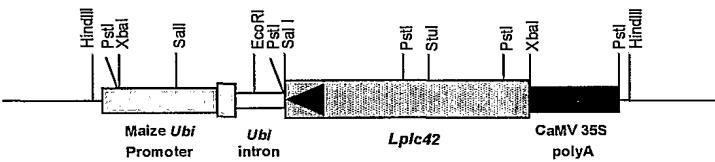
pUbilc41



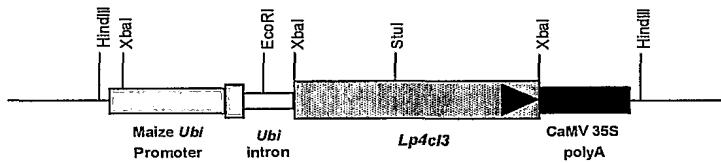
pUbi4cl2



pUbilc42



pUbi4cl3



pUbilc43

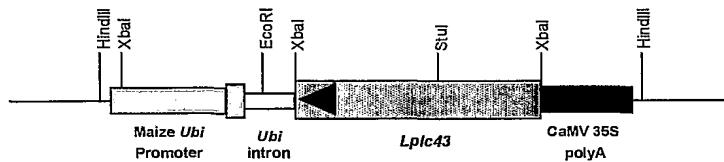
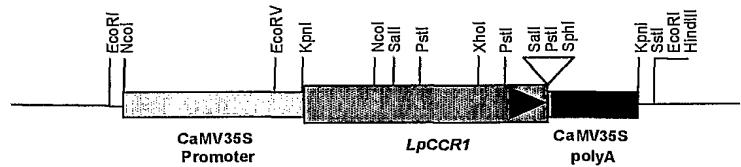


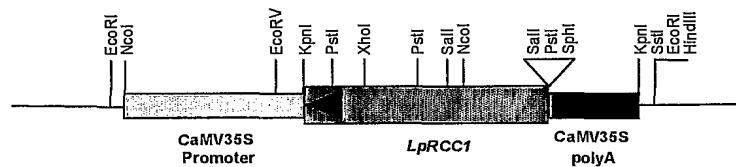
FIGURE 30

A)

p35SCCR1



p35SRCC1



B)

pUbiCCR1

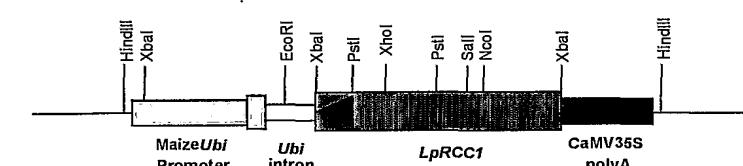
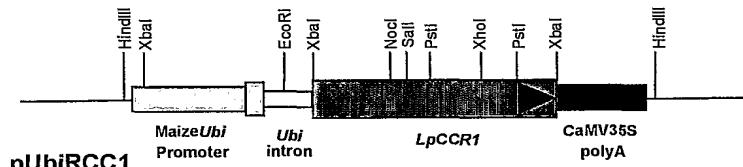
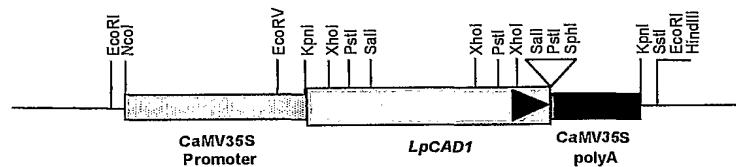
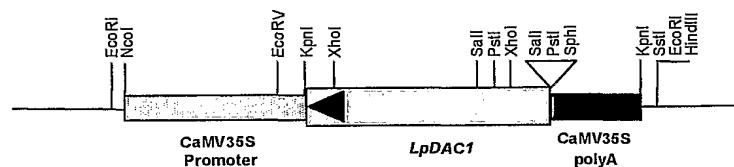


FIGURE 31

A)

p35SCAD1**p35SDAC1**

B)

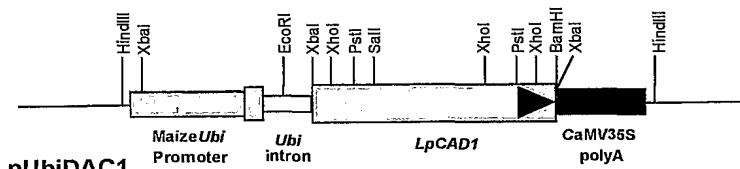
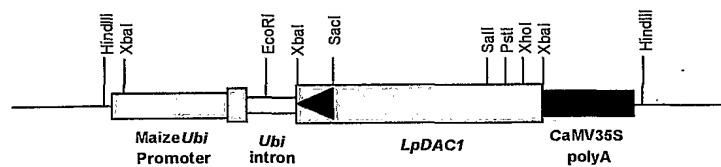
pUbiCAD1**pUbiDAC1**

FIGURE 32

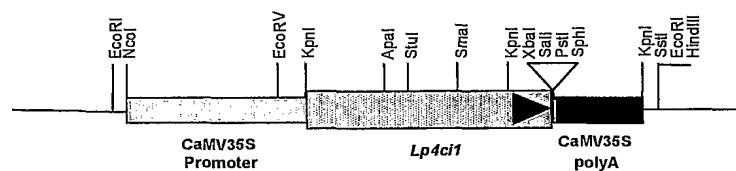
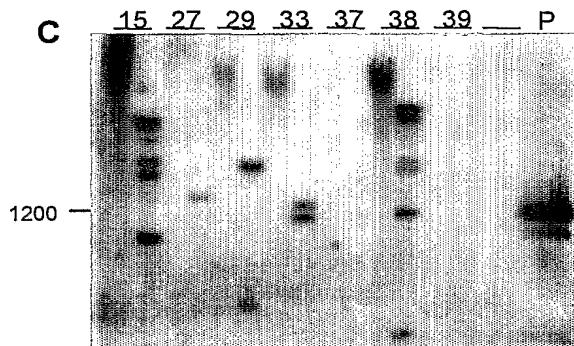
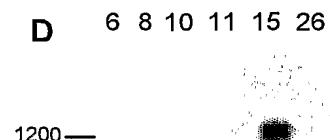
A**B****C****D**

FIGURE 33

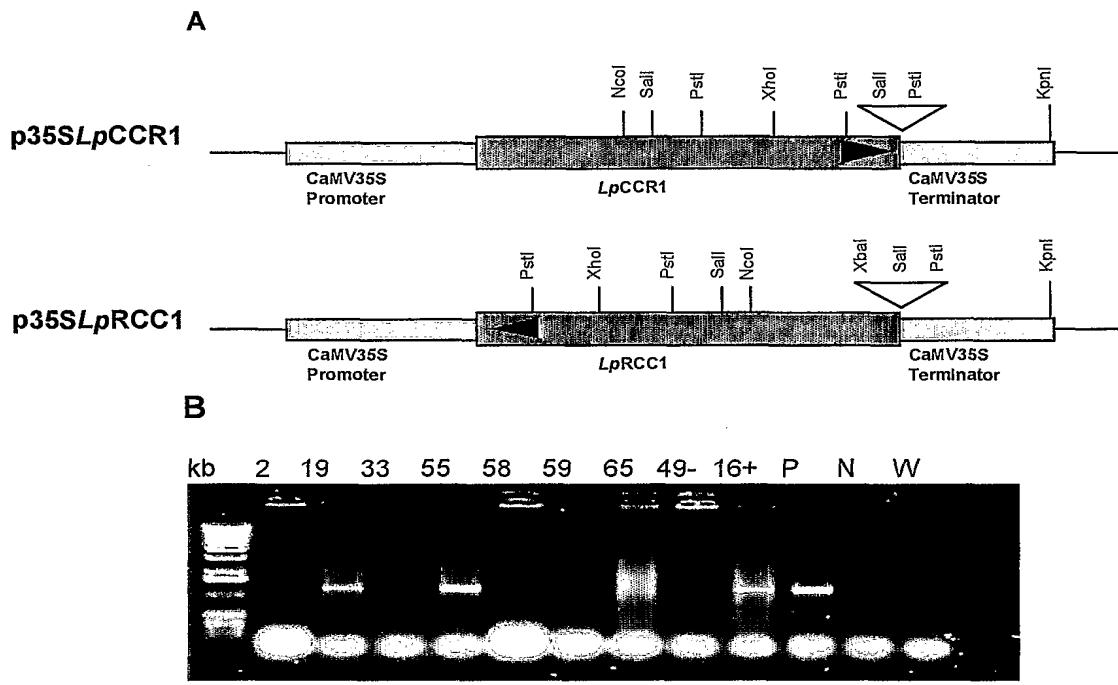
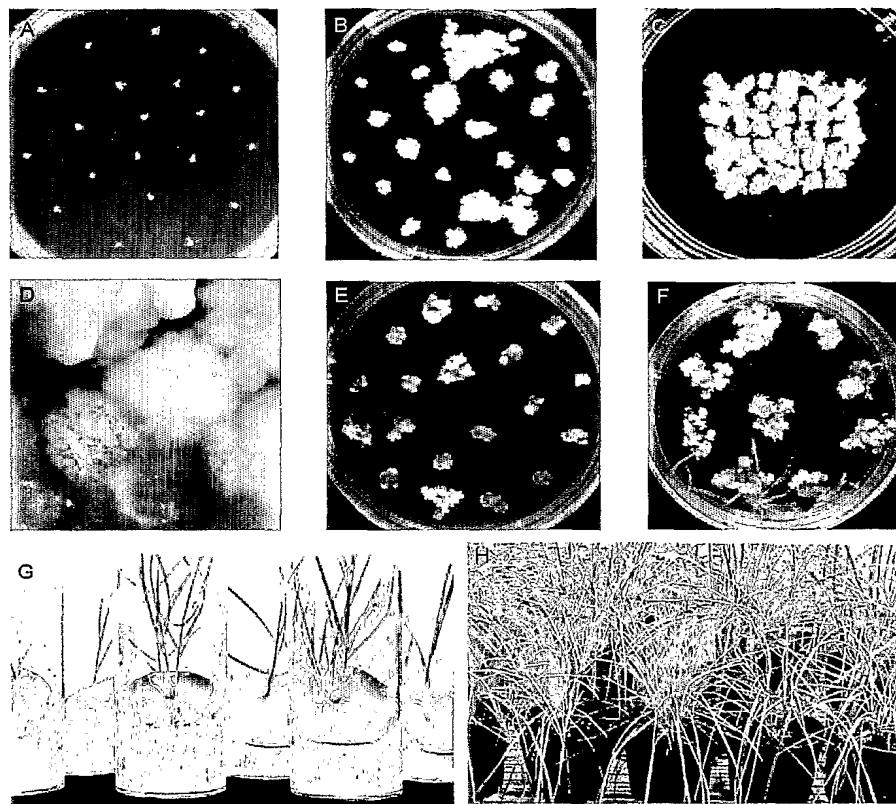


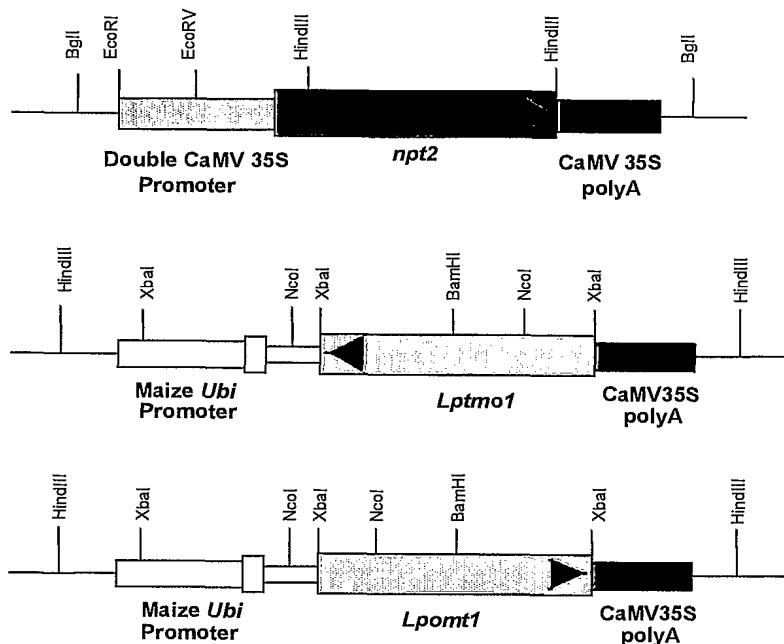
FIGURE 34



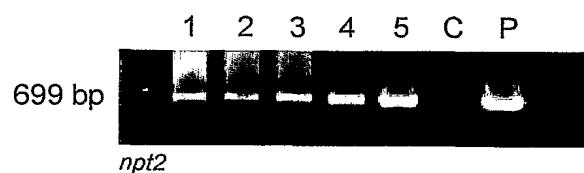
56/76

FIGURE 35

A

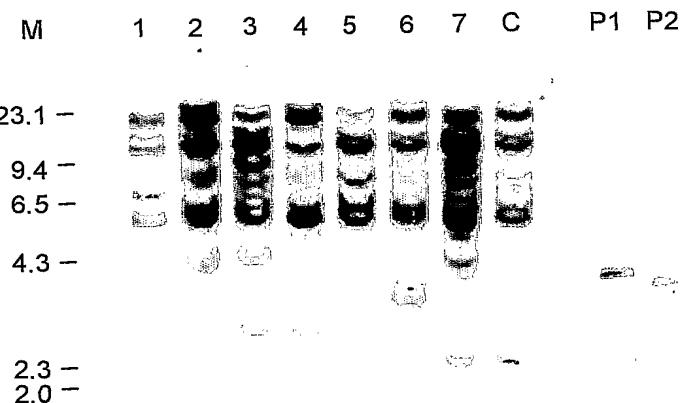


B

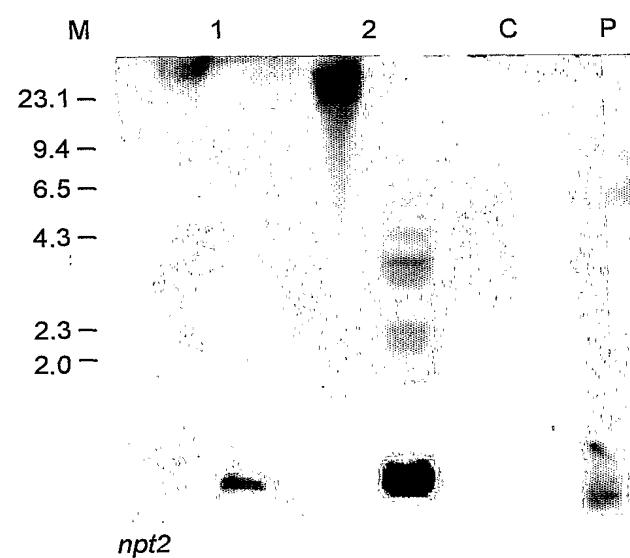


57/76

C



D

FIGURE 35
CONTINUED

E

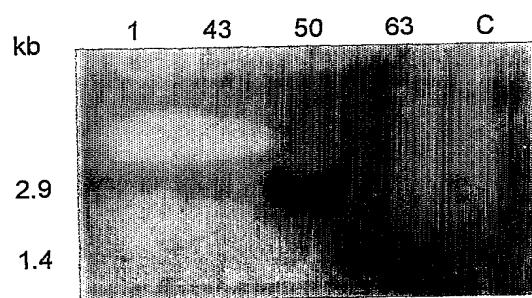


FIGURE 36

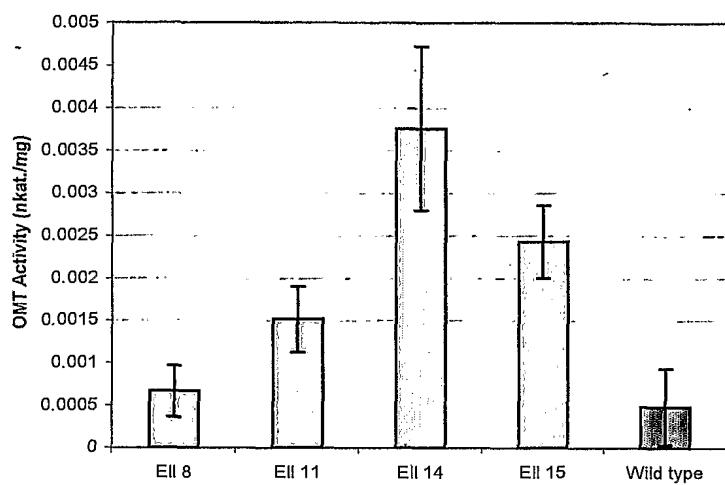
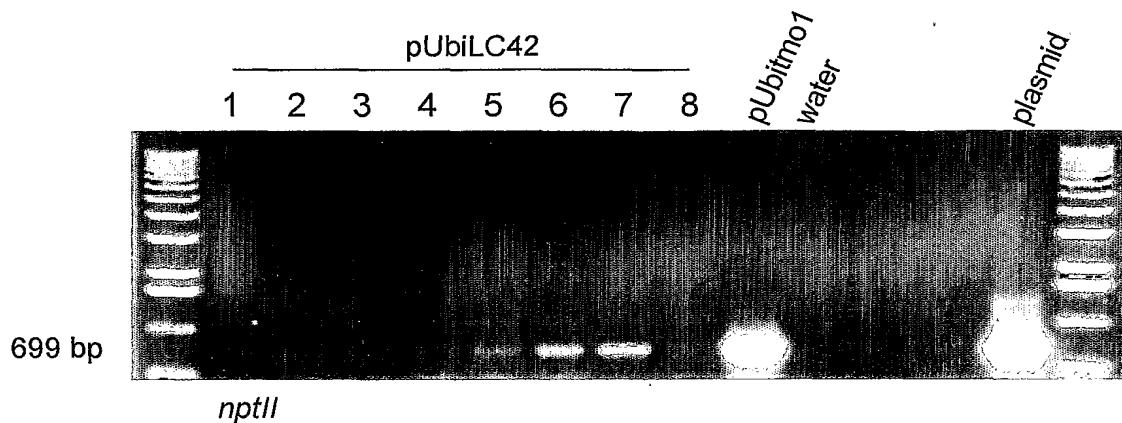


FIGURE 37



60/76

-2206	CGGGATCAACTGGATGTCCTTGCGGGCACGGTTCAGGAACAAACGACACATGCAGCAG	-2147
-2146	GGATCTCCTCCAAAGACTCACACAAAGGTGACATGAGCGCCCGCTTTTGAAGCCAAGT	-2087
-2086	TGGCTAAGAAATCGCAAAGCTTGGTAGTCGGCACCTCAGGATCTGCAACAAAAGGCA	-2027
-2026	CCAAGGGAGCTGCCAACACATCAACCACAAACATCATGTTCAAACGCAGTCTCCTCAAGCC	-1967
-1966	TCGAATGCTCAACCGAAAGAGAGGGCAGAAGCTCAACAAAAAACTCAGCCAACCCAAAGC	-1907
-1906	CCTCGACGTCATCAGAGATTAGGCTCTGAGGACCCGCAGGGAAAGCAACCTGTCAACAAAC	-1847
-1846	CGCATCCGGCAGAAAAGGAGCAAGACCGGAGCAACCCCTCAAGAGGCACACGAAAGACGTC	-1787
-1786	GAAGCCAAGAGGAGACGAGTCGCAGGGACGGCGGACAGGCAGAGGGGCCGTAGAACTC	-1727
-1726	CAAGAGCTGGCGTCCCTCGACCTAGCATCCGAAGCACTGACCGGGCACTCAATGCATA	-1667
-1666	ACTTTATCTTGATGGCATATGTACTCAAACCCATAACATGTTACCATGCATTATCTATG	-1607
-1606	GAACATTCTCATATACAACCTCTGAGTGGTCAGTCAGGAATTTCATTAAACACC	-1547
-1546	AAAAACATAACTGGGGCCTACACACACTTCACAGCATGGAAAATTGTTAGCTTTTAA	-1487
-1486	AGAGTTGCAAAATCTGTCAAGCGAATGTTCTGTGATAATTGGAACGAAGCATGTTCCC	-1427
-1426	CATTTCAATGTGTCTCTTACCCCTAACTAGCACCCGACCAACAAATCTGACCACCT	-1367

FIGURE 38

-1366	AGTTATATCATCATAGAGACCCACATGTAGGTTGACCCCCATAACACTTGTGTGGATATC	-1307
-1306	ATGGAAAATGGCCTTGATCAACACTTCTTCCTACTTGGTACAAATGGTTATGGACTTA	-1247
-1246	CTCAATTAGTGCTTAGAGAGCTTGGCTGCAGACTTGAGCTTCCAATATTCATAGG	-1187
-1186	TCCCTCCGGAGTGGGCAGCCCCATCTACATAGGCTCAAAACCAGATTTGTAACATGTT	-1127
-1126	AGACACTTTCAACTTCATCATAGACCATCAAGGAGCTGGCATGTGACAGTGATATATGTA	-1067
-1066	TCAATTACCCATTCAACACGAATAGCTTGCTCATGCATGGTAGTCTTGCGGGCGGGCGGG	-1007
-1006	CGGGACCACATCGAACACACCGCCGGCGGTCACTAGGGCTAGGGTTAGATAAAATCTAGCCG	-947
-946	TTTCATTCAAACTTGTGATATATAATCAAATTAAATAACCTTATTTCGTGCAT	-887
-886	TTTTATTTATTGAGGGCGTGTGTTGGGGACACGGCTGGAAAGTGACATCCCCAACACT	-827
-826	GCACGAAGAAAACCGTCGCCAAAAATTGATCCGGCGTCAGTCCTTGGAGACGATT	-767
-766	TGGATGACGGCTAGAGATGCTAAAGTTCTCCACGCCATGTTCTTCTATATATACA	-707
-706	CACAGCCCAAGGTCCATGAAAAGTAAACGGCACCGACAGCACCGACCGGACAACTTCA	-647
-646	CATTACGGCACATCGCTATTACGGACCACATACAACCTCCACCGCTATTCTCAGCCAAGTC	-587
-586	ATACATGACATGATCCAATGGACGACTTGTGAGCGAAACTAGAACCTTGGGGTTAG	-527
-526	ATTTTCCAATGTGGATAAGTTGTACGCGCCGACTAGCTTACACTTGGTTGAAAAAAGCT	-467

atactcctactgtatattggtaaaacaaaacatttcttttatttgcataaggagtgcgtcaaattaaagtctttgtca
 ttttcaaaaggaaaaaaaaacacccattaccactcttctccatgcatttttaccaaaggttgttctgtcaaa
 tgaacatataatagttcggtctatgtcagtgcattaccggccactagctagtaggactgcctatgtccagcaaattgt
 ctatgtggaccggagtgccaaaaggagccaattatgttagg
 gttgcaagcgggatcacacaaaaggcctcgcccttagttcaattaagtggtaacttctcagggaccccccgtca
 actctaccatcacatccgtccaaataaaagctagcatcagcaccagatttagtactccctccgttttatttagtgcga
 ttctaggttcagccaaagtctacatttgcattaaacgatttataacatagattttttattgcataatgtcaatatttttca
 catataatataaaggataaaacccatttgcattaaacgatttataacatagattttttattgcataatgtcaatatttttca
 catataaaattataaagggttactttgacttgcattaaacccatttgcattaaacgatttataacatagatggccaaaaggca
 catatggcaggagtaacatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 aatcgtgcacgtggcaatattatctaccgtcgccgtccagttccatgcatttttgcatttttgcatttttgcatttttgcatt
 gccggagctgcccgtgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 cttttttgcgaaggatatagttagtattacttctctgtatcacaaggaaactgtattgtgttgcatttttgcatttttgcatt
 cagaattctgtcatgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 tcatccatgtctgttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 ttttttttattaaattcagaactttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 cggcgtggacccttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 gccccctgtcagccacgggttaacccggcgtcgatgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 acggccgtcagccgtacgtggacgtccgcacgtggccgcacgcggccacccgttgcgttgcatttttgcatttttgcatt
 ccacccctgtccgcgcgttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 accagggtacgcgtacgcctgttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 tactacttgcgttaactattgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 gtccagtgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 tgggactaagcaaaactgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 agggtggatagttacttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 tgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 acagacacacacaaggccgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 agttcgaggttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 aagtaggtggacttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 tatccgcacggcagaaacgcgttagcatcaggccagaaagg
 agcgtgcgtgatatgttaacccagacggctttccatctggctacccgcataactacccgcgtgtgc
 gctgaccaatttgcgtcaccatgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 aacctcaagtgtgcactctgttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 gaccaacacacacatgttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 aaggcaggagaacatcttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 acacatactacccgttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 aacggagggagttatcgatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 gtgatacgtgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 tttcataccgaatgttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 aatgccacccatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 gtctggtttaagtcaaaatttttagactactaaaacacagaagtttgcatttttgcatttttgcatttttgcatt
 tcgaaatgcgtgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 gaacggctgtgttttagtcaacattatgttaggttcat
 gtgatacttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 catgtcaaatgtcgttacttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 caagatcaacgttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 tcgcaatcttgcatttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 caaggaaacgggaggaaatcttgcatttttgcatttttgcatttttgcatttttgcatt
 ctggaaatcttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 cagcatgttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 acaggtgttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 caggccgttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 gaggccgacaaaggaaacccatttttgcatttttgcatttttgcatttttgcatt
 catttttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 gatttgcatttttgcatttttgcatttttgcatttttgcatttttgcatt
 <212> Type : DNA
 <211> Length : 12175
 SequenceName : CCR1genomicseq (SEQ ID NO: 18.)
 SequenceDescription :

Custom Codon

Sequence Name : CCR1genomicseq (SEQ ID NO: 18)

62/76

FIGURE 38 CONTINUED

63/76

```

314  GCCACCACCACCGCCAACCCGTTCTACACGCCAACGAGATCCACCAGGCCAGGCGACCGCC
-----+-----+-----+-----+-----+-----+-----+-----+-----+-----+
A      T      T      T      A      N      P      F      Y      T      P      H      E      I      H      R      Q      A      T      A      .      .
373

GCCGGGGCCAGGGTCATCGTCACCGAGGCCTGCGCCGTCGAGAAGGTGCGCGCCTTCGCC
-----+-----+-----+-----+-----+-----+-----+-----+-----+-----+
A      G      A      R      V      I      V      T      E      A      C      A      V      E      K      V      R      A      F      A      .
374

434  GCCGAGAGAG
-----+-----  443
A      E      R

```

FIGURE 38 CONTINUED

64/76

-6735	TCGACGGCCCGTAATACGACTCACTATAGGGGAAGAATTGGATCATATGGATTG	-6676
-6675	ACACTGGAATTACTCCCATGGGAGCGTGAAACAAAAAGGTGTTAGCAAGAAGACA	-6616
-6615	CTGGCAACATTGCCAGCACAGAATTGTTACAATCATAGAAAGTTTATGACAGGACATT	-6556
-6555	GTTCACCGAAAGCAAGATTACAACAATATAATCAAGGGCTGGGTCTGGTGGACATG	-6496
-6495	CTCGGTCCAATGGACGATTATTGCCGAGACCAGCTCAAGGAGTTGACGAGCACACTTA	-6436
-6435	AGCGCCGAGATCTTAAAGGCACCCAAAGTCACAAAGTCGCCATCTGCTCTTGGCAGC	-6376
-6375	TCCTGGACATCTCTCGATATTGGCTTGAAAGCCATGACCCATCATAAGCTGAAAGGCT	-6316
-6315	AGGAGGGCACCATAGGTACCGAAGTACGTTGAATACCTCGAGGACCTCCCTCGTGTG	-6256
-6255	ATGGCGAAAGCATCGATCAGCTGCCCAAGGTCTGTTGATCGATCTGGGAAGATC	-6196
-6195	ATCGAGTGCATCCCGTCATGGATCCTTACCCCTCTGAAGGAGGTCTGAAAAAGCTGG	-6136
-6135	TGAGACCCGAGGGTCATTGACAAAGCATTGCCGGAGAATTATCGGAATTATCTAGA	-6076
-6075	GCCTCAGCAGGGATGTAGGCAGCTCTGGAGAAAGTGAAGAGGGAGGAGCTCACTAACCA	-6016
-6015	AAATCAAATCGATAAAGCAAAATCGGAAAGGAGGCCAAAGGGGATTACTGAGCAAGGC	-5956
-5955	CAAGGAAGATTGGCGAAGGAGCTCATCTTTCAATGCCGAGCTCGGCAGCAAGCCT	-5896
-5895	GGATGCCTCTTCATCCTTCAGCCTCTTAGCCCTCGAGCTCATCCTAAAGGAATC	-5836
-5835	AACCTCCTGGGGCCTCGCAGCTATTTATCGCACCCCTCAGCTTCGAGGAAGAAGA	-5776
-5775	CTCGACCTCTTGCAGCGAGTCTGTCAACTCCAGAGAAGTGTATTGGGAGGCAGA	-5716
-5715	GGCCTCCAGAGAAGAGATAACAGCTACAAATCCTTAAGAGATAAGGAAAAATAATTAGA	-5656
-5655	CGAAGAACTGGTTGTCAACAAACTTATAATTGATCAGGGAAATCGTCCCACATGGATAT	-5596
-5595	ATCGTTAAAACAGGAAAGCTTACAGGTTCCCTGGAGGAGAAGCTGTAACCACGGCAGT	-5536

FIGURE 39

-5535	CAAAGAAATCTCCTTCCCTTGGAAAGGGAAAGAAGTTGTCGATATTGAGCCATGGGGC	-5476
-5475	TGCAGGAGTCGAAGCCTCGGAAGCGGCTGGATTCGGCACGATGGCACCAGATTGGC	-5416
-5415	CTTCTTGGCCGGAGGCTCGATGAAGCCATCTCACTGCAAGAACAAAAACTAGCGAAGT	-5356
-5355	CAGAATTCAATGCATATGGCGAAGTTAGAACACAATCCTGGAAAAGGAAGCAAGGACTTA	-5296
-5295	CAATTCAATAGAGACCATCTCATCGGCAAAGCCGCCGGATGATCTCTTGGAGGTAGTGC	-5236
-5235	CTCGGCCTTCCGTAGCTGCATCAACAAAGGCAGCAGCAGTCAGCATCGTCATCATGCAT	-5176
-5175	TGACCCCGCTGTATCGCTCATATCATCGGCAGAGAATCGAGGATTGATGGAAAAAGCCTC	-5116
-5115	AGGATTCACTGGATCATCATGTTGATCTATCGGCTTGCATTCCCTAGAGTATGGACCC	-5056
-5055	TACAAGGACTAAGGAATCCCTTCTTGGAAAAATTGTTGACAGGTCTGCAAACGTT	-4996
-4995	AAGAGCCGTAAGGATCTGTCGTAGTTGACGAGTGAGAATAATGGCAGTTAAATAATCAA	-4936
-4935	AGGAACATGACAATAAGAGCATAAAGGGAAATTACCTCGGTTGGCAGATGACCAGCGT	-4876
-4875	CAAATGGCGGTTGAGGAGATATCAGTGGAAATTGAATCTCCTGGCTAAAGAGGGTGAGAC	-4816
-4815	ACCGGACTTCGTCAAGCAGTTCTTCGGATAATTCAAGCAATTACTCTAGTCTCGT	-4756
-4755	CCCTGGGACCCGAATACAACCACATCGGATGGGTCTAGACATGATCGGCTGAACTCGAT	-4696
-4695	GTTTTAAGAACACAGCGGCTACCTCAGTACCTATCATGGTTGACCATCGGATTCTTGA	-4636
-4635	TCCGAAGGAATCTCAAATAACTTGTCTACTGTTGGTTTCATCGGGTGAGAGGATAT	-4576
-4575	TTTCCAAGACTCTGGCTTGCTCTAGAACATCGGAGAATTGGGGGGAGCTGGAG	-4516
-4515	TCGGCTGCTGATGAGTCCTTAATATAAAACCACTTCAGCCTCCAGCCTGGCACGGATTCT	-4456
-4455	TTCATCGGGAAAGTTGAAGTAGTTGACTTCCTTACGAGCAACAAACCAACCCACCAATG	-4396

FIGURE 39 CONTINUED

-4395	ACGAAGGACCCACCACTGCTGTTATCTTTCACGAAGAAAATCTTCTCCACAAACCA	-4336
-4335	AAGTGGGGCTCAATGCCAAAAACGCTTCGCAGAGGGTGATAAAAGATGGCAAGGTGAAGG	-4276
-4275	ATTGAGTTGGGGTTAACCTCCATAATTGAATCTCATACACTCGAAGGAGGTGGTGAAGA	-4216
-4215	AATTGTGAGCGGGAAAGCGAAAGACCTCGGTACAAGAAGGATAAGAACATCACAGTAAA	-4156
-4155	CCGGCAGGAGGATTGGGCCGTGAAATTGCACCTGGAAGAATAACATTCCCTCGTCAGAA	-4096
-4095	GAAATTATTACGAGGCTCCGGGCCCTTTCATCTCGCTTCGTGGTAGAACGCTGGC	-4036
-4035	CAATGCCAGGGATAGGCCCGCGTGGAGCTTGACGGCGCTGGCGGTGCCGGAGCTGAG	-3976
-3975	GGAGGGAGCATCTGGCGCGCTTCTCCCGGGCGGATTGAAAGGAGCCCTGACGGTGGTGCCA	-3916
-3915	CTGCTCACGGCGCTGGTGGCGAGAGTGGGATTCTTCTTCACCATTGTGAGATTGAG	-3856
-3855	GGAGATCTGGAGTTGCGACGGTGGCGTAGTTGCAAACGAAAAGGATGAATGAGGAA	-3796
-3795	GAAGGGACGCAAGGATGAAGTGTGGAAAGGGGAGTTACCCCAAGAGATTATAAGTGAA	-3736
-3735	AGGAAAACCTGAGAATTGAGCGGGCACGTGTCGTTGCTCTCAATTATTGAGGGATT	-3676
-3675	TTCTCATCATAGATCGCGGAAATCGAGGAGTCACCTGGTAACTGCACGCAAGTAGTGGT	-3616
-3615	CATTCTTAAACAGAACCGCATAGAAGTAGGATGGGACCGTCAGGTACGTCTATCAGT	-3556
-3555	CAGATTACACAGTAATTACATCATCACTGACGTCAAAGTATGCTGAAGTATCCGAAG	-3496
-3495	AAAAGTCGAAATTGGCTCGAAGACTTCTGCAGAGAAGCGCGTAGAAAGGAATATCTA	-3436
-3435	AGGAAAGGGTCAAAACATTGGCTCGAGTCTACGCACGGATTGCAAGCATTCCGTACCTAG	-3376
-3375	ACTCGGGGGCTACTCCCATCGGAGCGCTGGACGTGCACCCGATAAAATTAGACGAGGAT	-3316
-3315	GAAAACCGGAAACCAAGTGCTACTCCCATGGGAGCGCCGATTACGCACCCGACAAACT	-3256
-3255	TTTTGCACTCCAGGATCATGCCGGGACTTAATTCTGTAGAGTAGCGTTGTTTGT	-3196

FIGURE 39 CONTINUED

-3195	CTTCGGCAGTTAACCAAGCAAAGCTGGACACGTTACTCAATATCCTTACGCATTAAACCC	-3136
-3135	TTACTTGAAGAATTGAAGCCCCGATGCAAATATATCGGATGACCTATGAAGGCCTGCGGA	-3076
-3075	AAGCTTCGGGAGAAGAAGACATTGAGTGGCACAACTTGAGTCTACGAACGGATTGCAAG	-3016
-3015	CATCCGTACCTAGACTCGGGGGTACTCCCATCGGAGCGCTGGACTCGCACCCGATAGA	-2956
-2955	AGGAGATGATGATATTACAAGAAGGACAAGAAGTATCAAGGGAGAAGAACATTGGTGGA	-2896
-2895	GGCATGCTTTAGTCTCACCGAAAAAACTCGGCTAGACACTCGGGGGCTACTGACGT	-2836
-2835	GGGCATTACCCCTCGGGTAACTGATATTGCCCTATCCTGTACGACCCAAGTGGAGGCCA	-2776
-2775	TGAAGACACTCGAAGGCAAGGTGGACACTACGTCGGTGCAGGGGGTTCCCTTGAAGA	-2716
-2715	ACAAGACGAAGAAAAGAAGAATACAAGAAAAGTATAGAAACTAGGATCTTGTAACTGG	-2656
-2655	TCGTACCCGGACAGATCTCTCGAGACCTGGCCCCCTACATATGGCTAGGAGAGGGCTG	-2596
-2595	CCGAGAGGGACACACACAATCTTAGCAATTAGCCACCATAGTCCAGAGCAAGGTCCC	-2536
-2535	CGTAGAACTTAGCCTCTCGACGAGATCACAGCCGAAACCTTCGGCACCCATTGTAACCC	-2476
-2475	GATATTTCATAGTCAGAGTCAGACAGGTAGGACGTAAGGGTTTACCTCATCGAGGGCC	-2416
-2415	CCGAACCTGGTAAATCGCTCTCCCCGCTTGTGATAACCGATGGCTTGTCTAGCTTA	-2356
-2355	CATGATTCCATCTACCTAAACCTAAACGGAGGGCATTGCCGAGGAGTACCCCTGACAT	-2296
-2295	TCCCCTCCACCAATGGTCTCACATAAATTCAACAAAGCAAACCTACATAAAAGTTAACG	-2236
-2235	GTTTCAGAAAGAAATAAAACTAGGCCCTCCTTGAGAATCTACGAATGATTACCATAT	-2176
-2175	CATCTCGCAGTTAGTGTAGTAACGAACTAAGTCTCAAATTCCGACGCATGGCGAAAAGG	-2116
-2115	TAGCGAACTTAAATGTGAGGAATGAATGCCACATATGCATGGTGCATCGAGTATTCTCA	-2056
-2055	TTTTAGTCTGGATTACTCCCTTAGATGTTGACACCACCCAAAAATACAACATTGGACA	-1996

FIGURE 39 CONTINUED

-1995	AGTTGTTCACTTCACTAGTATGAATTCAGTAAATCGGGCAATACTCCAACACTCATTCA	-1936
-1935	CCCCCTAGGCAGGGTTAGCTCAGATCAACGTCGGGTCTTCATCGAGTTAATGTCGTCA	-1876
-1875	CACGCACACACACGTACGCGCACACACACGTGCGCAAACAAAAAGAAAAGACCTT	-1816
-1815	CTCACGTAGCCTAGGTCTTGCCTGTAAGAAAAACCCAGGTCCACCCTAGTTCGAACCC	-1756
-1755	AAAATATTTGAAGATACTTAGTAAGATATTTGAAAATAAACCGCAAAAGGGAA	-1696
-1695	TTGAAAATATGGACTGGCTGTTTGTCCAAAACCACATCTTCGGAGAACCGACGAGGGT	-1636
-1635	ATCTATTGATGGGCTCATACTATACCTGGCATGTGTTGGCAGGCCTCATGTCGGCC	-1576
-1575	GAGGAAAGCCCGACGCTGAAAATCAGGCCAAGCTTAACCCGGCCGACCAAATACCCA	-1516
-1515	CCAAACCGTTGGGCCATCAGGTTGCAGGGCAGTAGTGTAAAACACCGATTCGGG	-1456
-1455	CTACATAGGCCGGCTCGTTGTCGGCAAACATTCTAGACCTAACGGCGAGTTTCG	-1396
-1395	GGCCGGGCTGCCATGGCAGGTATAGCTCATACGACGTATGACATTGAGCAATTGA	-1336
-1335	TGCAAAGCACGTGTAGGTTTATCCCATCCGTGTGGCGTGTAGGGTGTAAATGAATA	-1276
-1275	GGATAATTCCTCGCCGAAACTGGTCCAAATTGCTTGAAGTGTCCATATATGATTT	-1216
-1215	AAAGAATGTGACAAATAAAGATATCCAATTGAAATAGTGTCCGGATACGGTATAGGA	-1156
-1155	TATGGTATAGCAAATAACATGCTGATATGGATTGTCGATATTAAAGATAATCCAA	-1096
-1095	ATGTTTAAACCGCATAATTGATTTGAGTCAAAAGCGAATGCCAATTGAGGTTA	-1036
-1035	GCAGTTATTGAGTTCAAAATTATTGGCGAGCATATCTAGTTCTAAATTCTATCACGT	-976
-975	AAATTGTGTCTTTTTAATAACTACACAAGACTAAAAGTTAAATCTCTCAAGATT	-916
-915	GCGAAAATATAGCTATCTACTGATATATATCCGACTATTTGTTTCGGACCGCAT	-856
-855	GCGTCCTATTCCGATTGAACTGCACCTCCGATATCCACATTGAATCTAAACCGAT	-796

FIGURE 39 CONTINUED

69/76

-795	CAATATTGCTCCGATCTAAATCCGGAAAAATATGTGGTGAAGGGATATGGTATAAGCAAA	-736
-735	ATCCGATTTGATCCATTTGACCTCTAGGCCTGTGCAAGACCTGGAGGAAAGAATGGCGC	-676
-675	ATCTGTAGGGTGCAGTCCCACCGTGGAAAATGTGAGCTCACCGTATTGTCGGGATGG	-616
-615	AGCATCGAAACGGAGTCGGAACACGATTGCGCCACGTACAGAGCATGCATGATTCCCT	-556
-555	TGTATGCGGTCCAGGATCTTAAACTGCCTTCCATTTCAGGAACCTACCGATTGGCTGCA	-496
-495	AGCCGTAGCTAGCGGTTGAAGTCACGGCATTGCCGCCCCGATTAACCCACCCGTCGCG	-436
-435	CGCGCGGTGGTCGTTCACCGTCCTGCCCTAGGCTACGCACGCGCGCGCAGTTGGCC	-376
-375	AGTTGTAGGTAAGCCGACTCGAGATCACACACCCGGCCTCACCTACTACCTCTGCCGTC	-316
-315	GCGGTCAACCGTGTCAACTCACGCCAGGGAGCCACCCGCCACACGGCGCCTAGCTCA	-256
-255	TCCCCTCTCACTACTCTTCTCCTCCCTCTCACCTCGCCGTCGACCCAGCTCCGGCT	-196
-195	CTATAAATTCCGCACTACTCGAACCAACATGCCAGGCCTTGCCTTTACGACGAATC	-136
-135	CTACCAAACCGAGCTACCAAGATCCTCTACTAATCGAGCTCCCTACGCTGCTCCGCCT	-76
-75	GTCTCGTTCCGCCCTCACGCCGGCGTTCTCCGCTCCAAGCTACGTCCGTCGCCA	-16
-15	CATATATAGCATCGACATGACCATGCCGAGGTCTGGCTGCCGGAGACACCGCCGCCG	44
	M T I A E V V A A G D T A A A	
45	GGTGGTGCAGCCGCCGGAACGGCAGACCGTGTGCGTGACCGCGCCGGGTACAT	104
	V V Q P A G N G Q T V C V T G A A G Y I	
105	CGCGTCGTGGCTCGTCAAGCTGCTGGAGAAGGGTACACCGTCAAGGGCACCGTCAG	164
	A S W L V K L L E K G Y T V K G T V R	
165	GAACCCAGGCATGTCACCCATGCATTCACTATTTCTTAAGTCGTATGCGTTATGCGA	224
	N P G	
225	CTTGTGTATTAACATTGTGGACTGCATGCAGACGACCCGAAGAACGCGCACCTGAGGGC	284
	D P K N A H L R A	

FIGURE 39 CONTINUED

FIGURE 39 CONTINUED

71/76

FIGURE 39 CONTINUED

72/76

2445	GTGGACCTTGTGGTGGTGAACCCGGTGCCTGGTATCGGCCCCCTGCTGCAGCCGACGGTG V D L V V V N P V L V I G P L L Q P T V	2504
2505	AACGCCAGCATCGGCCACATCCTCAAGTACCTGGACGGGTGGCCAGCAAGTTCGCCAAC N A S I G H I L K Y L D G S A S K F A N	2564
2565	GCCGTGCAGGCCGTACGTGGACGTCCGGCACGTGGCCAGCAGCCACCTCCCGTCTCGAG A V Q A Y V D V R D V A D A H L R V F E	2624
2625	TGCGCCGCCCGTCCGGCCACCTCTGCAGCGAGCGCGTCCACCAGCGAGGACGTC C A A A S G R H L C A E R V L H R E D V	2684
2685	GTGCGCATCCTCGCCAAGCTCTTCCCCGAGTACCCCGTCCCCACCAGGTACGCGTACGAC V R I L A K L F P E Y P V P T R	2744
2745	CTGCTTGCTAGCCGCTTCCGTTAATTCCATTGCCTTAATTGATTGCATGATGCCGCTCCT A A T T T A C T C A C T T G C G T A A C T A A T T G C A T T C A T A T G A T C T A C C A A C C G T G G A G A A A T	2804
2805	TAGCAAGAGTCTGTCGGGCGTCCCGGTCCAGTGCAGTTAACCTGCATGTCGATGGCTG C A G G T T G C A G C T T A C T T G T G G T T C T T A G T T C A G A G A C A C A G A G C A A T T G G G C A C T A A G C	2864
2865	AAA A C T G A C A T C A C T G G T A A T T A G G T A G C T C C A C A C A C T G A A G T G G G T G G A T C C C A T C G G T A G T A G G T A A G G G T G G A T A G T A C T G G A C G A G A G C T C G A T C G T T G T A A A A A A G C G A G	2924
2925	TGACCAACCACCTCACCACACTGCAAGTAGCTGCTAGTGAACCATCCAACCAGCTCCCT G G A T C A C T C T G C T C C G T C C G T A C C T C A G C T A C A G A A G C G A C A T G A A C A C A C A G A C	2984
2985	A A A A C T G A C A T C A C T G G T A A T T A G G T A G C T C C A C A C A C T G A A G T G G G T G G A T C C C A T C G G T A G T A G G T A A G G G T G G A T A G T A C T G G A C G A G A G C T C G A T C G T T G T A A A A A A G C G A G	3044
3045	TGACCAACCACCTCACCACACTGCAAGTAGCTGCTAGTGAACCATCCAACCAGCTCCCT G G A T C A C T C T G C T C C G T C C G T A C C T C A G C T A C A G A A G C G A C A T G A A C A C A C A G A C	3104
3105	GGATCACTCTGCTCCGTCCGTACCTTCAGCTACCTACAGAAGCGACATGAACACACAGAC A C A C A A G G C C G G C T C A C C A T T C G C A T A G G T C A A A C C A A T G T G G T G A A C G G C A A C A T C G	3164
3165	CCACAAAGTCGCGTGCTAGTCGAGGTTGTCCGGTGTACCGAGGCCACACTATTGTCG A C A C A A G G C C G G C T C A C C A T T C G C A T A G G T C A G C G A A A G T A G G T G G A C T G A C A G A T A C	3224
3225	TGCCCGTCGCTGATATTGCACGCGTAGCTGTCGACGAAAGTAGGTGGACTGACAGATAC A C A T A T C C T C A T T G C C T T C T C T G C T C G G T T T C T G C T A G G A T T G C C A T C T T C A G G A G T G C C	3284
3285	TATCCGCACGGCAGAAACCGTAGCATCAGGCCAGAAAGCAGCGTGCCTGATATCGTAAC T A T C C G C A C G G C A G A A A C C G C T A G C A T C A G G C C A G A A A G C A G C G T G C G T G A T A T C G T A A C	3344
3345	ACATATCCTCATTGCCTTCTCTGCTCGGTTCTGCTAGGATTGCCATCTTCAGGAGTGCC T A T C C G C A C G G C A G A A A C C G C T A G C A T C A G G C C A G A A A G C A G C G T G C G T G A T A T C G T A A C	3404
3405	TATCCGCACGGCAGAAACCGTAGCATCAGGCCAGAAAGCAGCGTGCCTGATATCGTAAC T A T C C G C A C G G C A G A A A C C G C T A G C A T C A G G C C A G A A A G C A G C G T G C G T G A T A T C G T A A C	3464
3465	TATCCGCACGGCAGAAACCGTAGCATCAGGCCAGAAAGCAGCGTGCCTGATATCGTAAC T A T C C G C A C G G C A G A A A C C G C T A G C A T C A G G C C A G A A A G C A G C G T G C G T G A T A T C G T A A C	3524

FIGURE 39 CONTINUED

73/76

3525	CCAGACGGTCTTCACCTGTCCATTCTGGGCTACCTGGCATACTACCTCGGTGCCGCTGTG	3584
3585	CCGCTGACCAATTCTGCACGACCCTATAGCAAAACCCATGCATGTAAGCTTCAAG	3644
3645	ATCAGCAGTGACATGTGCAATATAAACCTCAAGTGTGCACTCTAGTGCCTACTGATAAAA	3704
3705	CCGTATAACTGGTGACCCAGTCATTCTCTTTTATTTGTTGGACCAACGAACAC	3764
3765	AGCATGTTATCCATACCAACAAGTGGCGCTGATTTTCAAACACTACACTGGGATCATACT	3824
3825	GGAAACCAAAGCAGGAGAACATCTCGAACCAAGAGATGTTACTAAATTGAAAGAAAA	3884
3885	TGTACTGACAAGTAATCTGTCGAAGCAAGACACATACTACCTCGGTTCGAACGTGGGAC	3944
3945	ACCATGCCCGTGCATATTGCTAGGCACCCTGCGCTGATTGTATCCAACCGGAGG	4004
4005	GAGTATCGATTGCGCAAAGTCCCTACATACATAGCCGCTCAAGATATAATCTTACGACC	4064
4065	TTCCGTCGAAATCGGTATACGTCGCAACCTATAGCTAACCTGGCAGAGCATAAAATAAC	4124
4125	TATCTAAGGTTGGGTCTCCCTCTTCAATCAACCTTCACTACCGAATGATGGAGTGT	4184
4185	TTGTGAAAACATCTTGGTCGACTCAGCATTAGCGCCCTACCAATTCTCTGTGGACAA	4244
4245	TGCCACCTTAAATCGTTTTAGTCTTCATGATTACTCCCCCTATATCTGGCCGTAGT	4304
4305	CCCTCTTCCATTTCCTGCTGGTTTAAGTCAAATTAGACTACTAAAACAACAGC	4364
4365	AAGATTTATGGAAGGGAGGTAGTGCAAAACAGAAAGTCCGATCGAAATGCGTGCCAATT	4424
4425	TGTCGTCGGCGGCCGGACTAAAATGGATCTGCATGTCATACCGTTCGTGGAGTATC	4484
4485	CTGCGAACGGTCGTGTTAGTCACATTAATGTGAGGTTCATGTGATACTCTGCTTG	4544
4545	AAAGATACTACTGCTACCTCGTAGAACCTGAATGAAAGTATGTGGACTGTTCAGCTC	4604
4605	TCTGCACATGTCAAATGTCGTTACTCATACCTTCGTCAGAGCATCCTGCGACGCGCGCC	4664
4665	GGTGCGAAATTGCGCGTGTGTTAGTCAGAGATCAACCGTGAGGTTATGCGGTACCCCTA	4724

FIGURE 39 CONTINUED

4725	TCTGGCTTCGAAGATAACCAAGCAGACTGCGGCTAGATTGTCATTTGATGTCGCAATCIT	4784
4785	CACCAAAACCTGCCCTTCCGGACCACAGCAGCAGTACGTAACAATGGTGTATGCCATGC	4844
4845	GTTGCTCGTGTCCAAGGAAACGGAGGAATCTCGGCTTCCCACAAGTCACGCATCGATGTT	4904
4905	CACACCTGAATTGGTCGACGTTCTCTTAGACTAGAAAAAGATTACAGAACACGCA	4964
4965	AGCTTCGTTCAAGTCATACTTCTGTTCACTGATGATTGCAGTTATATCAGC	5024
5025	ATGTCCTATTCTGAATTTCGACTTCTATTCAAAGGATGGGCTGGAATTGCTACTGACTT	5084
5085	TGGTGTGATGTGTGGCACAGGTGCTCTGATGAGACGAACCCGAGGAAGCAGCCATACA	5144
	C S D E T N P R K Q P Y K	
5145	AGATGTCGAACCAAGCTCCAGGACCTCGGACTCGAGTTCAAGGCCGGTGAGCCAGTCCC	5204
	M S N Q K L Q D L G L E F R P V S Q S L	
5205	TGTACGAGACGGTGAAGAGCCTCCAGGAGAAGGGCCACCTTCGGGTGCTCAGCGAGCAGG	5264
	Y E T V K S L Q E K G H L P V L S E Q A	
5265	CAGAGGCGGACAAGGAAACCCCTAGCTGCCGAGCTGCAGGCAGGGTTACCATCCGAGCAT	5324
	E A D K E T L A A E L Q A G V T I R A *	
5325	GAGGAACAAAGAAATCAACCATGTCCATACTGCTACTGTCATGTAACCAAGCTGTTGAATG	5384
5385	CCTAAAATCTAAGTTCTTGTAAATACTGTGTTCATGTGGACTAGATTGATCG	5439

FIGURE 39 CONTINUED

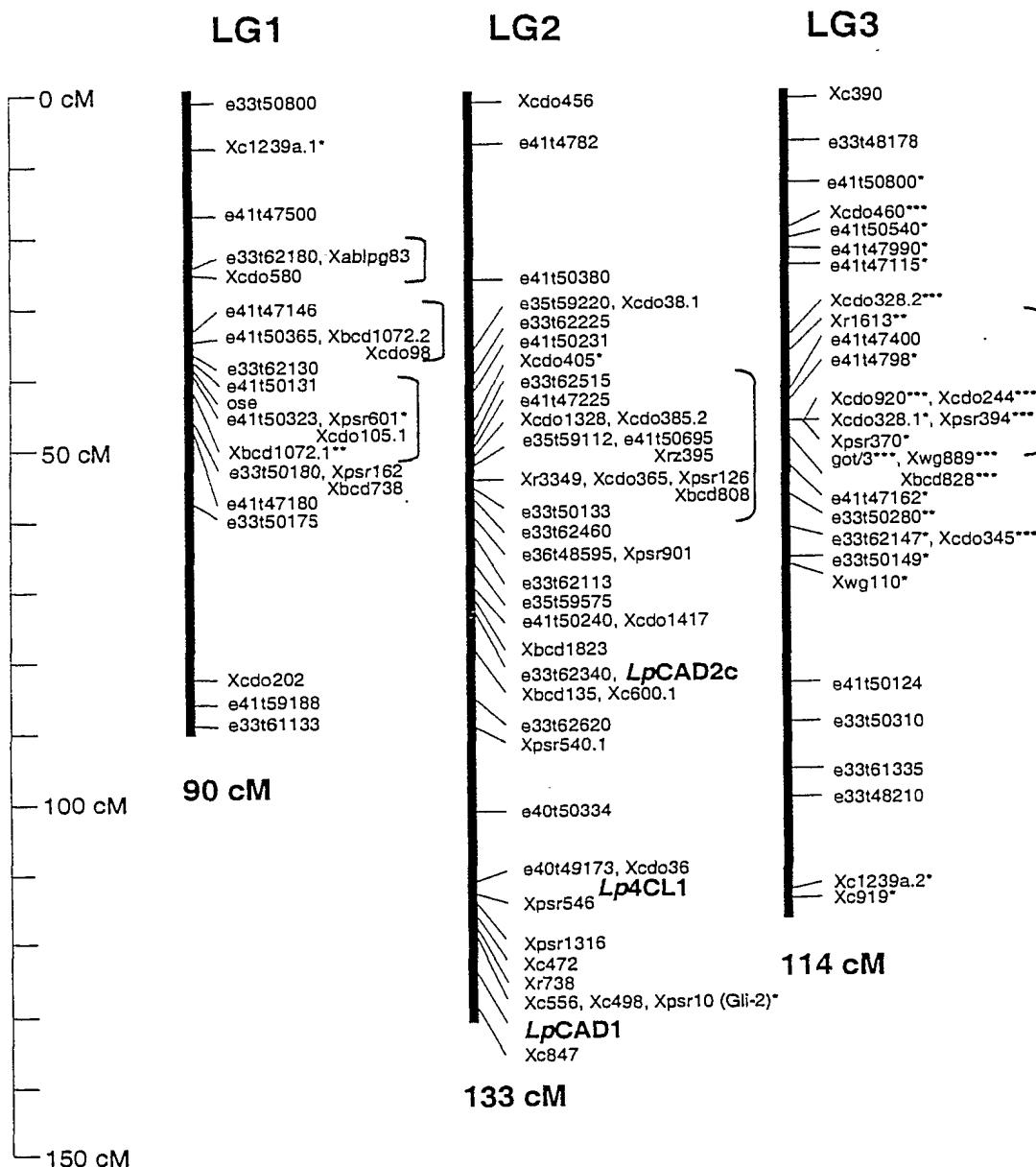


FIGURE 40

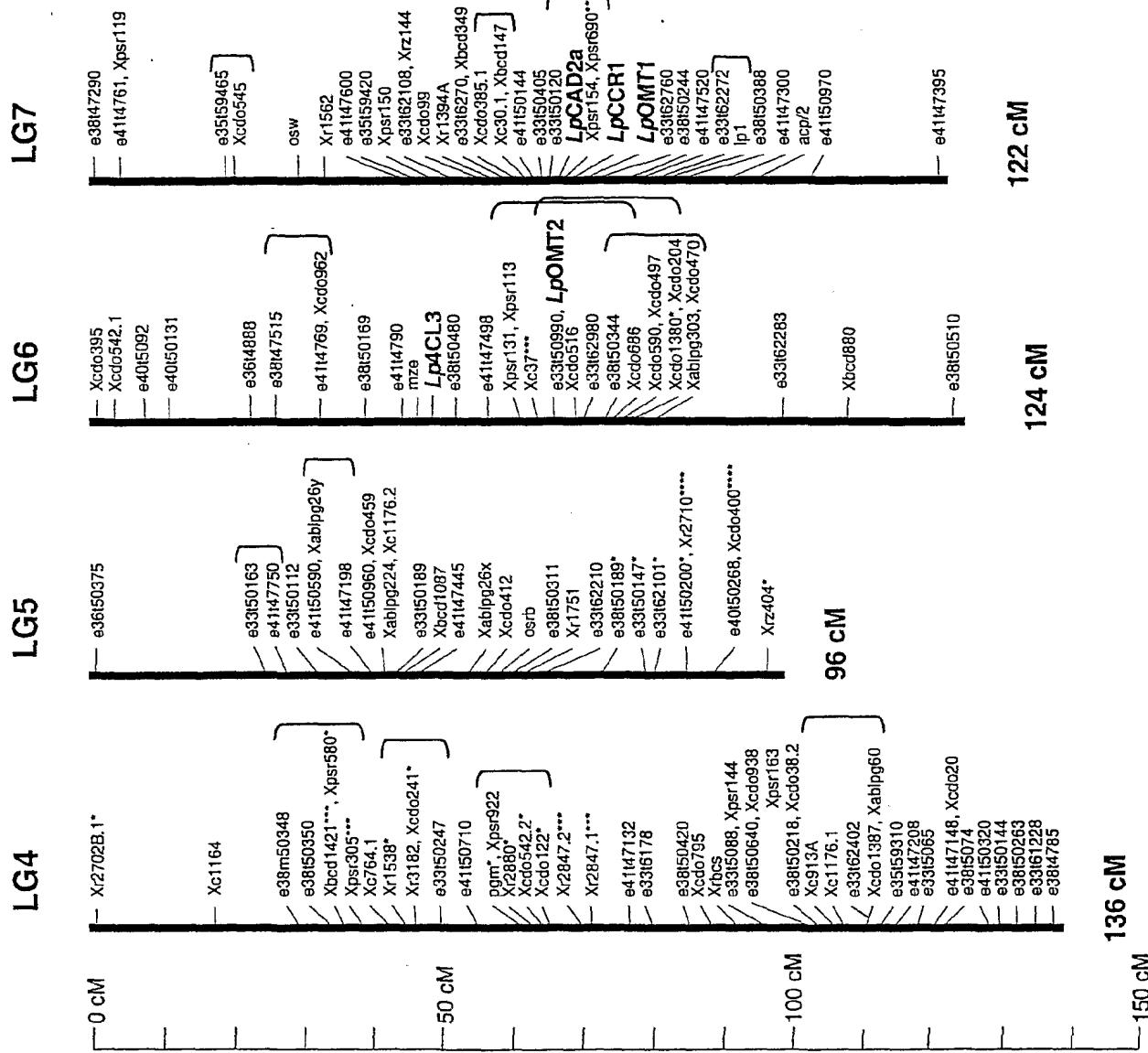


FIGURE 40 CONTINUED

Organization Applicant

Street : 15th Floor, 8 Nicholson Street
City : East Melbourne
State : Victoria
Country : Australia
PostalCode : 3002
PhoneNumber :
FaxNumber :
EmailAddress :
<110> OrganizationName : State of Victoria as represented by Department of Natural Resources and Environment

Organization Applicant

Street : North Terrace
City : Adelaide
State : South Australia
Country : Australia
PostalCode : 5005
PhoneNumber :
FaxNumber :
EmailAddress :
<110> OrganizationName : The University of Adelaide

Organization Applicant

Street : Lisboa 27, Apartado Postal 6-641
City : Mexcio
State : DF
Country : Mexico
PostalCode : 06600
PhoneNumber :
FaxNumber :
EmailAddress :
<110> OrganizationName : International Maize and Wheat Improvement Center

Organization Applicant

Street : Waite Road
City : Glen Osmond
State : South Australia
Country : Australia
PostalCode : 5064
PhoneNumber :
FaxNumber :
EmailAddress :
<110> OrganizationName : State of South Australia as represented by South Australian Research and Development Institute

Organization Applicant

Street : Military Road
City : Lismore
State : New South Wales
Country : Australia
PostalCode : 2580
PhoneNumber :
FaxNumber :
EmailAddress :
<110> OrganizationName : Southern Cross University

Organization Applicant

Street : Level 3, 84 William Street
City : Melbourne
State : Victoria
Country : Australia

PostalCode : 3000
 PhoneNumber :
 FaxNumber :
 EmailAddress :
 <110> OrganizationName : Dairy Research and Development Corporation

Application Project

 <120> Title : Modification of Lignin Biosynthesis
 <130> AppFileReference : 40494788
 <140> CurrentAppNumber : AU PQ8154
 <141> CurrentFilingDate : 2001-06-14

Earlier Applications

 <150> PriorAppNumber : AU PQ8154
 <151> PriorFilingDate : 2000-06-14

Sequence

 <213> OrganismName : Lolium perenne
 <400> PreSequenceString :
 cggcacgagt ggactttccg acgccccggact cggcgatgtat gaccggcatttgg aggaggtagt 60cgttagtcgt
 c ctccgcctcg tacgcgcgc tgcccgccat ttcccttcctc gcctcgccgg 120tccttcctccc cgacctg
 cgc taggctctgg atctcgccgg gtttgggcgc ggcgtccctcg 180ctgtgagctc gtgccgaatt cggca
 cgagc cacccgcgcg gcgtgcactg gtacgagctc 240gcgagccatt gtcagtgcag tgtaggcctc gct
 actcggtt ggcattccaa agaaacctc 300tgctccctga aaccaggagga tcatgtatcac ggtggccggc
 ccggagggtgc acgagccgc 360gatcgcggcg gctgctgcgg ccgtggagggc ggccggcaccgg gaggcgacgc
 cgatcttcgc 420gtccaggcgc cccgacatcg acatccgcac ccacatgcgc ctgcacgact attgcttc
 gc 480gacggcagcc tcggcccccgg acgcgcgcgtg cctcatcacc gcggccacgg ggaagaccta 54
 0cacgttcgcg gagacgcacc tgctgtgcgc caagggccgcg gcggccgtgc acgggctcgg 600cgtgcgc
 ac ggggaccggc tcatgctgtc gctccagaac tccctggagt tgcgcgtcgc 660cttcttcggc gcgtcc
 atgc tcggccgcgt cagcacggcg gcgaaccgt tctgcacgc 720gcagggagatc cacaagcagc tgc
 ggcctc cggcgcgaag ctggcgtca cgcgcgtccgc 780ctacgtcgac aagctccggc acgaggcctt cc
 cccgaatc ggcgaggccc tcaccgtat 840ca
 ccatcgac gaggacgcgc gcacccggaa cggctgcgcag ccgttctggg ccctcggtgc 900agccggcgc
 gagaacagcg tcccgagtc tcccatctcg cccgacgcgc cgggtggcgct 960gcctactcg tcggccacg
 a cggggctgcc caagggcggtg gtgcgtacgc acggggggct 1020gggtgtcgagc gtggccgcgc aggtgg
 cgg cgagaaccccg aacctgcaca tgcggccgg 1080ggaggacgtg gtgcgtcgctg tgcgtccgccttcc
 acatc ttctcgctca actcggtgtc 1140gctgtgcgc ctgcggccgg gcgcgcgcgt gatgctgtatgc cct
 aggttcg agatgggggc 1200catgctggag ggcacatcgaggc ggtggccgcgt cacgggtggcg gccgtgtgc
 gccgctgtt 1260gctcgccgc gccaagaacc ccgggggttga gaagcgcac ctcagctcca ttccgatcg
 1320gcttcggc gccgcgcgc tcggcaagga gctcgaggac gcgcgtacgt gccgcgtgc 1380g
 caggccatc ttccggacagg gctacgggat gacggaggcc gggccggcgtgc tgcgtatgt 1440cccccgcgttc
 gcgcggggagc cgacgcgcgc caagtccggc tcgtgcggc ccgtgggtgc 1500caacgcgcgc ctcaagg
 gg tcgcaccccgca caccggcgtc tccctcgcc gcaacctccc 1560cgccgagatc tgcacccgc gcccgc
 agat catgaaaggaa tacttgaatg atccctggc 1620cacggccgcg accatcgacg tgcgggggtg gctc
 cacacc ggcgcacatcg gctacgtcg 1680cgac
 gacgac gaggtttca tcgtcgaccgc cgtcaaggag ctcatcaagt tcaagggtt 1740ccaggataccgc
 cc gggcgcgc tcgaggctct gtcacatcgacgc catccgttca tcgcgcacgc 1800ggccgtcgac ccgcacaaagg
 atgatgcgcgc cggcgaggc cccgttgcct tcgtgggtcc 1860cgccgcgcac tccgcacatcg cggaggagg
 c catcaaggag ttctgtatcca agcagggtgt 1920gttctacaag aggctgcaca aggtctactt caccac
 ggcg atacccaaatg cggcggtcgg 1980gaagataactc agggaaagaaac tcagagctaa actcgccgc cccgc
 cactg cctgtggat 2040gggttcatggc ttcatgtctaa tcatttcgtatc cggaaaggca cttcttagcat at
 ttttcca 2100ccctttgtttt catttggaaatg attgttattcc agctgtggc cagtgtactga gtaaggatg
 2160gggataaaaatg ttttgttcatc gttttctttt acgttactct ctcattggg ggttacaatg 2220tat
 cagggga ttcgttattt aagttatca agattgggttca aattataaaa aaaaaaaaaa 2280aaaa
 2284

<212> Type : DNA
 <211> Length : 2284
 SequenceName : 4CL1cDNA (SEQ ID NO: 1)
 SequenceDescription :

Custom Codon

 Sequence Name : 4CL1cDNA (SEQ ID NO: 1)

Sequence

<213> OrganismName : *Lolium perenne*
<400> PreSequenceString :
MITVAAPEVQ QPOIAAAAAA VEEAAAPEATT IFRSRLPDID IPTHMPLHDY CFATAASAPD 60APCLITAAT
G KTYTFAETHL LCRKAAAALH GLGVRHGDRI MLLLQNSVEF ALAFFGASML 120GAVSTAANPF CTPQEIH
KQL VASGAKLVVT QSAYVDKLRH EAFPRIGEAL TVITIDEDDG 180TPDGCQPFWA LVSAADENSV PESPI
SPDDA VALPYSSGTT GLPKGVVLTH GGLVSSVAQQ 240VDGENPNLHM RAGEDVVLCV LPLFHIFSLN SVL
LCAALRAG AAVMLMPRFE MGAMLEGIER 300WRVTVAAVVP PLVLALAKNP GVEKHDLSII RIVLSGAAPL G
KELEDALRG RLPQAIFGQG 360YGMTEAGPVL SMCPAFAREP TPAKSGSCGT VVRNAQLKVV DPDTGVSLGR
NLPGEICIRG 420PQIMKGYLND PVATAATIDV EGWLHTGDIG YVDDDDDEVFI VDRVKELIKF KGFQVPPA
EL 480EALLIAHPSI ADAAVVPQKD DAAGEVPVAF VVRAADSDIA EEAIKEFVSK QVVFYKRLHK 54
0VYFTHAIPKS ASGKILRKEL RAKLAAPATA RVVHGFMLII SIRKALLAYM FHLLFHLED C 600IPASGQ
606

<212> Type : PRT
<211> Length : 606
SequenceName : 4CL1pep (SEQ ID NO: 2)
SequenceDescription :

Sequence

<213> OrganismName : *Lolium perenne*
<400> PreSequenceString :
cgccacgagc gccatttcctc caccttcagc tccggccaaa gatttccatc cggcgagatc 60catgggctc
c atcgcggccg acgcgcctcc cgcggagctg gtgttccggc ccaagctccc 120ggacatcgag atcccgaa
ccc acctgacgct gcaggactac tgcttccagc gcctgcccga 180gctctcccg cgccgcctgccc tcata
gacgg cgcacgggc gccgcgcctca cctacggcga 240gggtggacgccc ctgtccccc gctgcgcggc ggg
gctgcgc cgcctcggc tcggcaaggg 300cgacgtcgatc atggcgtccc tccgcaactg ccccgagttc g
ccttcgtgt tcctcggcgc 360ggccggcgc ggcgcgcgc ccaccaccgc caaccgcgtt tacacgcggcc
acgagatcca 420ccgcggccg accgcgcgc gggccagggt catcgatcacc gaggcgtcg cggtcgag
aa 480gggtgcgcgc ttcgcgcgc agagaggat tcccgatcg tccgtcgacg agggcgtcg 54
0cgccggcgtgc ctcccgatcg ccgagactct gctcggggaa gaaagcggg agcgggtcg 600cgacgagg
cg gtcgaccccg acgacgtggt ggcgcgtccg tactcgatcg gcaccaccgg 660cctgcggcaag ggcgtc
atgc tcaccaccgc cgcctcgatc accagcgatc cccagcagg 720ggacgggtgag aaccgcgacc tgca
cttcag ctcgtcgac gtgtcgatcg 780gctgttccac atctactcgatc tcaactcggt gc
tgctcgcc ggtctccgcgc cccgggtcg 840ga
tcgtgtatc atgcgtcaatc tcgaccacgg cgcgcgtgg gacgtggatc gcacgcacgg 900cggtcaccgtg
gcgcattcg tgccgcgc tgcgtggatc atgcgtcaatc gcgcgcggg 960gaccgcgcgc gacctggcg
t ccatccggat ggtcatgtcg gggggccgc ccatggggca 1020ggagctgcgc gacgcgttca tggccaa
gat ccccaacgcgc gtgcgtccgc agggatatgg 1080gatgaccgc gccggccctg tgctggcgat gtgc
tggcc ttgcgtccgc agccgttcgc 1140gggtcaatcg tgcgtccgc gcaccgtcg caggaacgc ggg
ctcaaga tcgtcgaccc 1200cgacaccgc gcctccctcg gccgcacact gccggggggag atctgcaccc g
cgcaagca 1260gatcatgaaa gtttacctaa atgtccgtt ggcacaaag aacaccattt acaaggacgg
1320ttggctgcgtactggatc ttggatcgatcgatc gacgatgttgc 1380c
agactgaag gagataatc aatataaggg attcaatcgatc cctccgcgcg aacttgc 1440ccttctcatt
acacaccctcg aaatcaatcgatc tgctgtcgatc gtatcgatgc aagacgaaact 1500tgctggatcgatc
tg cgttggatcgatc ggcgtcgatcgatc gtttacttgcgtatcgatc 1560cgagatcgatc cagttcgatcgatc
aggt tggatcgatc aagaggatcgatc gcaaatcgatc 1620cttcgcggat tccattccaa agatccatc tggc
aagatc ctcaggacgg acctcgatcgatc 1680aaag
ctcgcc gcaggcattc ccaggatcgatc taccacacag tccaaaatcgatc aagtcgatcgatc 1740tattgttcc
acccatcgatc cacctctcgatc caacaccatcgatc ttttacttgcgtatcgatc 1800gaaattatcgatc
ggctgtatcgatc ttttacttgcgtatcgatc 1860cttgcgtatcgatc cgtatcgatcgatc
a ttttacttgcgtatcgatc 1920aggctgatcgatc ttttacttgcgtatcgatc 1980aaatcgatc
ttc agggatcgatc aaaaaaaaaaa aa
1992

<212> Type : DNA
<211> Length : 1992
SequenceName : 4CL2cDNA (SEQ ID NO: 3)
SequenceDescription :

Custom Codon

Sequence Name : 4CL2cDNA (SEQ ID NO: 3)

Sequence

<213> OrganismName : *Lolium perenne*
<400> PreSequenceString :
MGSIAADAPP AELVFRSKLP DIEIPTHLTL QDYCFQRLPE LSARACLIDG ATGAALTYGE 60VDALSRCA

A GLRRLGVGKG DVVMAILRNC PEFAFVFLGA ARLGAATTAA NPFYTPHEIH 120RQATAAGARV IVTEACA
 VEK VRAFAAERGI PVVSVDEGVD GGCLPFAETL LGEESGERFV 180DEAVDPDDVV ALPYSSGTTG LPKGV
 MLTHR SLVTSVAQQV DGENPNLHFS SSDVLLCVP 240LFHIYSLNSV LLAGLRAGCA IVIMRKFDHG ALV
 DLVRTHG VTVAPFVPPV VVEIAKSARV 300TAADLASIRL VMSGAAPMGK ELQDAFMAKI PNAVLGQGQG M
 TEAGPVILAM CLAFAKEPFA 360VKSGSCGTVV RNAELKIVDP DTGASLGRNL PGEICIRGKQ IMKGYLNDPV
 ATKNTIDKDG 420WLHTGDIGYV DDDDEIFIVD RLKEIIKYKG FQVPPAEELEA LLITHPEIKD AAVVSMQD
 EL 480AGEVPVAFVV RTEGSEISEN EIKQFVAKER VFYKRICKVF FADSIPIKSPS GKILRKDLRA 54
 OKLAAGIPSSN TTQSKS 556
 <212> Type : PRT
 <211> Length : 556
 SequenceName : 4CLpep (SEQ ID NO: 4)
 SequenceDescription :

Sequence

<213> OrganismName : *Lolium perenne*

<400> PreSequenceString :

```

cggcacgaga tctcccccaga ctaattttaga agaagatcta cttagtcgtat gcttctcgct 60cgatcgccg
g cccggtaggt agcttagctag ctactcgtag tagaccatata ccatgggttc 120cgtgccggag gagtcag
tgg tggcggtggc accggccggag acgggtgtcc ggtcgaagct 180ccccgacatc gagatcaaca acgag
cagac gctgcagagc tactgtttcg agaagatggc 240cgaggtcgctg tccccccct gcacatcatcgac cgg
ccagacg ggcgcctctt acacactac 300ggaggtcgac tcctgtaccgc gtcgcgcgc ggcggggctc c
ggccatgg gctgtggggaa 360ggggcgacgtg gtgtatgacc cgtcgccaa ctggccggag ttgcgccttct
ccttcctggg 420cgccgcgcgctg cttggcgccg ccaccacac cggccaaaccc ttctacaccc cgcacag
at 480ccacccggccag gcccggaggccg cggggccaa gctgtatcgat accggggctt ggcggctgg 54
0gaagggtctg gatgtcgccg cggggccgggg cgtggccgtg gtcacccgtcg acggggaggcg 600cgacgggt
gc gtggacttcg cggagctgtat cgcggccgag gagctggccg aggccggacga 660ggccggggtc ctcccc
gacg acgtcgctgc cctgccttac ttctccggca ccacccggct 720ccccaaaggcc gtcacatgtca cccaa
ccgcag cctcgtaacc agcgtcgccc agctggtcg 780cggggtcgac cctaaacgtgt gttcaaccaa gg
acgacgcg ctgtgtgtcc tgctggcgct 840ct
tccacat tactcgctgc acacgggtgt gctggcgccgg ctccgcgtcg ggcgcgcct 900cgtcatcatg
cgcaaggttcg acgtcgccgcg gctgggtggc ctcgtcccgccg cgcacccat 960caccatcgcc ccattctgtg
c cgcgcattatcgat cgtggagatc gccaaggagcg accgcgtcg 1020cgccgcacgc ctcgcatcca tcccgat
ggc gctctccggc gcccggccca tgggcaaggaa 1080cctccaggac gcttcatgg ccaagatccc caaccc
ccgtg ctccggacagg ggtacgggat 1140gaccgggct gggccgggtgc tggccatgtg cttggcggtc gccc
aaggagc cggtcaaggt 1200caagtccggg tcgtgcggaa ccgtgggtgcg caacggccgag ctcacgggtcg t
cgaccccgaa 1260caccggcgca tccctcgcc ggaaccagcc tggcgagatt tggccatggg ggaaggcagat
1320catgataggt tacctgaacg acccagagtc gaccaagaac accatcgaca aggacggct 1380g
ctgcacacc ggagacatcg gcttgggtga tgacgacgac gagatctca tcgtcgacag 1440gctcaaggag
atcatcaagt acacggggctt ccaagtggcg cggccggagc tcgaggccct 1500cctccctcaacg aaccggaa
gg tcaaggacgc cgcgcgtgtat ggggtgaagg atgatctctg 1560cgccgaaatgc ccgggtcgccat tcattaa
agag gatcgaaatc tctggatca acggaaacaa 1620gatcaagcaa ttctgtctcaa aggaggttgt ttcc
tacaag aggtcaaca aggtctactt 1680cacc
gactcc attcccaaga acccttccgg caagatctta aggaaggact tgagagccag 1740gctcgccgct gg
catccccca ccgaagttgc cgcgcggaga agctaaaggcc cgcttctcg 1800gaacgcagtc acccatgggt
ctgttttaggt gctgtttagt accacacccaa atggggaaag 1860aaactacggg aggggatcat attattgtt
g caggagatc cagtttgcatttgcctg 1920cttgcgtat gttgataaaaa taaaatgata taataga
tgt gttgttttat tttttgacca 1980tgcgttataaa aggctgtttt atacactact tttttttaaaaa
aaaaaa aaaaaaaaaa 2038
<212> Type : DNA
<211> Length : 2038
SequenceName : 4CLcDNA (SEQ ID NO: 5)
SequenceDescription :

```

Custom Codon

Sequence Name : 4CLcDNA (SEQ ID NO: 5)

Sequence

<213> OrganismName : *Lolium perenne*

<400> PreSequenceString ::

MGSVPEESVV AVAPAETVFR SKLPDIEINN EQTLQSYCFE KMAEVASRPC IIDGQTGASY 60TYTEVDSL
R RAAAGLRRMG VKGDVVMNL LRNCPEFAFS FLGAARLGAA TTTANPFYTP 120HEIHRQAEAA GAKLIVT
EAC AVEKVLEFAA GRGPVVTVD GRDGCVDFA ELIAGEELPE 180ADEAGVLPDD VVALPYSSGT TGLPK
GVMLT HRSLVTSVAQ LVDGSNPNVF FNKDDALLCL 240LPLFHIYSLH TVLLAGLRVG AAIVIMRKFD VGA
LVDLVRA HRITIAFPFV PIVVEIAKSD 300RVGADDLASI RMVLSGAAPM GKDLQDAFMA KIPNAVLGQG Y
GMTEAGPVL AMCLAFAKEP 360FKVKSGSCGT VVRNAELKVV DPDTGASLGR NOPGEICVRG KOIMIGYLND

PESTKNTIDK 420DGWLHTGDIG LVDDDDEIFI VDRLKEIIKY KGFQVAPAEI EALLLTNPEV KDAAVGV
 KD 480DLCGEVPVAF IKRIEGSEIN ENEIKQFVSK EVVFYKRINK VYFTDSIPKN PSGKILRKDL 54
 ORARLAAGIPT EVAAPRS 557
 <212> Type : PRT
 <211> Length : 557
 SequenceName : 4CLpep (SEQ ID NO: 6)
 SequenceDescription :

Sequence

 <213> OrganismName : Lolium perenne
 <400> PreSequenceString :
 ggcacgagga atcctaccaa accgagctac cagatccttc tctactaatac gagctcccta 60cgctgctcc
 g cctgtcttcg tttccgcctc accgcccggcc gtttctccgc tccaagctac 120gtccgtccgt ccacata
 tat agcatcgaca tgaccatcgc cgaggtcgtg gtcgcccggag 180acaccgcccgc cgcgggtgggt cagcc
 ccccg ggaacgggca gaccgtgtgc gtgaccggcg 240ccgcccggta catcgctcg tggctcgta agc
 tgctgct ggagaagggg tacaccgtca 300agggccaccgt caggaaccca gacgacccga agaacgcgc a
 ctgaggggcg ctcgacggcg 360ccgcccggaccg gctggctcgc tgcaaggccg acctccctcga ctacgacgc
 atccggcgcg 420ccatcgacgg ctgcccacggc gtcttccaca cgcgtcccc cgtcaccggc gaccccg
 ga 480aaatggtgg a cccggcggtg agggggcaacgc agtacgtcat agacgcggcg gcgaggccg 54
 0gcacggcgcg gcgatggtg ctcacctcct ccacccggcgc cgtcaccatg gaccccaacc 600gcggggccg
 ga cgtggcgctc gacgagtcgt gctggagcga cctcgacttc tgcaagaaaa 660ccaggaactg gtactg
 ctac gggaaaggccg ttgcggagaca ggccgcacatcg gagttggcgc 720ggcagcgcgg cgtggacatt gtgg
 tggta acccggtgct ggtgatcgca cccctgctc 780agccgacggt gaacgcgcacg atcggccaca tc
 ctaagta cctggacggg tcggccagca 840ag
 ttcgcacaa cgcggcgtcag gcgtacgtgg acgtccgcga cgtggccgac gcccacctcc 900gcgtcttcga
 gtgcgcgcgcg cgcgtccggcc gccacctcgc cgcgcgcgcg gtcctccacc 960gcgaggacgt cgtgcgcac
 t ctcgcacaaacgc tcttcccgca gtaccccgctc cccaccagg 1020gctctgatga gacgaacccg aggaagc
 agc catacaagat gtcgaacccagg aagctccagg 1080acctcggaact ctagttcagg cccgtgagcc agtcc
 ctgta cgagacgggtg aagagccctc 1140aggagaaggg ccacccggcgt gtcgcacatcg agcaggcaga ggc
 ggacaag gaaaccctag 1200ctgcggagct gcaaggcagggttaccatcc gacatgagg aacaagaaat c
 aaccatgtc 1260cataactgcta ctgtcatgt aaccagctgt tgaatgccta aaatctaagt tcttgcata
 1320ctgtgttggt tcatgtggac tagattgatc gaataaacat ctctacacaa gttgtctaaa 1380a
 aaaaaaaaaaaaaa 1395
 <212> Type : DNA
 <211> Length : 1395
 SequenceName : CCR1cDNA (SEQ ID NO: 7)
 SequenceDescription :

Custom Codon

 Sequence Name : CCR1cDNA (SEQ ID NO: 7)

Sequence

 <213> OrganismName : Lolium perenne
 <400> PreSequenceString :
 MTIAEVVAAG DTAAAVVQPA GNGQTVCVTG AAGYIASLWV KLLLEKGYTV KGTVRNPDDP 60KNAHLRALD
 G AADRLVLCKA DLLDYDAIRR AIDGCHGVFH TASFVTDDEP QMVEPAVRGT 120QYVIDAAAEA GTVRRMV
 LTS SIGAVTMDPN RGPDVVVDES CWSLDLDFCKK TRNWYCYGKA 180VAEQAASELA RQRGVDLVVV NPVLV
 IGPLL QPTVNASIGH ILKYLDGSAS KFANAVQAYV 240DVRDVADAHL RVFECAAASG RHLCAERVLH RED
 VVRILAK LFPEYPVPTR CSDETNPRKQ 300PYKMSNQKLQ DLGLEFRPVS QSLYETVKSL QEKGHLPVLS E
 QAEADKETL AAELQAGVTI 360RA
 362
 <212> Type : PRT
 <211> Length : 362
 SequenceName : CCR1pep (SEQ ID NO: 8)
 SequenceDescription :

Sequence

 <213> OrganismName : Lolium perenne
 <400> PreSequenceString :
 ggcacgagca acaagtcatc aatggcgaa ggcttgcggc cgctcggtt ggctgcgagg 60gacgcctcc
 g gtcacacccctc cccttacacgc ttctcgagaa gcgttccgaa ggacgacat 120gtgacgatca aggtgtct
 ctt ctgcgggatc tgccacactg acctccacat catcaagaac 180gactggggca acgcctcta cccca
 tcgtc ccagggcatg agatcggtgg cgctcgcc 240agcgtcgca gccgcgttag cagcttcaag ggc
 ggcaca cggtggcggt gggctacttc 300ctcgactcct gccgcacccgt ctacagctgc agcaaggggt a

cgagaactt ctgccccacc 360ctgacgctca cctccaaacgg cgtcgacggc ggccggcgcca ccaccagggg
 cggcttctcc 420gacgtctcg tcgtcaacaa ggactacgtc atcccgctcc cggacaacct gcccctgg
 cc 480ggcgccac ctctccctcg cgccggcgac acagtctaca gccctatggt ggagtacggc 54
 0ctcaacgcac ccgggaagca cytcggcgac gtcggcctgg gcgggctgg ccacgtcgcc 600gtcaagtt
 cg gcaaggcctt cgggatgacc gtcacccgtca tcagctcctc ggacagagaag 660cgcgacgagg cgctcg
 gccg cctccggcgac gacgccttcc tcgtcagcag cgaccccgag 720cagatgaagg cggcggcgccc cacc
 atggac ggcacatcg acacgggtgc cgccggccac 780ccgatcggtc cgctgctcga cctgctcaag cc
 catggggc agatggtcgt ggtgggcg 840cc
 cagcaagc cgctcgagct cccggcccttc gccatcatcg gccgctcgcc 900ggggacggca
 cccggcagcg cgcacatgc caggccatgc tcgacttcgc gggcaagcac 960ggcattaccgg cgacgtcg
 a ggttcgtcaag atggactacg gtcacacccg ccatcgagcg 1020gctagagaag aacgacgtca ggtaccg
 ctt cgtcatcgac gtcggccggca gccacccgtca 1080gggcacccgccc gcttaacttg tgctacacaa tgtgg
 acgcg cgctcggtt gtcggaaaaaa 1140aggttcgccc gtcacagcc acatgaacaa gtcaatgagt cgt
 tgggtgt ttgttatct 1200tcattccaca tatgggacgc agttccagat tttcatgtca aataattgcg t
 cgtgtgcgg 1260ttgtcaagac tcaaatagga gaaaaaaaaga ctctgtgattt cggtttgcaaa aaaaaaaaaaa
 1320aaaaaa 1325

<212> Type : DNA
 <211> Length : 1325
 SequenceName : CAD1cDNA (SEQ ID NO: 9)
 SequenceDescription :

Custom Codon

Sequence Name : CAD1cDNA (SEQ ID NO: 9)

Sequence

<213> OrganismName : Lolium perenne
 <400> PreSequenceString :
 MAEGLPALGW AARDASGHLS PYSFSRSVPK DDDVTIKVLF CGICHTDLHI IKNDWGNALY 60PIVPGHEIV
 G VVASVGSVGS SFKAGDTVGV GYFLDSCRTY YSCSKGYENF CPTLTLTSNG 120VDGGGATTQG GFSDVLV
 VNK DYVIRVPDNL PLAGAAPLLC AGVTVYSPMV EYGLNAPGKH 180XGVVGLGGLG HVAVKFGKAF GMTVT
 VISSS DRKRDEALGR LGADAFLVSS DPEQMKAAG 240TMDGIIDTVS AGHPIVPLLD LLKPMGQMVV VGA
 PSKPLEL PAFAIIGGGK RLAGSGTGSV 300AHCQAMLDFA GKHGTTADVE VVKMDYQH RRAAREERRQ V
 PLRHRRRRQ PPAGHRRRLTC 360ATQCGRALVW SRKRFAGSQP HEQVNESLVC CLSSFHIWDA VPDFHVK
 407

<212> Type : PRT
 <211> Length : 407
 SequenceName : CAD1pep (SEQ ID NO: 10)
 SequenceDescription :

Sequence

<213> OrganismName : Lolium prenne
 <400> PreSequenceString :
 ggcacagtc gcctcaacg tctccctta accggccgtc cctacgcttg caccaccacc 60acgcacaga
 c agacgagttt cccagcccccc gccggaaaccg gatggcaccc acggccggccg 120agcagacggc gcaccac
 cag cacaccaggaa aggccgtggg gctggccggc cgcgcacgacg 180ccggccacct ctcccccgtc gcccat
 cacac ggaggagcac aggagacgac gatgtgggt 240taaaagatttt gtactgcggc atctgcact ctg
 acctgca cgcctgaag aacgactgg 300agaactcaag gtacccgatg atccccggc acgagatcgc c
 ggcgaggc acggagggtgg 360gcaagaacgt gagcaagttc aaggccggc accgcgtggg cgtcgggtgc
 atggtaact 420cgtggccgtc gtgcgagagc tgccgacaagg gtttcgagaa ccactgccc ggcgtat
 cc 480tcacccataa ctcggctgac gtcgacggca cccgtaccta cggccgtctc tccagcatgg 54
 0tgggtgggtca cgagccgggtc gtggccgggt tccccacgc catggccgtc gacaaggccg 600cgccgctg
 ct gtgcggccgc atcaccgtgt acagcccccataa gaaatggaccac gggctcaacg 660ttcccggtgc
 cggc gtgcggccgc tggccgggtt gggccacgtt gggtcaagt 720tcggcaaggc cttcgaaatgaa
 tggccggccgc tggccgggtt gggccacgtt gggtcaagt 780aggccctggg gccgctggc gccgacgcgt tc
 atccgtcgtat gacccatgg atggcatcat aaacacggta tctgcaaaca tccccctgac 900ccctcttcc
 gggctgtca agcccaacgg caagatgatc atggccggcc tccccgagaa 960gccccatcgag attccctcc
 t tcgtctctgt tgccacgaat aagaccctgg ccggagcat 1020catcgccggc atgagcgaca cgcaggaa
 gat gctggaccc tcgacggccac acggcgatgac 1080ggccgacatc gaggtggctcg gccggagatgatg
 acacg gccttggagc gccttggccaa 1140gaacgcacgc aggtatcgat tcgtcatcgacatcgacacc
 ctcgacatgttgcgc 1200caccaccggag tgaacgtact cagcactgt tacatctac gttgttccac t
 gttatgtct 1260ccgtatgtaaa caataaacga tcaaaaactct tggatctgg tgcattggtagacatgtt
 1320tggatgtatgg gaaactgagt tgaaggatgg atggataaaa aaaaaaaaaaaa aaaaaaaaaaa 1377

<212> Type : DNA
 <211> Length : 1377

SequenceName : CAD2cvEllettcdNA (SEQ ID NO: 11)
SequenceDescription :

Custom Codon

Sequence Name : CAD2cvEllett cDNA (SEQ ID NO: 11)

Sequence

```

<213> OrganismName : Lolium perenne
<400> PreSequenceString :
MAPTAAEQTE HHQHTRKAVG LAARDDAGHL SPLAITRRST GDDDVVIKIL YCGICHSDLH      60ALKNDWKNS
R YPMIPGHEIA GEVTEVGKNV SKFKAGDRVG VGCMVNSCRS CESCDKGFFEN      120HCPGMILTYN SVDVDGT
VTY GGYSSMVVVH ERFVVRFPDA MPLDKGAPLL CAGITVYSPM      180KYHGLNVPGL HLGVVLGGGL GHAV
KFGKA FGMKVTVISS SPGKKEEALG RLGADAFIVS      240KDADEMKAIV APWMASXNTV SANIPLTPLF GLL
KPNKGKMI MVGLPEKPIE IPPFALVATN      300KTLAGSIIGG MSDTQEMLDL AAKHGVVTADI EVVGAEVNT A
LERLAKNDV RYRFVIDIGN      360TLDNVAATTE
                                370
<212> Type : PRT
<211> Length : 370
      SequenceName : CADcvE11ettepep (SEQ ID NO: 12)
      SequenceDescription :

```

Sequence

<213> OrganismName : Lolium perenne
 <400> PreSequenceString :
 gcagcgggtn caaatcgccg gtcctgggtt ggaagtgnag cagtgggaag atgtgtgcga 60gggggttg
 t tttggatgna agacaggccg gccagtggag aacaagagag aacgcgagag 120gccaaagtat ccgcagc
 ccc gcaaacaagg cctagattt ggttaagttt gggtcgtctc 180agacaccgcg gccatccctt taggt
 ggtcc ggcgcgtgga ccgttattttt atctgagg 240accattcag acgcgcagac acgagatgga tgg
 tgcagtw agagatgacc taagtacaa 300aacctctcc cgagctggcc ccatcgtca cttaccgagc g
 acaaagttt cccacttcat 360cacactcagc ccagcaagca tactgatggt gagcgcactc gcggctgtgc
 ccaccgaccc 420cacgcattc aaaaccaact ctactttca ccmccaccaac aaaagacaaa atatgg
 ga 480ttttgtatg agatggaagc ggagcttgc agaatggaa acgcataaat cgagaacacg 54
 0tatacagtgc tggaaattgg atgactaagc cccaagggtt agaaaaaaa tnagaccatg 600tcttagat
 ga attagacatt ttttgatata atagaagcgg gacttggcgc gacaatttca 660aacttcgtcc ctaaca
 ggttca tgcactttc gatagttagc gtgtgctact gcgggacccc 720aaccacttgt gttaagccca catc
 gtttaa ggcaccaaggg ttagatgaaa gtaccaatct 780caactcatttgc gcaactagcta caaaacttgc tt
 ttcacatg tacggtcata ctacaattt 840ga
 ccttggta acgtaagtat gactgtatg gtgtgctaag gtgtgttggc agctcaaata 900aacccaaaaa
 tttcaacaca cgtcaacccat gaactgagat tcaaccaac ggctgagccg 960tctccctttaa aagatag
 g gagaaccaacca taatcaccat tgggtgtat gtgtgagttt 1020gcaagaaaa aaaaatggag aagccaa
 aac ccgttggagag agtgcgagag catacaagaa 1080caccacaaaca aagtgtgaag gaaaaaaaag atatg
 agata agatttcgga aataattttt 1140cacacccatg catgggtgtg gtgttcccg tcaccgtcta tgt
 atttctc gaaattcatg 1200cccacccatgg tagataaaaaa tattttttc tctctctct ttttattcaa a
 tctcaaagc 1260ataakrartg gtgacagaaac gataagattc ctacctagct ttctgagatc ccactagtt
 1320atcttcaagc tgggtgattga aggattaacc atgcttgaat tagattggct tcaaacttgg 1380t
 agtagctt tttcataactt tgattactt ggtatggta gttgggttga gatgggttgc 1440aatgtagaat
 cagattttag agcgattgtc agcttgaatt gccgcagtt tagcacatac 1500tagtttggat agatgaac
 ag tttggagaga caaataatgt ctatcgagc tcatcgata 1560atatttagtct atggcttttgc cttccgg
 tgtc ccctctgcaaa actttacccc tctgttagat 1620gttaggatttt ctgatatact ttcatggttt aagg
 gtgtgc gtgtaggaa cggagatc 1680cgga
 tcacac cttttcgctc acattttaca agcatgtaaac acctaagatt gattgatatac 1740taggcttaca cc
 ccaatggaa ggttaaactaa tattttgaa atgcgacttt tcaaaagtcc 1800caatataacc ttgacgatga
 tcttacaact actcgccca gtcttgcgt atatcgatt 1860ggccgaggat cgtgggtacc ttgttagtgg
 a ctagatgct catggaggtt gtatggacat 1920gtttaatgc tggtttctc tagttttt ctaatca
 act tggcatttctt ctccttaaca 1980cataataaga gggaaatacc tccatatactt ttctgaaaaa agcat
 ggcacca acaatgaaac 2040agaacaaagt acgcacagct atacccgacc caaacaatgg ctcaaggctt tca
 cgtgc 2100tagtttggta gcatgtattt tatagtagga actaaaattt aaagacaact tgcnaaaaca
 2160attttgc ttcgtgttt ttaaggatg cggcatttat cgattataca ttacatatgt 2220gat
 tggatgat gccaactttt tgcctccga tgatcatatg aaagggttgc atctttagggc 2280atctccaaatg g
 gnagactca aatgcaaaaa aatngtgcgt ttgggttcc cnngacaaaa 2340cctgctccca acggggcaac
 ccaactaaa aacggcagg tgcagcgtcc ggcntgacc 2400aaaactgacg caaatgg anatttt
 gg ggcacccatg acgaacgcgg ggcgtccact 2460tatccgacta tgcgtccatc ctggcccatc tgacag
 tgac acaaataca accacatgcg 2520cccccc
 accc ttctctctcc tccgtccgc ttttccatg gaancngtcc tcgctccctcg 2580ccggaaattga tctc
 gcctaa ccatgctccg cccacccct cgcctkaaqq ccccaaccc 2640cgctacactcc ttttggatcag cc

ctatttgg a gtcgccccg gttgaaacga gcgccgcag 2700cctcgacacc gccgagcaag acgaagactg
 ggcggagctc gccgagacgg gacggggacg 2760gagctcgcca tgcgtgcctc gcaggggcgc gatgggggg
 g gagctcccg tggctggctg 2820cagcacctcg gcccgtctg agccgtccca cgacgcgagc atgcgc
 tcg acgcccggc 2880gtgtcacctc gtcgcgcgc cagggccgc ccccccctgc cgaccggcgg cgg
 acgcg 2940accccgacg acgtgcccgg cggcagagac gcttcctcg cgacagcgcc ctccctcgatc
 3000tccgtcgagc cgacatacgcc gctaggaggg acgcgggcgt ccccggtgtc ggcctccgtt 3060gtggc
 gcatc gcggccggg cttccgtcga ggcgcacatcgc gggcgtggcc tcgtggcgca 3120gcctgcctg att
 cgtctg aggccggcgg cggagcttcc tgcggcgcgc gggggcggag 3180cctcctcgct gccggcgcac c
 tgcgtcgcc ggggtcgag acgcggcgcgc ggcagagctt 3240cctcgccgg gctcgccgc ggcctccctcg
 cggcgttgc gcttccaggc tcgcacgcgg 3300cctccggcgt ggcgcagcga gagcgcagcc tccggta
 gt taggcacagg cgcacacga 3360catcccc
 gc ctcggcctcc ggcgtggcgc agcgcgcg acgcgttagcc taggttggca 3420actagtacta cgagga
 agaa agaggagaaa caattattt ggtcacagcg ttggcgatcc 3480tgtgcgtatcc aaacggacac ccgg
 acgcga aacgatgtca gctgtccgc ttggcgacc 3540aaacgcacccg aaacggacgt ccgtttgggt cg
 gtgcgttg gagatgcct tactccccat 3600cctcaaatga gtctaattat atatcttggtaat 38
 aaaaagttaa actttgatca 3660acatttagtaa tgatagtagc aacgaataca aaattaaatt gtaaaaaata
 t attatgaaac 3720tttattttaa gatggatcta gttataactaa ttttctgcgg atggaggaag tagctaa
 ata 3780ttgttaattt ctaaataaaa aattaaaact ttaacttaaa acaaagttaa caagcataat 38
 40tatctgttgg a tggagaaatg agctaagata caccatctt ctctctacat taccttagcat 3900gccacat
 cag gaaacttattt aggataagctt ccaaggaaacc acccagaaca acaattttaca 3960tggcctggctt aaccc
 aatga caatttccga gcaacttggc gtgggtgtac gcttccttg 4020ttcaatttgc tcttattacaa gag
 tggccct gtataaggtaa aaaaaataaa caagcttcca 4080aggacggcca tggcccttgc tccgtcaggc t
 gcaacgtact cacgacgaag tttatctcg 4140gttctggaca tttgtctcgcc gcatatttgcgaccatgaaat
 taaaaatgtt 4200atatctgtat
 gggggatca tgcactcctt cgcagaggaa tccagacgac gatttacacg 4260tgtttccacc ttagcttt
 t ttaagtgtgt gtgttggaa cgtatcatata actgcccctg 4320aaatgtctgc atatataaac cgactcc
 atc atgtactcgaa gacaaggctc tcaaggaaaa 4380caaactatgc ctatctactt agcaatgtt tgaga
 gtaca gctttccgg tgccatattt 4440tttccatata atcttttctt gaagaacaag aaaaaaaaaa cag
 ttgggtgt ggtgggtgtt 4500gaaggcagaa accccatata aagccctgtt caccctcccc gcaaagcaca a
 ctcatagct 4560cgggtctctc gtcacacca aaatcgccca ccagcaccag catctctcgatcggcagacg
 4620catagatcgaa tgggtccac cggccggcgcg atggccgcgtt cggccggacga ggacgcgtgc 4680a
 tggtcgccc tccagctcgc ttctcgctg gtcctccga tgcgtgaa gaacgcac 4740gagcttggcc
 tcctggagat cctgggtggcc gccggccggca agtcgtgcac cccgaccgg 4800gtggccgcca agctcccg
 tc cggccgaac cggaaagcgc cggacatggt ggaccgcata 4860ctccggctgc tgcgtcgta caacgt
 cgtg acgtgcctgg tggaggaggg caaggacggc 4920cgcctctccc ggagctacgg cggccggccc gtgt
 gcaagt tcctcacccc caacgaggac 4980ggcgtctcca tggccggcgt cgcgtcatg aaccaggaca ag
 gtcctcat ggagagctgg 5040tgagttctc ag
 tggagctt gttactgtat atccgaattt gttcccttta gtgagggtta 5100attccggcgc cgctcgac
 5119

<212> Type : DNA

<211> Length : 5119

SequenceName : LpOmt1promoter (SEQ ID NO: 13)

SequenceDescription :

Custom Codon

Sequence Name : LpOmt1promoter (SEQ ID NO: 13)

Sequence

<213> OrganismName : Lolium perenne
 <400> PreSequenceString :
 tcccgatatct tcaacgtgc acccctacac ttccgtcttg tcttggagat ttacacac 60acggcaatt
 a ccaggatctt cttccatagat tattttttc gataaggatc ttccagat 120agcatgtgaa tctctgt
 act actactgtttt gtcaagcaaa attaacattt acatcagtgt 180ttttttgggg ggcagcggaa tcttt
 gacgc ctcttcttgc ctctcaagac atgtcaccct 240cacttagt tagtgcgtact ggttagtacta cgt
 acgatgc tccctccctc cgtaattttt 300caacctttt gctctcttctt ttataaaatg caaacctttt a
 aatctgacc agatatctgc 360taaaaaattt gcaacatgc atacatcaaa gcagtagtcc tccctccgtt
 taaaattacc 420tgggtttattt caaataaaatg caaactctgt aaaattcaat taaatatttta gaaaaatc
 ta 480acacgcacccg tagtataaaa gtatgcctcc tctgtttgttta aaaaagctaa gcaactttttt 54
 0tgagataggc ataaatctttt agctaaaaca tgcgttatata cctttgtatc tagataaaatg 600tggaaagc
 tt tttagaaac agacaaatg tgcgtttgttcaatttgaatgttggatattt 660ttccctctaat cttgtat
 caaa tttagaaat tttggcttgc atagaggac cattattttt 720atgaaactac ataaatttttaaaa
 cactca acataatttgc cgtgggtca gtgtatgc 780taacttagct tttcataat gcaactgtt tt
 caatagag catgaaggac gacaaatttgc 840tt
 cgtgtac ttgtatagag ggacgcgtt ctgggtcaac tcaccctgc tgcgtgttctt 900catcccttt
 gctcttccctc tctgtgttgc caatttgcgtt tccacgtgc atgtgggcgc 960aacttgcacc tagaaattt
 a catgctccca ctggccggag cggagttatctt tttgtgttgc tactaca

Custom Codon

— — — — —

Sequence Name : CAD2cvBar1anogenomic (SEQ ID NO: 14)

SequenceDescription ::

— — — — —

Sessions

<213> OrganismName : *Lolium perenne*
<400> PreSequenceString :

RSTGDDDVVI KILYCGICH S DLHALKNDWK NSRYPMPG H EIAGEVTEVG KNVSKFKAGD 60R VGVGCMVN
 S CRSCECSDKG FENHCPGMIL TYNSVDVDT VTYGGYSSMV VVHERFVRF 120PDAMPLDKGA PLLCAGI
 TVY SPMKYHGLNV PGLHLGVLGL GGLGHVAVKF GKA FGMKVT V 180ISSSPGKKEE ALGRLGADAF IVSKD
 ADEM K AVMSTMDGII NTVSANIPLT PLFGLLFALV 240ANKTLAGSII CGMSDTQEML DLA AKHGVTA DIE
 VVGA EYV NTALERLAKN DVRYRFVIDI 300GNTLDKVAAT TE
 312

<212> Type : PRT
 <211> Length : 312
 SequenceName : CAD2cvBarlanopep (SEQ ID NO: 15)
 SequenceDescription :

Sequence

<213> OrganismName : Lolium perenne

<400> PreSequenceString :

ggcacgagtc gcctccaacg tcttccctta accggccgtc cttacgcttg caccaccacc 60acgcacaga
 c agagcaggtt cccagcccccc gcccggaaacccg gatggcaccc acggccggcg 120agcagacgga gcaccac
 cag cacaccagga aggccgggtggg gctggccggcg cgcgcacgacg 180ccggccaccc ctcccccgtc gccat
 cacac ggaggaggcac aggagacacgac gatgtgggtga 240taaaagatttt gtactgcggta atctgcact ctg
 actgcga cgcctgaag aacgactgg 300agaactcaag gtaccgcgtatcccccggc acgagatcgc c
 ggcgagggtc acggagggtgg 360gcaagaacgtc gagcaagttc aaggccggcg accgcgtggg cgtcggtgtc
 atggtaact 420cgtccgggtc gtgcgagagc tgcgcacaagg gttcgagaa ccactgcccgg ggcgtatgt
 cc 480tcacccatcaa ctcggtcgac gtcgcacggca cggtcaccta cggccggctac tccagcatgg 54
 0tggtggtgca cgagccgggtc gtggccgggt tcccccacgc catggcgtc gacaaggccg 600cgccgctg
 ct gtgcgcggc atcaccgtgt acagcccat gaagtaccac gggctcaacg 660ttcccggtc gcaccc
 cggc gtgctggggc tggccgggtt gggccacgtt gccgtcaagt 720tcggcaaggc cttcggaaatg aaag
 tgcgg tgcgcacgc gtcgcggggg aagaaggagg 780aggccctggg gccgcgtggc gccgacgcgt tc
 atcgtcag caaggacgccc gacgagatga 840ag
 gctgtgat gaggccatg gatggcatc taaacacggg atctgcacaaac atcccccgt 900cccctctt
 cgggctgctc aagcccaacg gcaagatgat catggcggc ctcccccgg 960agcccatcgat gattccctcc
 c ttcgctcttag ttggccacgaa taagaccctg gccggggagca 1020tcatccggcg catgagcgc acgcagg
 aga tgctggaccc cgcggcgaag cacggcgtga 1080cggccgacat cgaggtggc ggcgcggagt atgt
 aacac ggccttggag cgccttgc 1140agaacgcgt caggtatcgc ttgcgtatcg acatcggcaa cac
 cctcgac aatgttgcgg 1200ccaccaccga gtgaacgtac tcagcactgc ttacgatcta cgttggccca c
 ttttagtgc 1260tccgtatgaa acaataaaacg atcaaaactc ttgtcatctg gtgcattgtt gttagacatgg
 1320ttgtttgcga ggaaacttagt ttgaaggatg gatggataaa aaaaaaaaaa aaaaaaaaaa 1378

<212> Type : DNA

<211> Length : 1378

SequenceName : CAD2cvBarlanocDNA (SEQ ID NO: 16)

SequenceDescription :

Custom Codon

Sequence Name : CAD2cvBarlanocDNA (SEQ ID NO: 16)

Sequence

<213> OrganismName : Lolium perenne

<400> PreSequenceString :

cgggatcaac ttggatgtcc ttgcggca cggttcagg aacaacgaca catgcacg 60ggatctcc
 c caaaagactca cacaagggtg acatgagcgc cccgtttttt gaagccaaatg 120tggctaaagaa atcgca
 agc ttggtggtt cggccaccc agatctgca acaaaaaggca 180ccaaggggagc tgccaaacaca tcaac
 cacaat ccatatgttc aaacgcacgc tcctcaagcc 240tcgaatgtc aaccggaaaga gaggcagaag ctt
 caaaaaa aacactcagcc aacccaaacg 300cctcgacgtc atcagagatggctctgag gacccgcagg g
 aaccaaccc tgcacaaac 360cgcatccggc agaaaaaggag caagacggc gcaaccctca agaggcacac
 gaaagacgtc 420gaagccaaaga ggagacgatg cgcaggagc gcccgcggc gagaaggggc cgttagac
 tc 480caagagctcg gcgtccctcg acctagcatc cgaagcactg accggggcac tcaatgcata 54
 0actttatctt gatggcatat gtactcaac ccatacaatg ttccatcatgc attatctatg 600gaacattc
 ct tcataataaa ctctgtgt gtcagtgcat aggaattttt attaacaacc 660aaaaacatac ttgggg
 ccttta cacacacttt cacagcatgg aaaactgtt agctttttaa 720agagttgcaaa aatctgtcaa gcga
 atgttc ttgtgataat tggacgaaat catgtttccc 780cattttcaat gtgtgtctct taccctaact ag
 caccgcac caacaaaatc tgaccatct 840ag
 ttatata tcatacgtac ccacatgttag gttgacccccc ataacacttg tttggatatc 900atggaaaatg
 gcttgcata acactttttt tcctacttttgg tacaatggt tatggactta 960ctcaattatgt gtttagag
 a gttttggctg cagatggat agcttcccaaa tattcatagg 1020ccctccggc gtggcagcc ccatcta
 cat aggctaaaaa ccagatttt gtaacatgtt 1080agacactttc aacttcatca tagaccatca aggag
 ctggc atgtgacagt gatataatgt 1140tcaatttaccc attcaacacg aatagttgc tcatgcattt gta
 gtcttgc ggcggcgggg 1200cgggaccatc gaacacacccg ccggcggcgtc agtaggttagataa a

atcttagccg 1260ttttcattca aacttgtat atataatcaa atttaataaa aaacctttat ttctgtgcata
1320ttttattttat ttgaggcgt gtttggggga cacggctgga aagtgacatc cccaaacact 1380g
cacgaagaa aacgcgtcgc caaaaaattc gatccggcgt cagtccttg ggagacgatt 1440tggatgacgc
ggctagagat gctctaagtt ctccacgcca tgtttcttc tatataata 1500cacagccaa ggtccatg
aa aagtaaaacg gcacgacgac accgaccggc gacaacttca 1560catatcggca catcgctatt acggac
caca tacaactcca ccgctattct cagccaagt 1620atacatgaca tgatccaatg gacgactttg tgag
cgaaac tagaaccttgcgggttttag 1680attt
tccaaat gtggataagt tgtacgcgcc gactagttt acacttggtt gaaaaaaagct 1740tattttagca cg
acttctca ctgacatagg aatgtaaaaca gtctctccac gccatgtttc 1800tttcttagtag tagcataacta
gttagtaactt ctctttgtcc tacacacacc cagggtccaa 1860gaaagggaaa cggcacgacg gcacccacc
g acgacgacga ctccacatca cggtcggta 1920aaaaaaagtc aaaaactcgctg acgtggcacc accggtc
gca gtcaactgac ggcgtcctct 1980gcgcaggtyt cacttcaagt ttacccatc actgtgggccc caccg
ccaaat gtggggcccg 2040cgagtttctt actcaactgac ctgtctccca ccagcccttcc cggcggtata tta
ccccggc 2100ccccaaatttc ctctgccttc ccacgagcag cagccggagc acggaaatccc ggccgcccatt
2160cctccaccc cagctccgc caaagattc catccggcga gatccatggg ctccatcgcc 2220gcgc
gacgcgc ctcccgccga gctgggtttc cggtccaagc tcccgacat cgagatcccg 2280acccacccctgac
gtgtcagga ctactgttcc cagggccctgc cggagctctc cgcgcgcgc 2340tgctctatcg acggcgccac
ggggccgcgc ctcaacctacg cggacgttggc cggccctacg 2400cgccgtcg cggcgggact cggccgccc
tg ggggtccgcga agggcgacgt cgtcatggcg 2460ctgctccgcga actgcccccg aatcccttc gtgttc
ctcg ggcggcccg gtcggcgcc 2520gccacc
acca cccgccaaccc gttctacacg ccccacgaga tccaccggca ggcgaccggc 2580gccggggccaa gggt
catgt caccgaggcc tgcggcgcc agaagggtcg cggccctcgcc 2640gccggagagag
2650

<212> Type : DNA

<211> Length : 2650

SequenceName : 4CL2promoterseq (SEQ ID NO: 17)
SequenceDescription :

Custom Codon

Sequence Name : 4CL2promoterseq (SEQ ID NO: 17)

Sequence

<213> OrganismName : *Lolium perenne*

<400> PreSequenceString ::

INTERNATIONAL SEARCH REPORT

International application No.
PCT/AU01/00699

A. CLASSIFICATION OF SUBJECT MATTER

Int. Cl. ⁷: A01H 5/00, C12N 9/00, C12N 9/02, C12N 9/04, C12N 9/10, C12N 15/29

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

GenBank, EMBL, PDB Nucleic Acids, GenPept, TREMBL, SWISS-PROT, PIR, Medline, ChemAbs, AGRICOLA, WPIDS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 5 952 486 A (BLOKSBERG <i>et al</i>) 14 September 1999 whole of document	1-26
X	Civardi L <i>et al</i> , "Molecular Cloning and Characterization of two cDNAs Encoding Enzymes Required for Secondary Cell Wall Biosynthesis in Maize", <i>NATO ASI Series, Volume H 104 (Cellular Integration of Signalling Pathways in Plant Development)</i> , 1998, pages 135-146 whole of document	1, 2, 4-12, 14-17, 20a, 19b, 23-26
X	WO 99/31243 A (INTERNATIONAL PAPER COMPANY) 24 June 1999 whole of document	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26

Further documents are listed in the continuation of Box C See patent family annex

* Special categories of cited documents:	
"A" document defining the general state of the art which is not considered to be of particular relevance	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"E" earlier application or patent but published on or after the international filing date	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"O" document referring to an oral disclosure, use, exhibition or other means	
"P" document published prior to the international filing date but later than the priority date claimed	"&" document member of the same patent family

Date of the actual completion of the international search

23 July 2001

Date of mailing of the international search report

29 August 2001

Name and mailing address of the ISA/AU

AUSTRALIAN PATENT OFFICE
PO BOX 200, WODEN ACT 2606, AUSTRALIA
E-mail address: pct@ipaaustralia.gov.au
Facsimile No. (02) 6285 3929

Authorised officer

GARETH COOK

Telephone No : (02) 6283 2541

INTERNATIONAL SEARCH REPORT

International application No.
PCT/AU01/00699

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	GenBank accession AF052223, Heath RL <i>et al</i> , "Lolium perenne 4-coumarate-CoA ligase 4CL3 mRNA, complete cds", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenPept accession AAF37734, Heath RL <i>et al</i> , "4-coumarate-CoA ligase 4CL3 [Lolium perenne]", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenBank accession AF052222, Heath RL <i>et al</i> , "Lolium perenne 4-coumarate-CoA ligase 4CL2 mRNA, complete cds", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenPept accession AAF37733, Heath RL <i>et al</i> , "4-coumarate-CoA ligase 4CL2 [Lolium perenne]", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenBank accession AF052221, Heath RL <i>et al</i> , "Lolium perenne 4-coumarate-CoA ligase 4CL1 mRNA, complete cds", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenPept accession AAF37732, Heath RL <i>et al</i> , "4-coumarate-CoA ligase 4CL1 [Lolium perenne]", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	WO 98/39454 A (ZENECA LIMITED) 11 September 1998 whole of document	1, 2, 4, 6-12, 14, 17, 20a, 19b, 22-26
X	Pichon M <i>et al</i> , "Cloning and characterization of two maize cDNAs encoding Cinnamoyl-CoA Reductase (CCR) and differential expression of the corresponding genes", <i>Plant Molecular Biology</i> , 1998, 38:671-676 whole of document	1, 2, 4, 6-12, 14, 17, 20a, 19b, 22-26
X	GenBank accession AJ231134, Selman-Housein G <i>et al</i> , "Saccharum officinarum mRNA for cinnamoyl-CoA reductase" 25 January 2000	1, 2, 4, 6-12, 14, 17, 20a, 19b, 22-26
X	WO 93/05159 A (IMPERIAL CHEMICAL INDUSTRIES PLC) 18 March 1993 whole of document	1, 2, 5-12, 15-17, 19b, 23-26
X	WO 93/24638 A (ZENECA LIMITED) 9 December 1993 whole of document	1, 2, 5-12, 15-17, 19b, 23-26
X	Baucher M <i>et al</i> , "Down-regulation of cinnamyl alcohol dehydrogenase in transgenic alfalfa (<i>Medicago sativa</i> L.) and the effect on lignin composition and digestibility", <i>Plant Molecular Biology</i> , 1999, 39:437-447 whole of document	1, 2, 5-12, 15-17, 19b, 23-26
X	GenBank accession AF010290, McAlister FM <i>et al</i> , "Lolium perenne cinnamyl alcohol dehydrogenase mRNA, complete cds", 23 September 1997	1, 2, 5-12, 15-17, 19b, 23-26
X	GenPept accession AAB70908, McAlister FM, "cinnamyl alcohol dehydrogenase [Lolium perenne]", 22 September 1997	1, 2, 5-12, 15-17, 19b, 23-26

INTERNATIONAL SEARCH REPORT

International application No.
PCT/AU01/00699

C (Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	Heath R <i>et al</i> , "cDNA Cloning and Differential Expression of Three Caffeic Acid O-Methyltransferase Homologues from Perennial Ryegrass (<i>Lolium perenne</i>)," <i>Journal of Plant Physiology</i> , 1998, 153:649-657 whole of document	17, 18, 19b, 20b, 23-26
X	GenBank accession AF033540, Heath RL <i>et al</i> , "Lolium perenne caffeic acid O-methyltransferase (OMT3) mRNA, complete cds", 29 January 1999	17, 18, 19b, 20b, 23-26
X	GenBank accession AF033539, Heath RL <i>et al</i> , "Lolium perenne caffeic acid O-methyltransferase (OMT2) mRNA, complete cds", 29 January 1999	17, 18, 19b, 20b, 23-26
X	GenBank accession AF033548, Heath RL <i>et al</i> , "Lolium perenne caffeic acid O-methyltransferase (OMT1) mRNA, complete cds", 29 January 1999	17, 18, 19b, 20b, 23-26
X	GenBank accession AF010291, McAlister FM <i>et al</i> , "Lolium perenne bispecific caffeic acid/hydroxyferulic acid O-methyltransferase mRNA, complete cds", 3 June 1998	17, 18, 19b, 20b, 23-26
X	Capellades M <i>et al</i> , "The maize caffeic acid O-methyltransferase gene promoter is active in transgenic tobacco and maize plants," <i>Plant Molecular Biology</i> , 1996, 31:307-322 whole of document	17, 18, 19b, 20b, 23-26
Note that the application as filed contains two claims numbered 19 and two claims numbered 20. To distinguish them, 19a and 20a refer to the first claims 19 and 20, and 19b and 20b refer to the second claims 19 and 20.		

Supplemental Box

(To be used when the space in any of Boxes I to VIII is not sufficient)

Continuation of Box No. II, Observations where unity of invention is lacking

The international search report has been drawn up in respect of the entire international application but the International Searching Authority is of the opinion that the application does not appear to comply with the requirements of unity of invention as set forth in the PCT regulations (Article 34(3), Rule 68(1) PCT).

The separate groups of invention are:

1. Claims 1, 2, 6 to 12, 16, 17, 19a, 19b and 23 to 26 (partial) and claims 3, 13 and 21 (complete) are to 4-coumarate-CoA ligase (4CL) from ryegrass (*Lolium*), the nucleotide sequence encoding it, the promoter from its gene and various uses of them. 4CL from ryegrass is considered to be the first "special technical feature".
2. Claims 1, 6 to 11, 16, 17, 19a, 19b and 23 to 26 (partial) are to 4CL from fescue (*Festuca*), the nucleotide sequence encoding it, the promoter from its gene and various uses of them. 4CL from fescue is considered to be the second "special technical feature".
3. Claims 1, 2, 6 to 12, 16, 17, 20a, 19b and 23 to 26 (partial) and claims 4, 14 and 22 (complete) are to cinnamoyl-CoA reductase (CCR) from ryegrass, the nucleotide sequence encoding it, the promoter from its gene and various uses of them. CCR from ryegrass is considered to be the third "special technical feature".
4. Claims 1, 6 to 11, 16, 17, 19b, 20a and 23 to 26 (partial) are to CCR from fescue, the nucleotide sequence encoding it, the promoter from its gene and various uses of them. CCR from fescue is considered to be the fourth "special technical feature".
5. Claims 1, 2, 6 to 12, 16, 17, 19b and 23 to 26 (partial) and claims 5 and 15 (complete) are to cinnamyl alcohol dehydrogenase (CAD) from ryegrass, the nucleotide sequence encoding it, the promoter from its gene and various uses of them. CAD from ryegrass is considered to be the fifth "special technical feature".
6. Claims 1, 6 to 11, 16, 17, 19b, and 23 to 26 (partial) are to CAD from fescue, the nucleotide sequence encoding it, the promoter from its gene and various uses of them. CAD from fescue is considered to be the sixth "special technical feature".
7. Claims 17, 18, 19b and 23 to 26 (partial) and claim 20b (complete) are to caffeic acid O-methyltransferase (OMT) gene promoter from ryegrass and various uses of it. Caffeic acid OMT from ryegrass is considered to be the seventh "special technical feature".
8. Claims 17, 18, 19b and 23 to 26 (partial) are to caffeic acid O-methyltransferase (OMT) gene promoter from fescue and various uses of it. Caffeic acid OMT from fescue is considered to be the eighth "special technical feature".

In order for there to be unity between the four types of enzymes claimed, they have to share a significant structural element, that is a structural element that defines the specific biological activity of the enzymes, and the significant structural element must be disclosed in the specification. No significant structural element has been identified as being shared by the four types of enzymes, hence there is lack of unity between the enzymes. In addition, all four types of enzymes are known in the prior art, for example, in US 5 952 486. Hence unity of invention is also lacking between the enzymes from ryegrass and the enzymes from fescue.

Note that the application as filed contains two claims numbered 19 and two claims numbered 20. To distinguish them, 19a and 20a refer to the first claims 19 and 20, and 19b and 20b refer to the second claims 19 and 20.

INTERNATIONAL SEARCH REPORT
Information on patent family-members

International application No.
PCT/AU01/00699

This Annex lists the known "A" publication level patent family members relating to the patent documents cited in the above-mentioned international search report. The Australian Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent Document Cited in Search Report				Patent Family Member			
US	5 952 486	AU	44036/97	BR	9711745	EP	929 682
		US	5 850 020	WO	98/11205	ZA	9710451
		ZA	9810574	US	6 204 434		
WO	99/31243	US	6 252 135	ZA	9811568		
WO	98/39454	AU	63041/98	EP	970 222		
WO	93/05159	AU	16581/92	BR	9205934	CA	2 109 222
		EP	584 117	US	5 451 514	US	6 066 780
WO	93/24638	AU	43345/93	BR	9306445	EP	643 774
		US	5 633 439				
END OF ANNEX							